

# Neutron diffraction for deciphering protein-carbohydrate interactions in bacterial infection

Lukáš Gajdoš

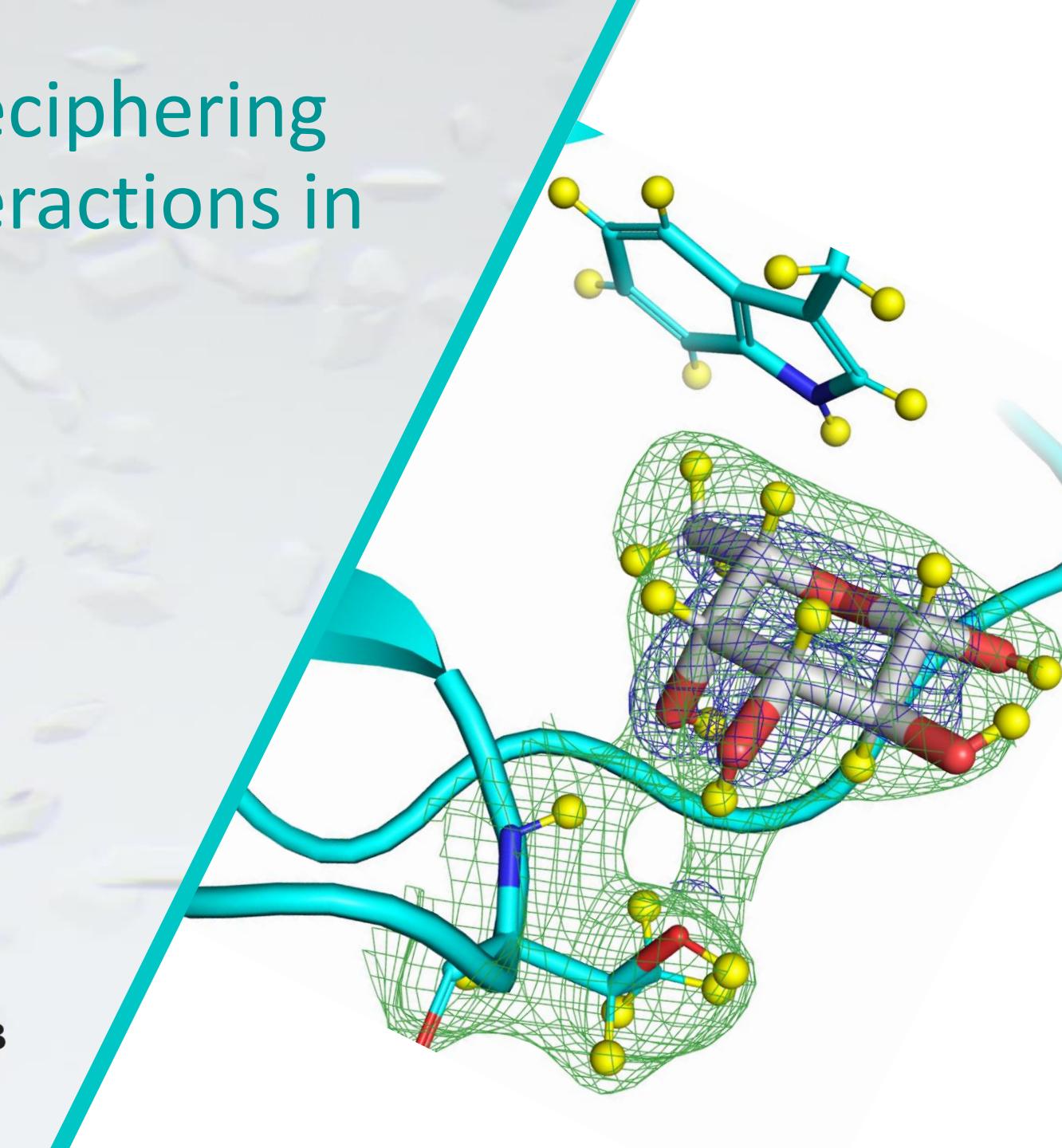
ESS/ILL User Meeting

Anne Imberty (CERMAV)

Juliette Devos (ILL)

Trevor Forsyth (ILL, LINXS)

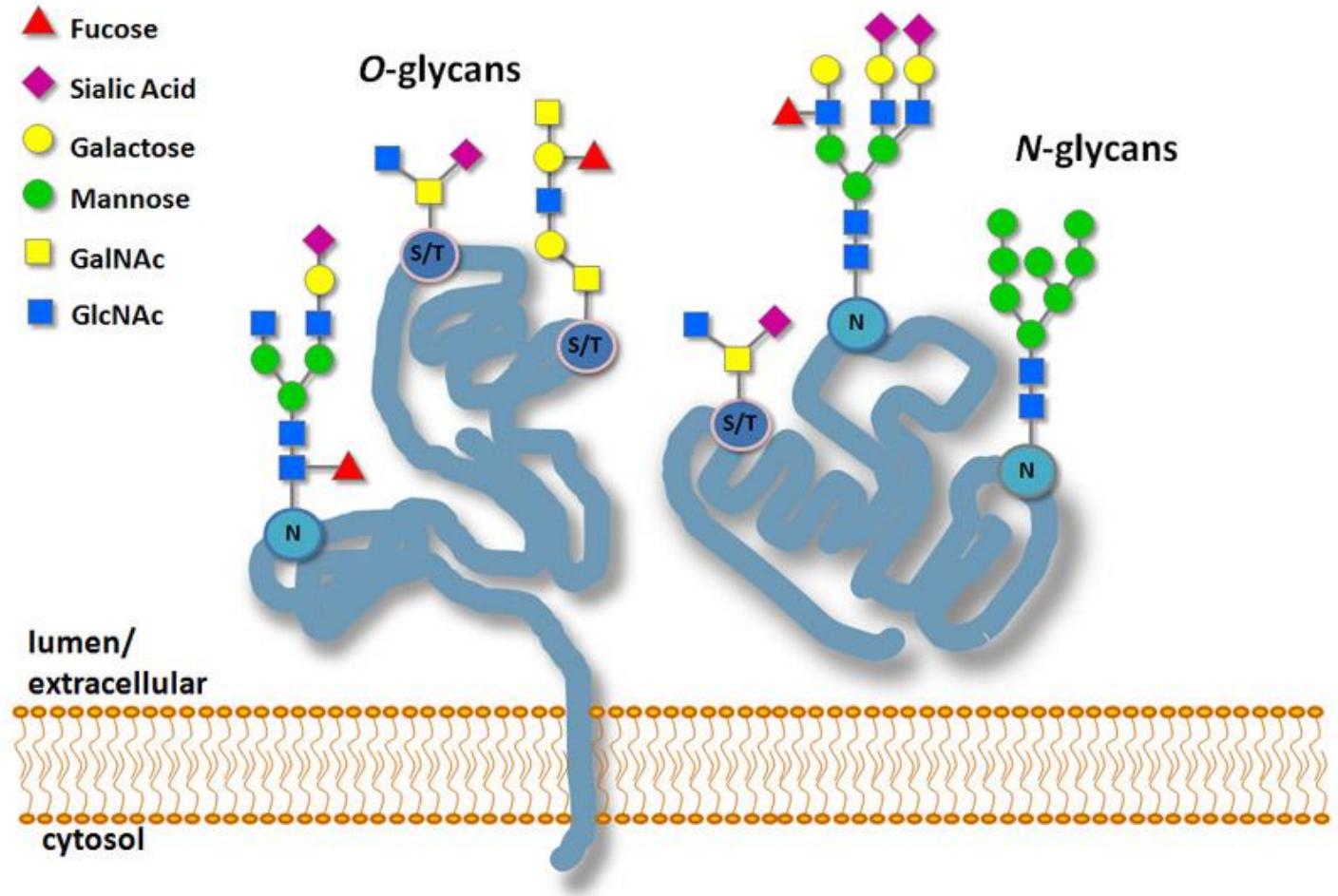
Matthew Blakeley (ILL)



# Glycans

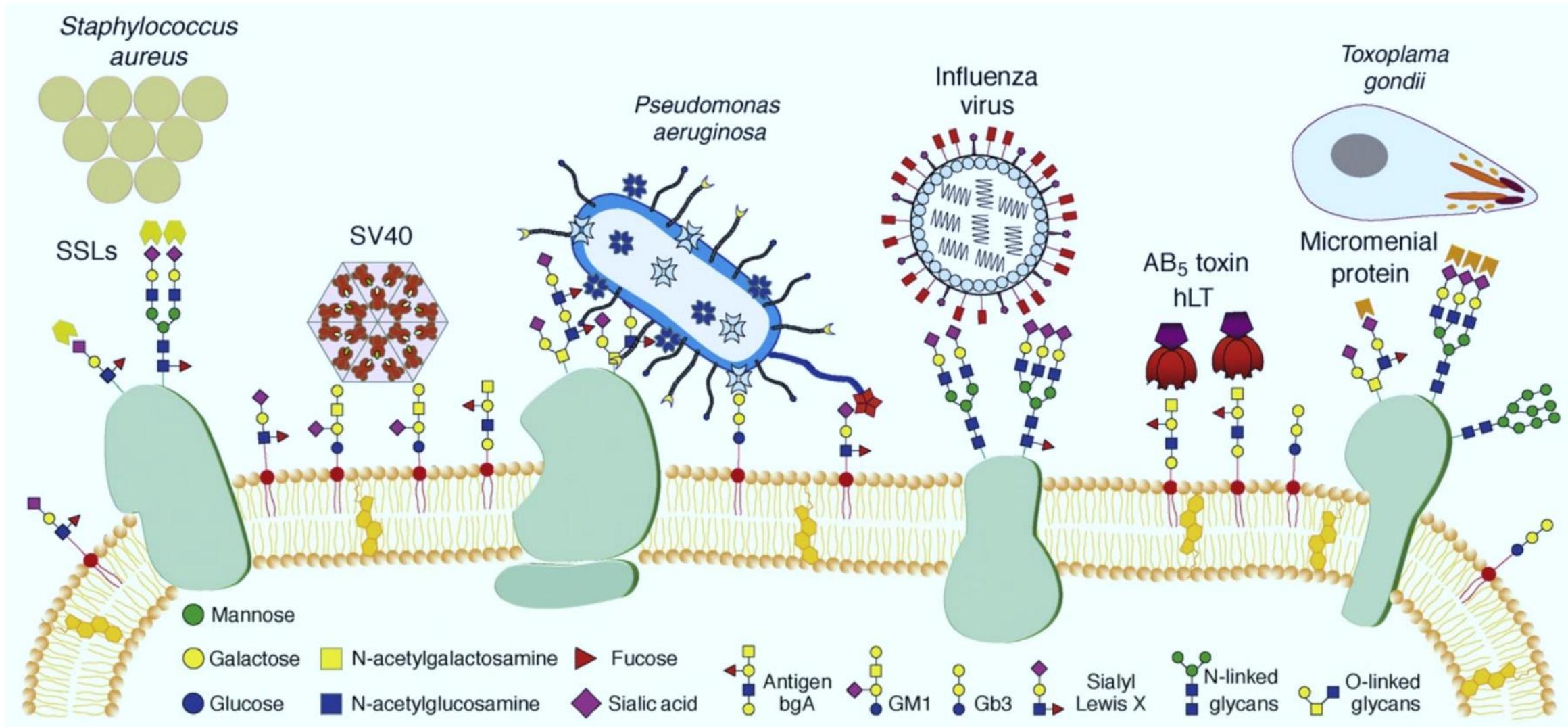


Nieuworp M., et al. 2005



Lectins are proteins that can decode this complex „glycocode“

# Lectins from pathogenic organisms



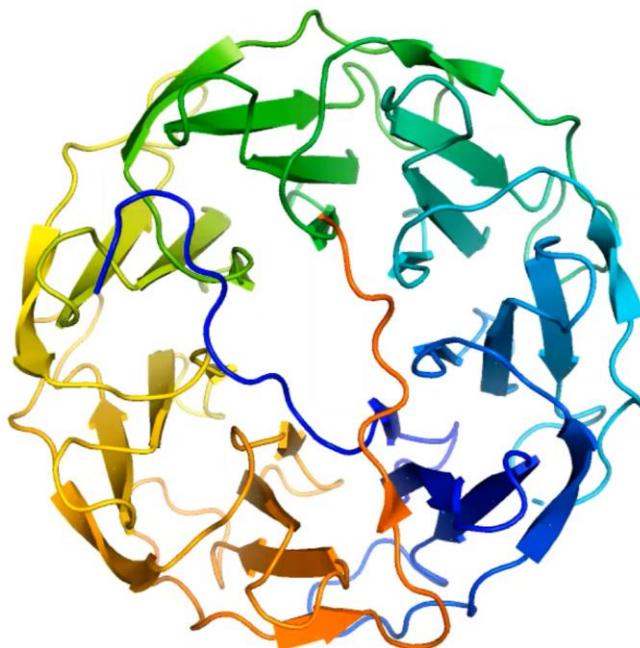
# *Photorhabdus luminescens & Pseudomonas aeruginosa*

# *Photorhabdus luminescens & Pseudomonas aeruginosa*

- Bioluminescent bacteria
- Insect pathogen
- Symbiosis with nematodes
- Complex life cycle
- Production of **toxins**
- Produces several **lectins**



Courtesy of Dr. A. Regaiolo  
and Prof. R. Heermann

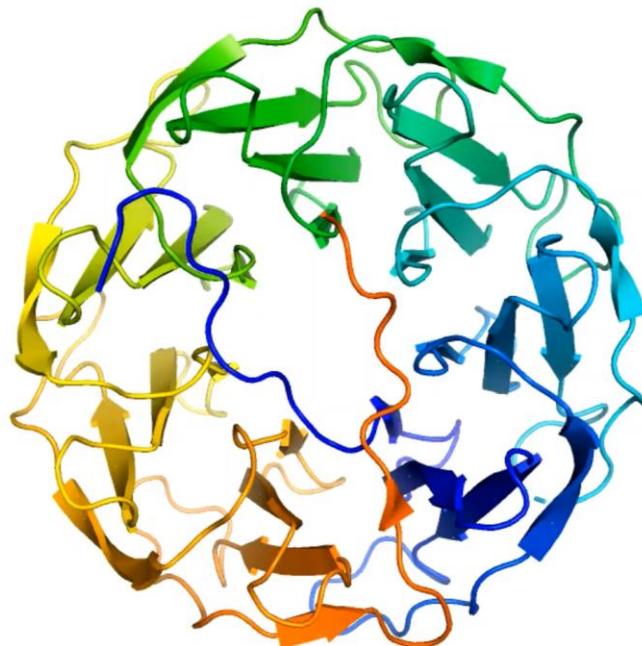


**PLL lectin**  
Fucose-specific  
 $K_D = 5 \text{ mM}$   
PDB: 6C9P

Kumar A. et al., 2016

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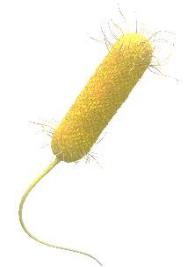
**PLL lectin**  
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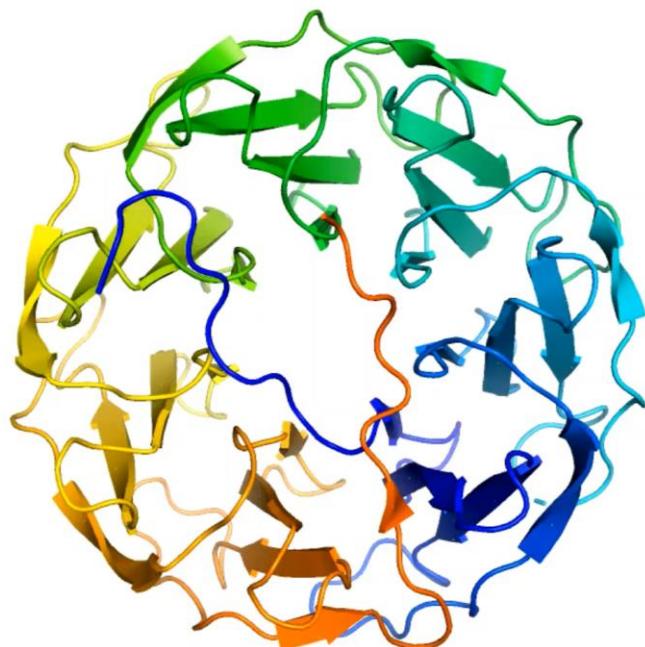
Courtesy of Dr. A. Regaiolo  
and Prof. R. Heermann

- Human **opportunistic** pathogen
- Lung infections (**cystic fibrosis**)
- **LecA** (PA-IL), **galactose**-specific
- **LecB** (PA-ILL), **fucose**-specific
- Roles in attachment and biofilm formation
- Targets for novel glycomimetics



# *Photorhabdus luminescens & Pseudomonas aeruginosa*

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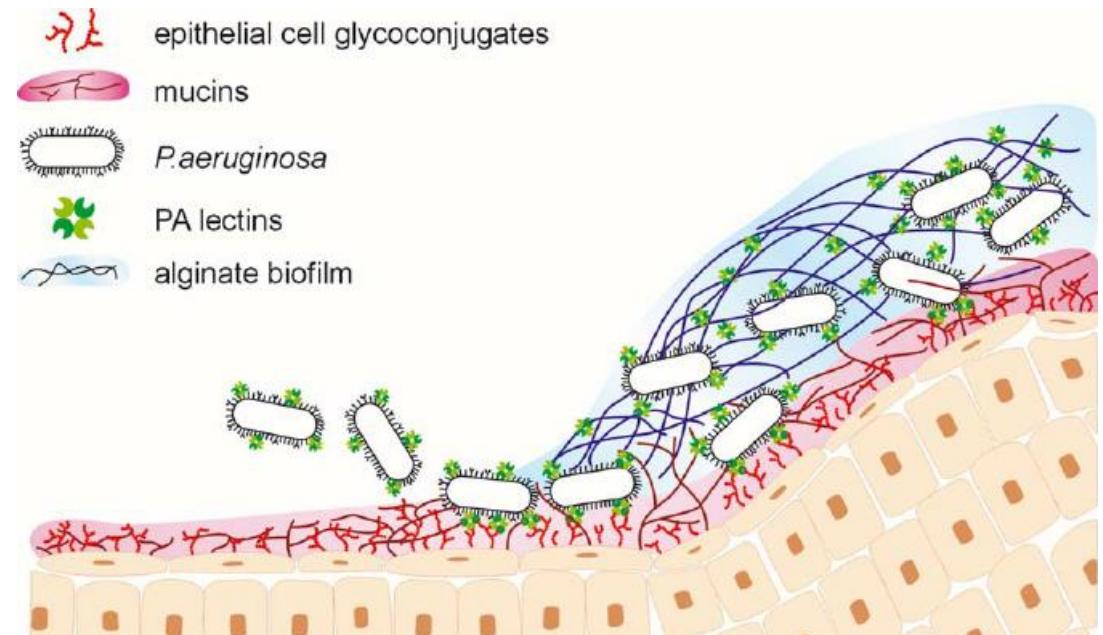


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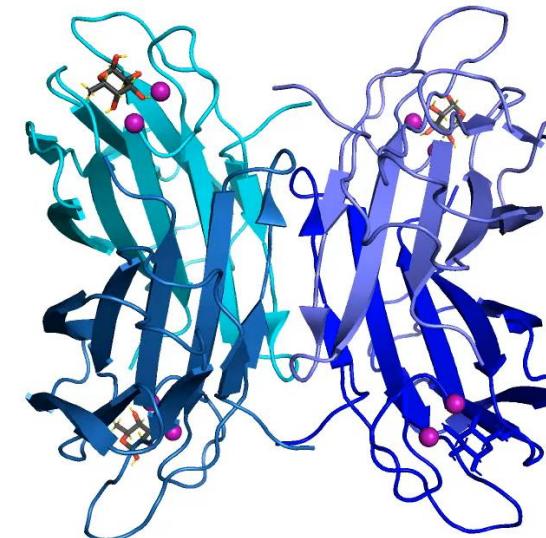
Kumar A. et al., 2016



Courtesy of Dr. A. Regaiolo  
and Prof. R. Heermann



Imberty A., et al., 2014



**LecB lectin**  
Fucose-specific  
 $K_D = 4 \mu\text{M}$   
PDB: 1GZT

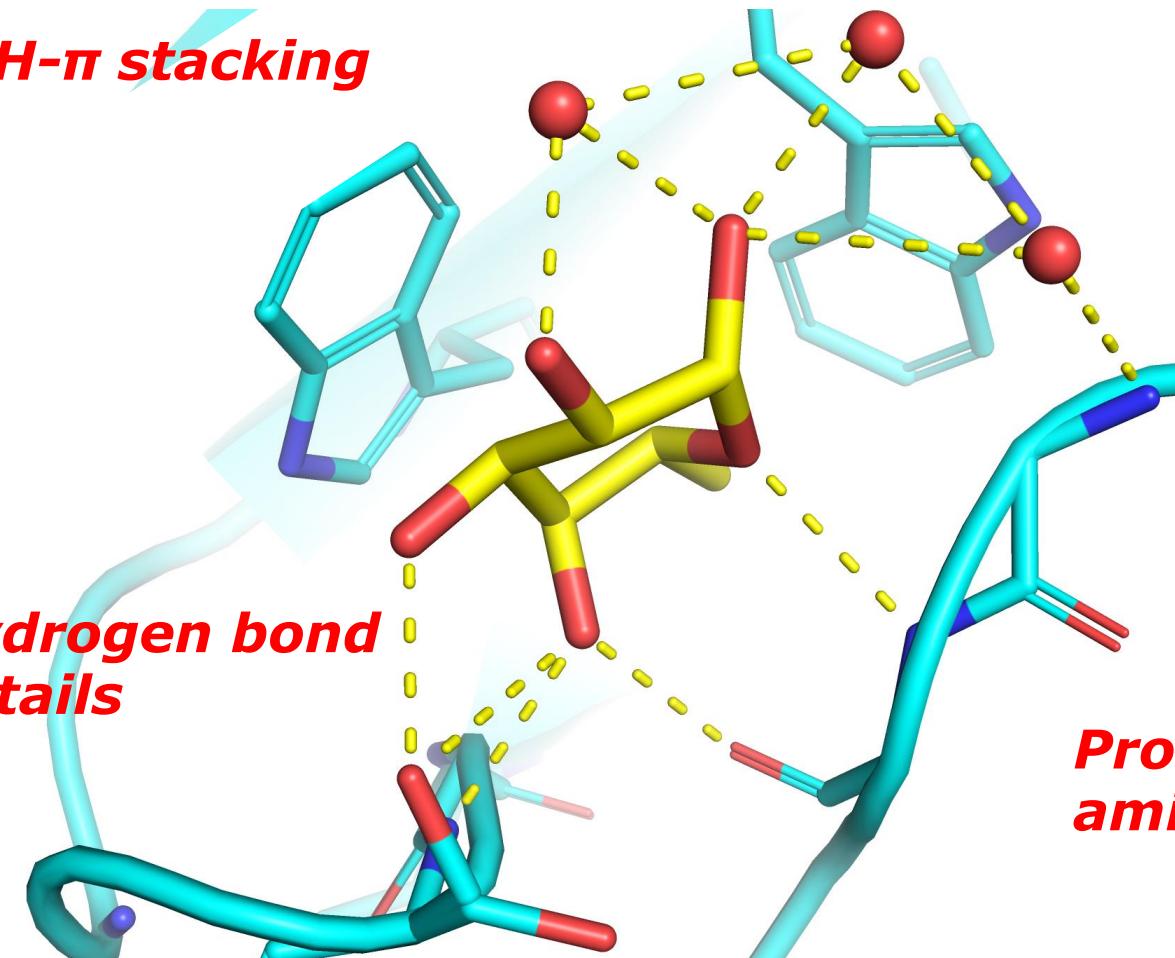
Mithell E. et al., 2002

# Protein-carbohydrate interactions

## *Fucose in PLL*

*CH- $\pi$  stacking*

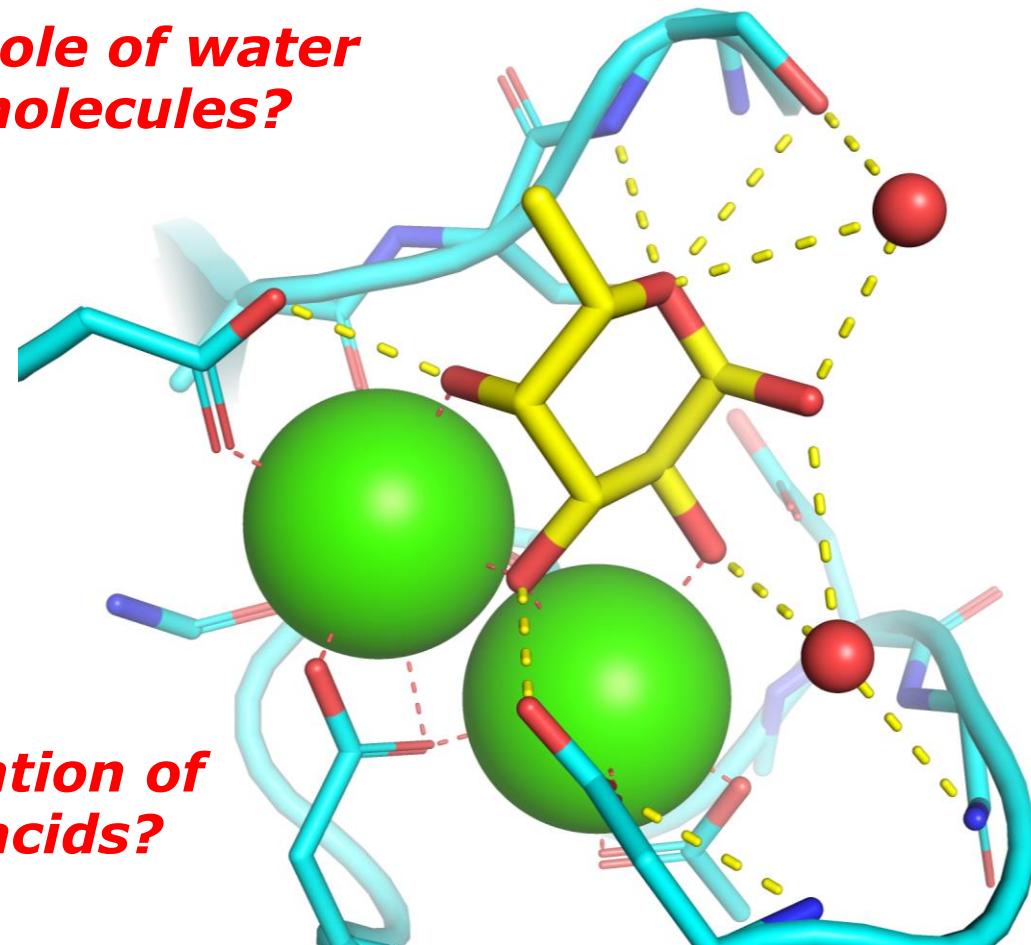
*Hydrogen bond details*



## *Fucose in LecB*

*Role of water molecules?*

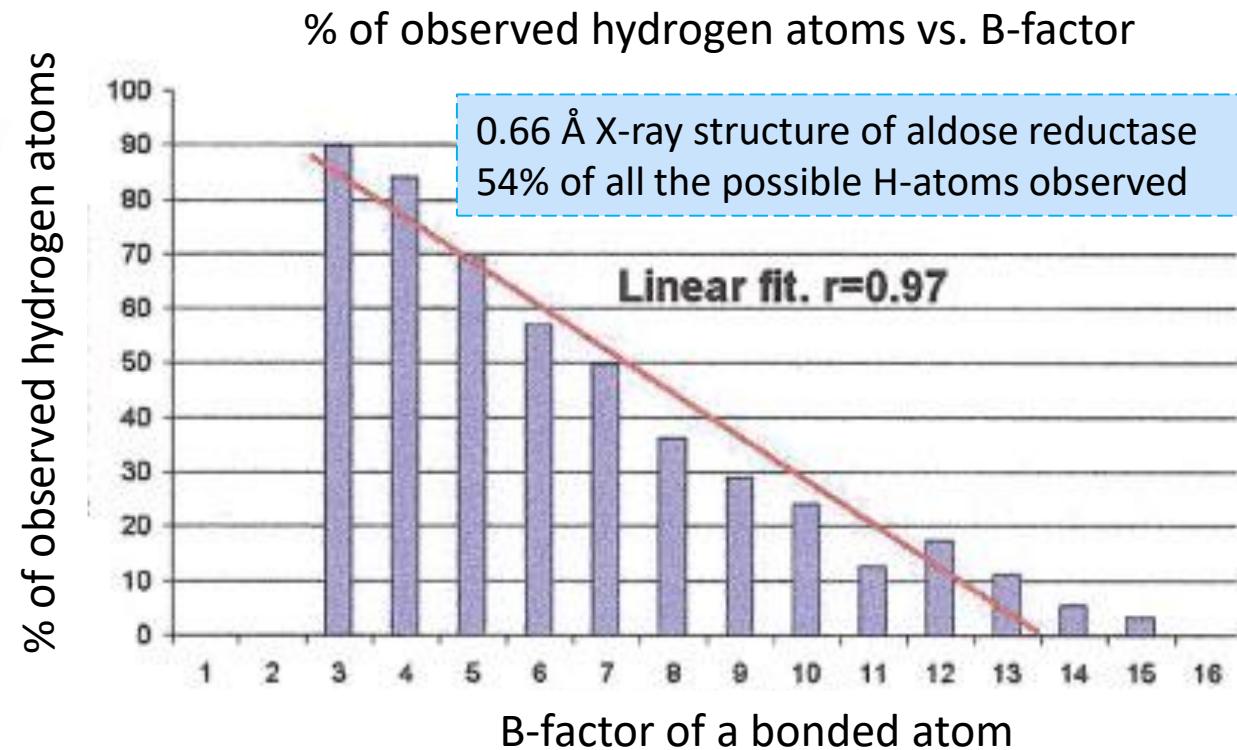
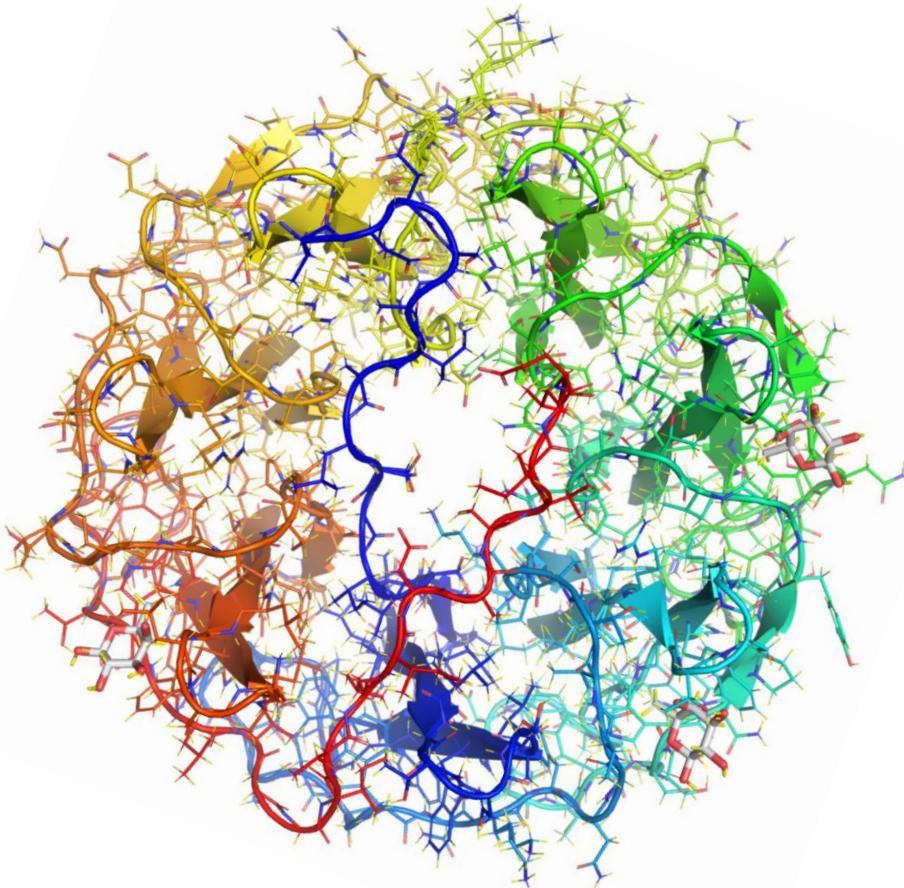
*Protonation of amino acids?*



**Wanted : Location of hydrogen atoms**

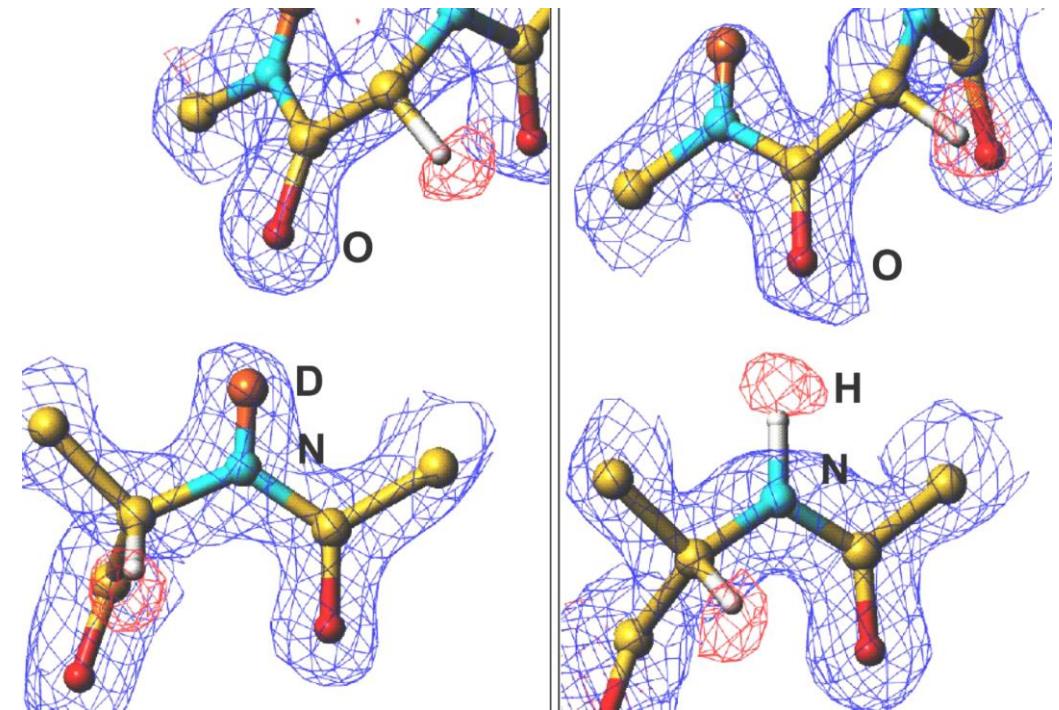
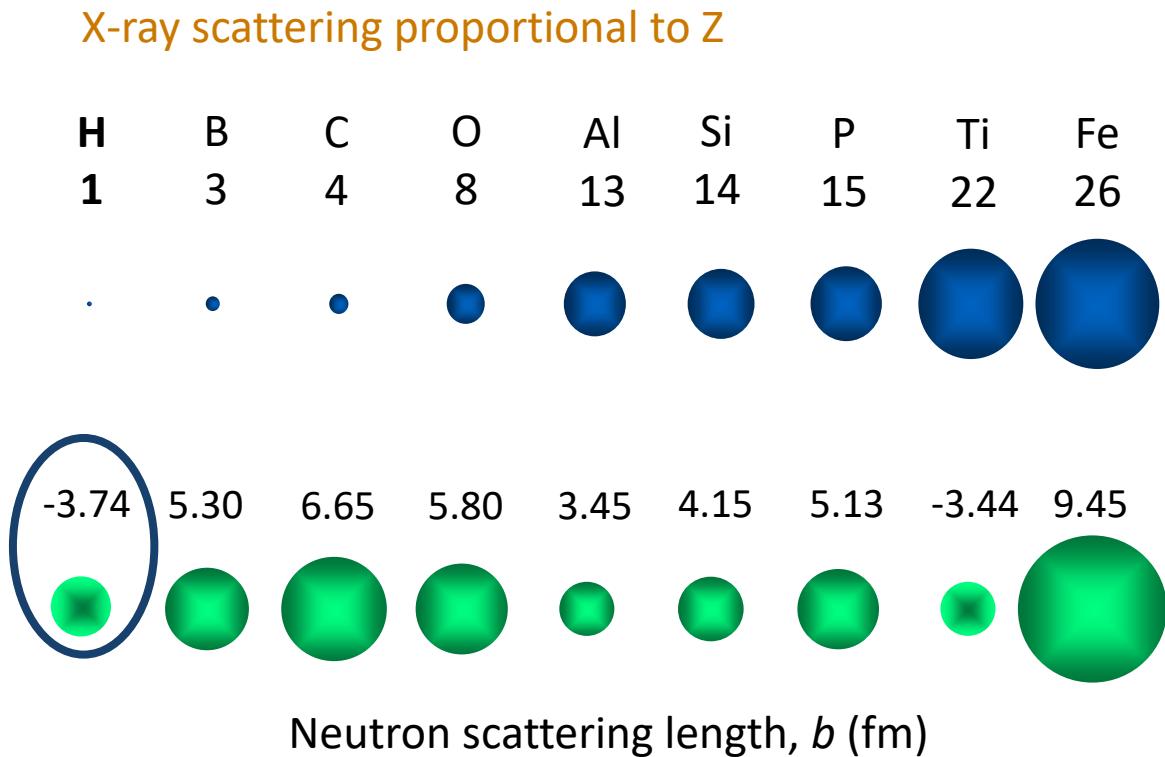
# H atoms „invisible“ in X-ray structures

- **Hydrogen atoms** account for about **half** of all the atoms in proteins
- Critical roles in **biological functions** (enzyme mechanisms, ligand binding)
- **Rarely observable** in X-ray diffraction experiments



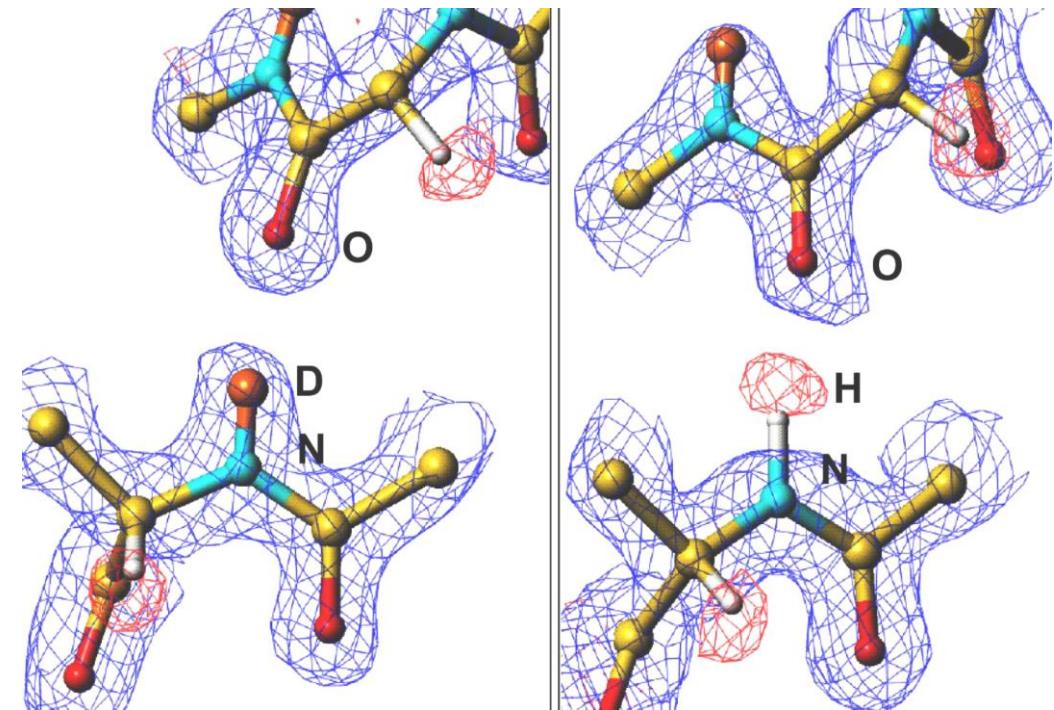
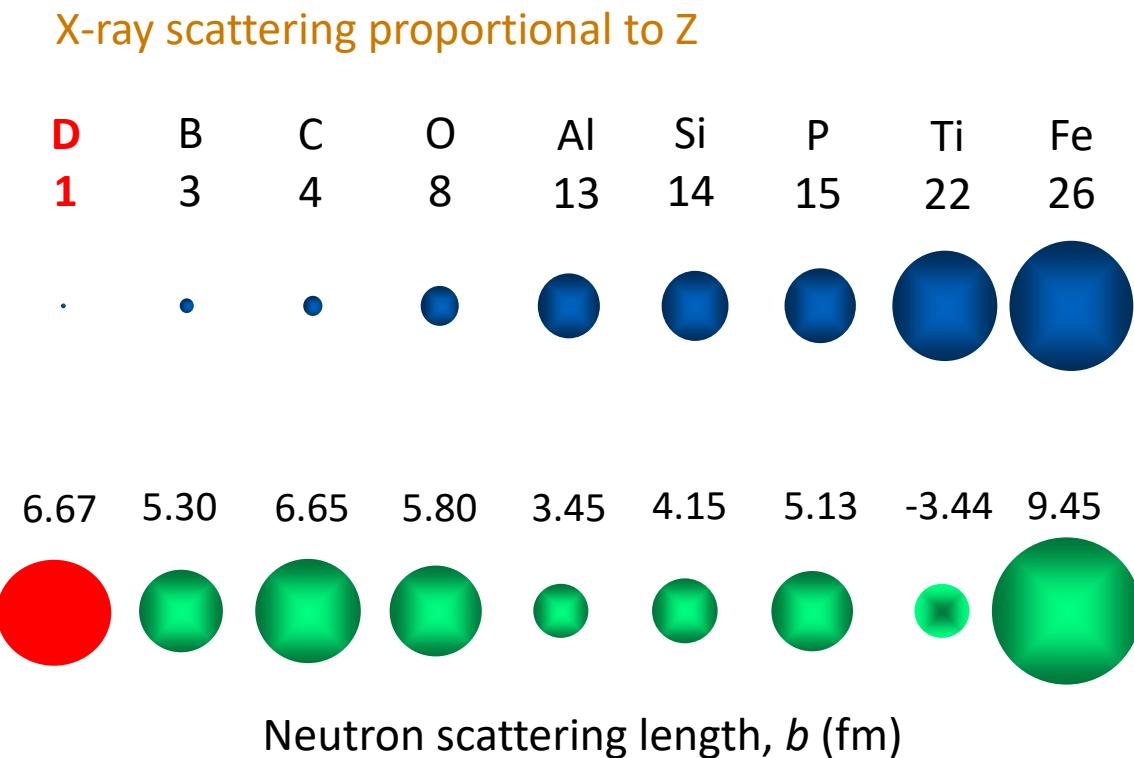
# Neutrons as a diffraction probe

- Interaction with atomic **nuclei**
- Scattering **varies** with elements and even **isotopes** of the same element (**H/D**)
- Non-destructive probe (room-temperature data collection)



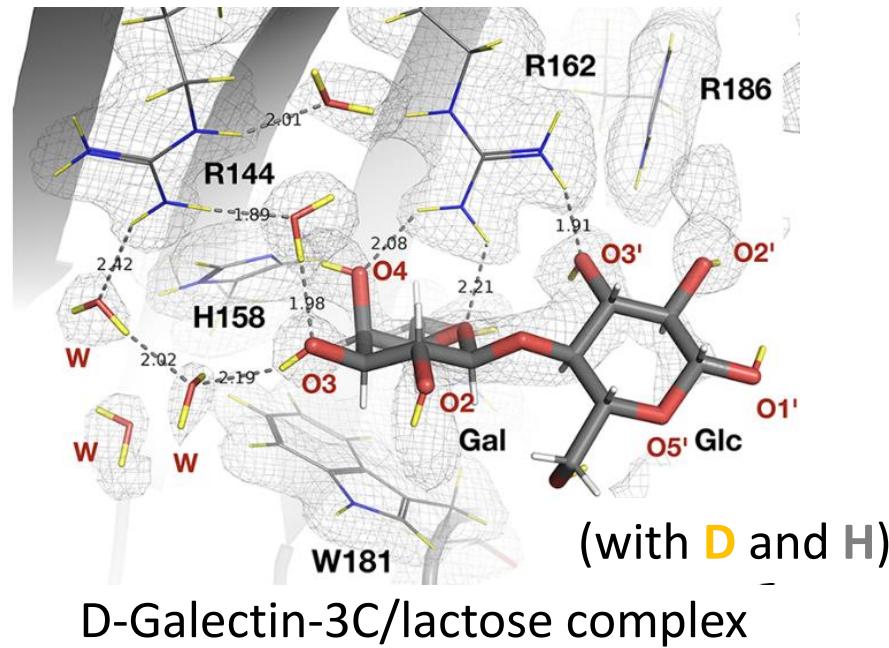
# Neutrons as a diffraction probe

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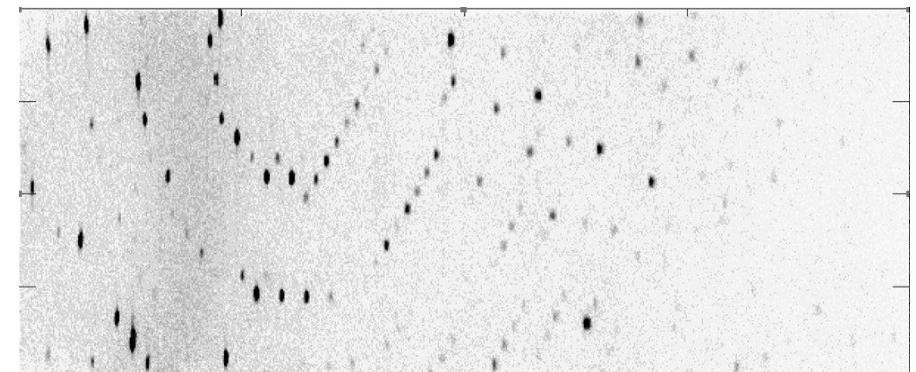
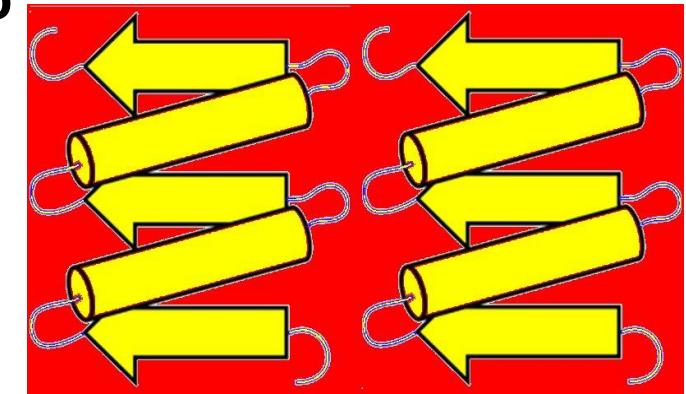


# Need of perdeuteration

- Full replacement of all **hydrogen** (H) atoms by **deuterium** (D) atoms
- Reduces the large **incoherent** scattering of H (~ 40 times larger than for D)
- **Reduces the background and increases the signal-noise ratio**
- Clearer visualization of neutron maps
- **Cancellation effects** limit visualization of CH<sub>n</sub> groups



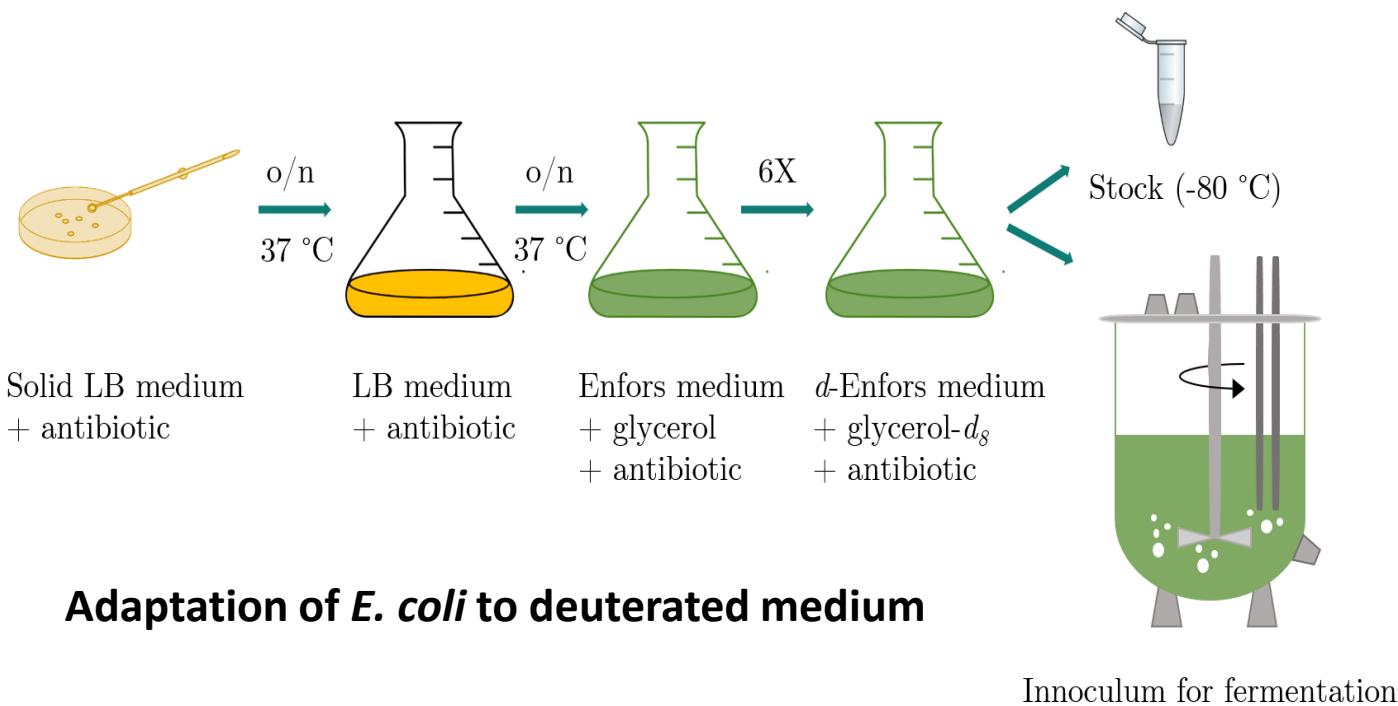
Manzoni F. et al., J. Med. Chem., 2018



Courtesy of Prof. Trevor Forsyth

# How to obtain perdeuterated biomolecules?

- **Adaptation** of *E.coli* cells to deuterated medium
- **Production** of recombinant proteins **in D<sub>2</sub>O** --- > D-Lab at ILL
- **Fermentation** (high cell-density cultures) of *E.coli*
- **Deuterated carbon source** (glycerol-d<sub>8</sub>, glucose-d<sub>12</sub>)



# How to obtain perdeuterated biomolecules?

- Proteins:

Production of recombinant protein in D<sub>2</sub>O ---> D-Lab at ILL

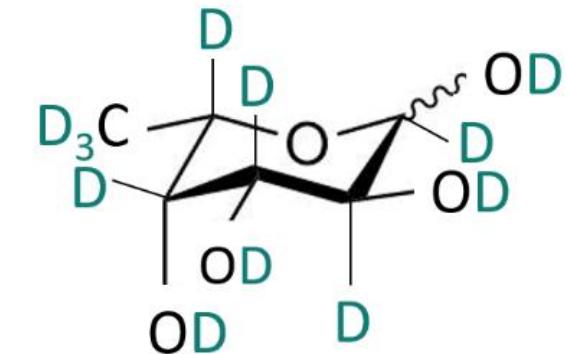


- Sugars:

Glucose-d<sub>12</sub> : grow plant/algae in D<sub>2</sub>O and degrade cellulose

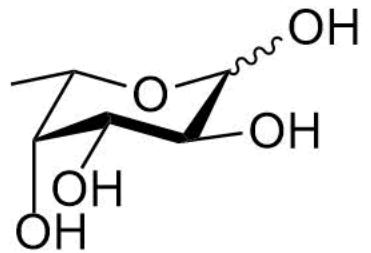
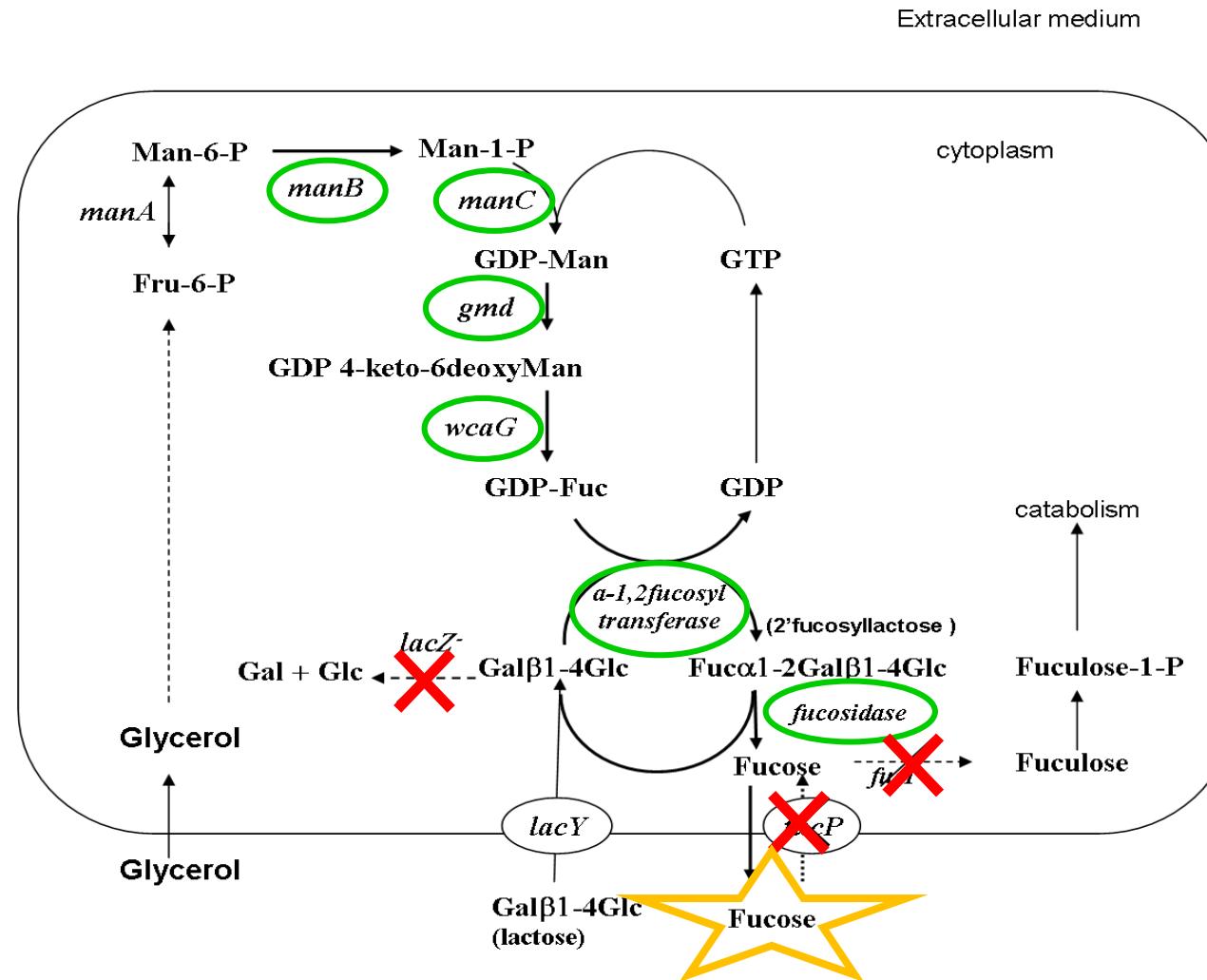
***Synthetic chemistry « isotopic hydrogen-exchange technique »***

***Synthetic (glyco)biology***



# *In vivo* production of L-fucose-d<sub>12</sub> in *E. coli*

Fucose-producing strain of *E. coli* designed and engineered by Dr. Eric Samain at CERMAV



Overexpressed genes

**manB**: phosphomannomutase  
**manC**: Man-1-P-guanyltransferase  
**gmd**: GDP-Man 4,6-dehydratase  
**wcaG**: GDP-L-fucose synthase  
 **$\alpha$ -1,2-fucosidase**  
 **$\alpha$ -1,2-fucosyltransferase**

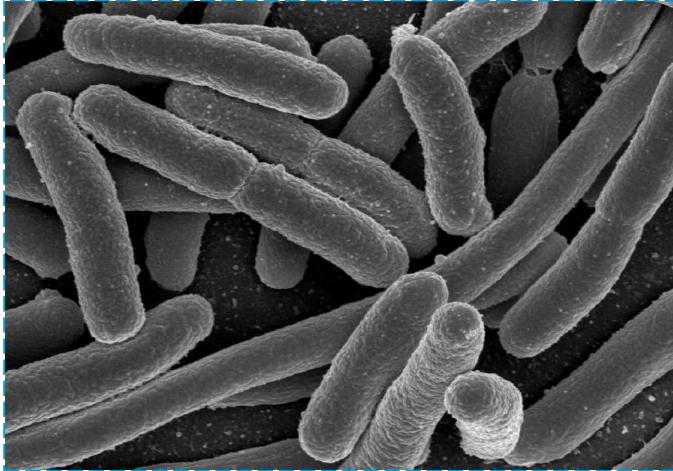


Knocked-out genes

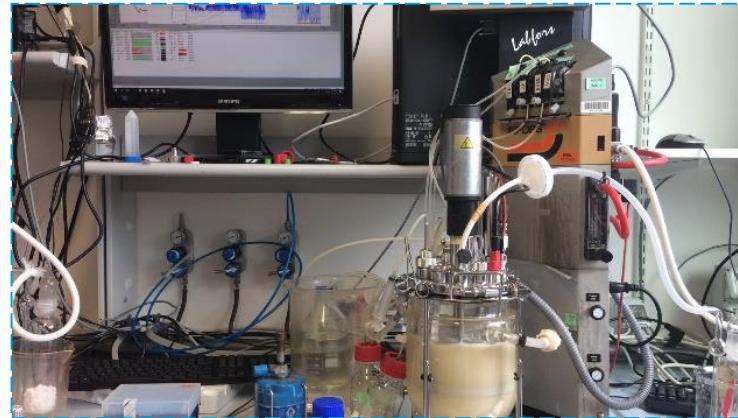
**lacZ**:  $\beta$ -galactosidase  
**fucI**: fucose isomerase  
**fucP**: fucose permease

# Production, purification and characterization of L-fucose-d<sub>12</sub>

Adaptation to D<sub>2</sub>O and glycerol-d<sub>8</sub>



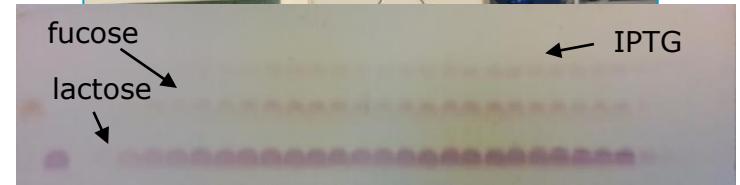
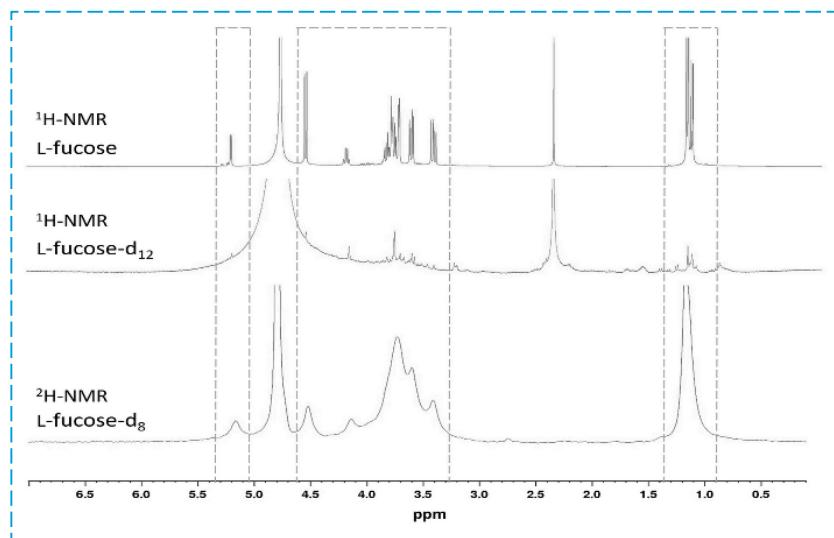
Batch production



Purification



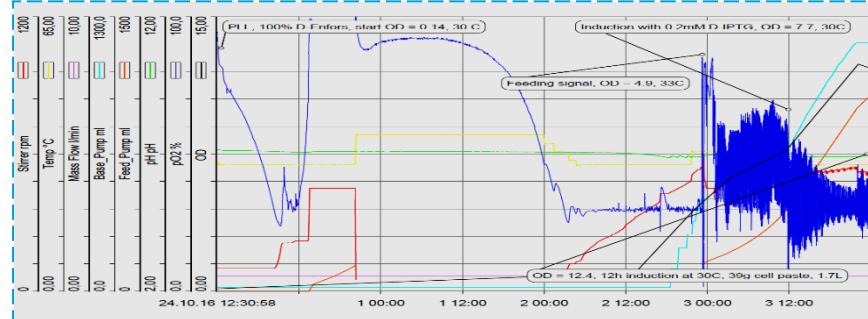
Characterization (Mass spec, NMR)



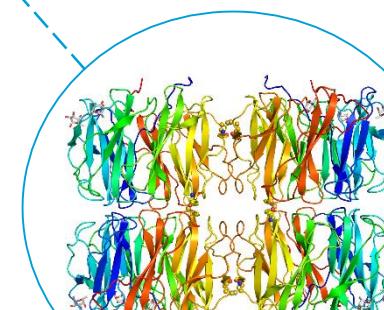
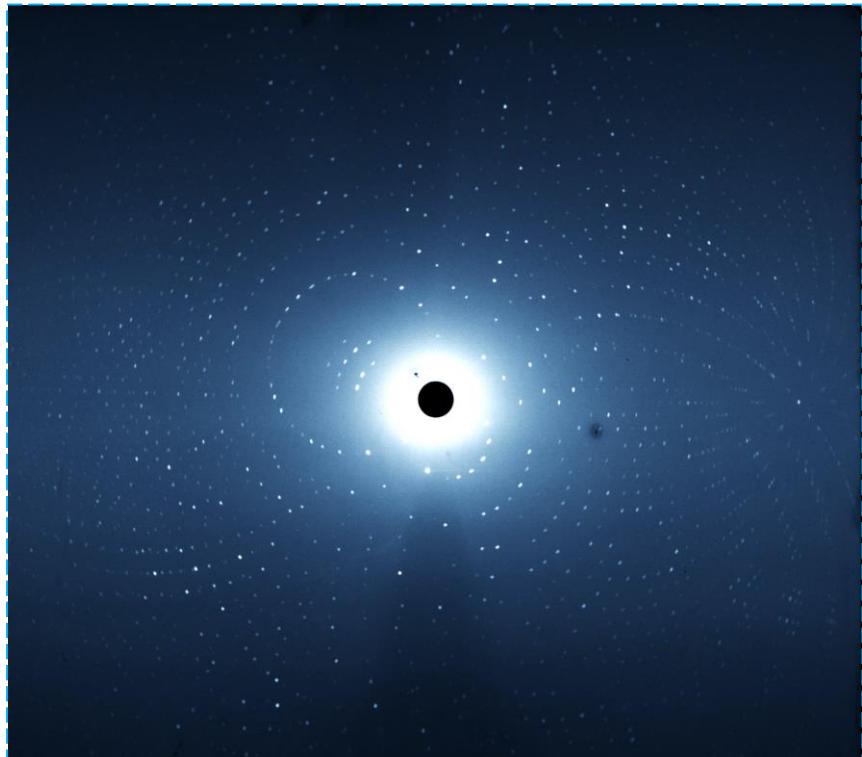
**"After purification, 220 mg of lyophilized Fuc-d<sub>12</sub> was obtained from a culture using 1.5 L of D<sub>2</sub>O and 45 g of deuterated glycerol"**

Gajdos L. et al., Glycobiology, 2021

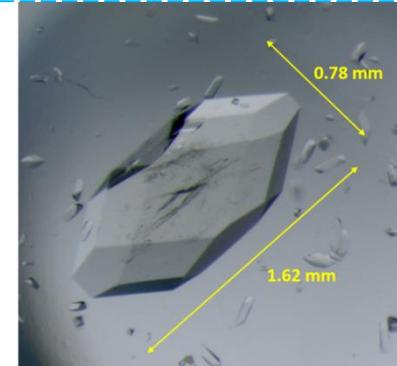
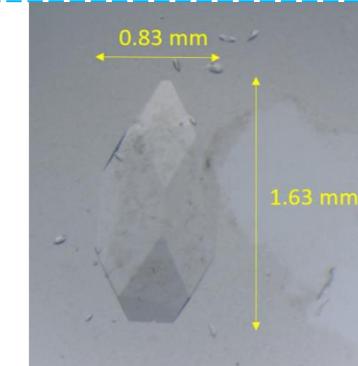
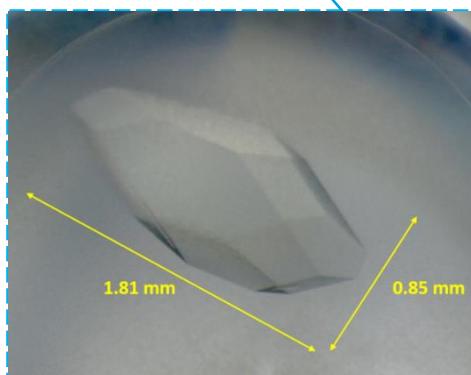
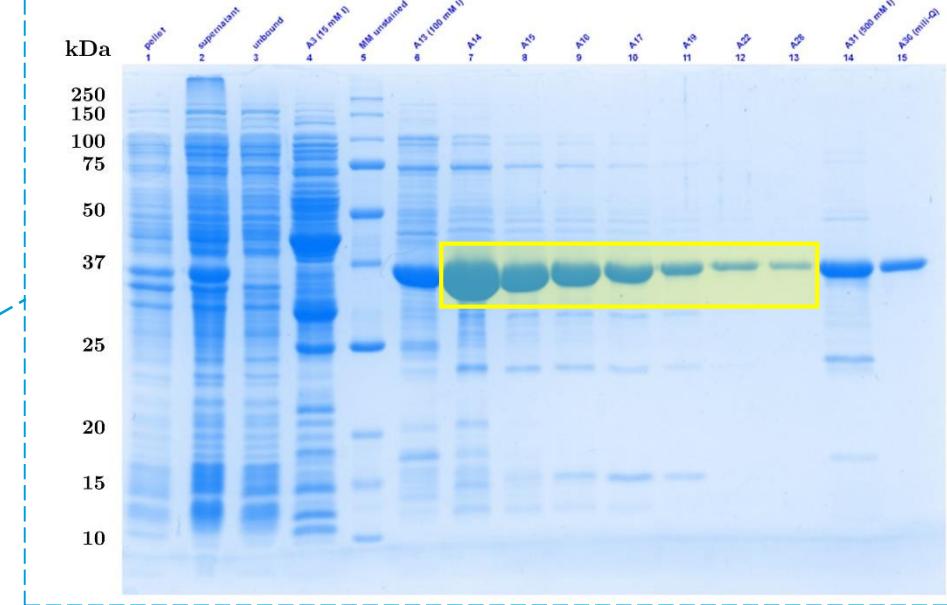
# Production and crystallization of lectins



Fermentation  
66 g of wet cell paste



5 mg of protein /g of cell paste  
Total 330 mg of protein

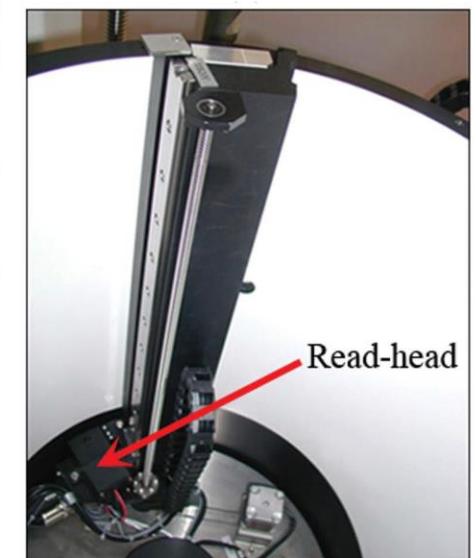
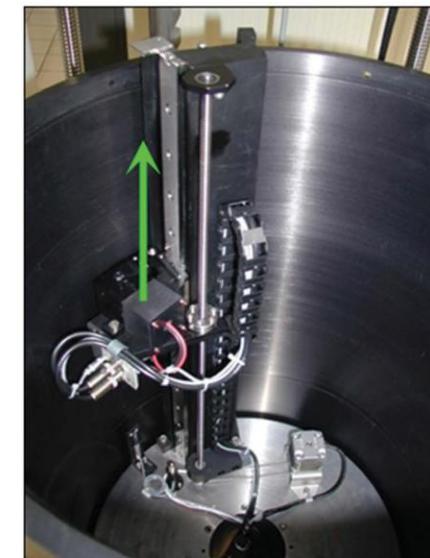
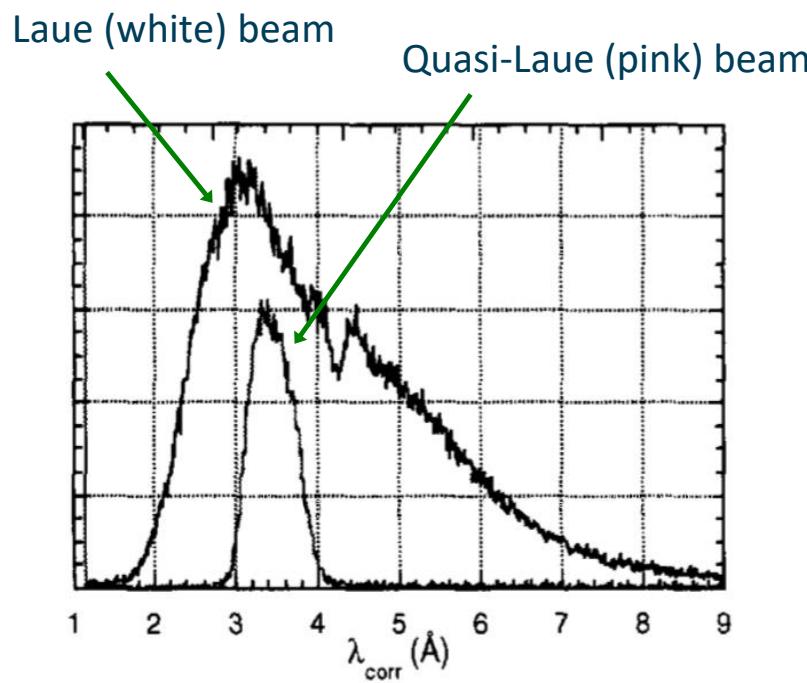
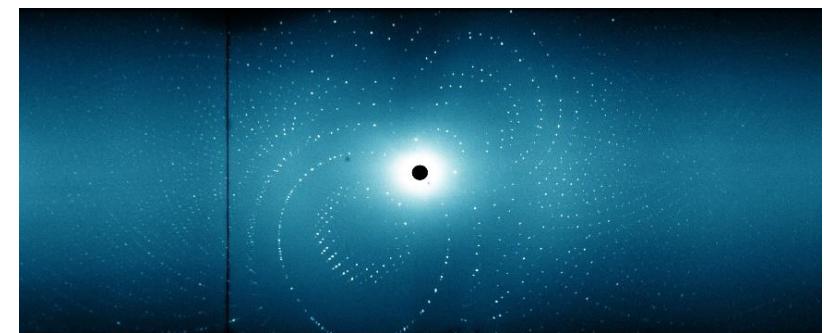


# Neutron data collection summary (LADI-III at ILL)

|  | PLL lectin        |                           | D-LecB lectin       |  |
|--|-------------------|---------------------------|---------------------|--|
|  | H/D-exchanged apo | D-PLL/Fuc-d <sub>12</sub> | Fuc-d <sub>12</sub> |  |
| Number of proposals                      | 400 µm<br>1       | 400 µm<br>1               | 400 µm<br>1         |  |
| Number of crystals<br>(tested/collected) | 4/2               | 6/1                       | 3/1                 |  |
| Crystal volume (mm <sup>3</sup> )        | ~0.5              | ~0.5                      | ~0.1                |  |
| Neutron data collection (days)           | 8                 | 15                        | 18                  |  |
| Unit cell dimensions                     |                   |                           |                     |  |
| <i>a, b, c</i> (Å)                       | 72.7, 89.3, 159.4 | 72.7, 89.2, 159.2         | 52.9, 73.9, 55.0    |  |
| $\alpha, \beta, \gamma$                  | 90, 90, 90        | 90, 90, 90                | 90, 94.6, 90        |  |
| Resolution (Å)                           | 2.2               | 2.2                       | 1.9                 |  |
| Completeness (%)                         | 82.9 (73.5)       | 84.9 (74.7)               | 73.8 (61.7)         |  |

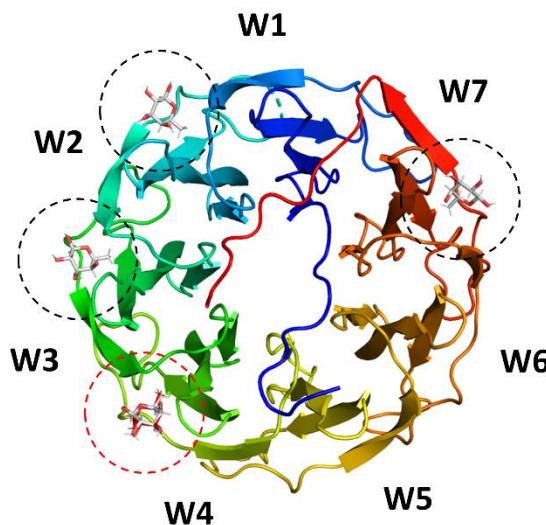
# Laue diffractometer LADI-III at ILL

- Institut Laue-Langevin (ILL) in Grenoble, France
- Quasi-Laue diffraction method (pink beam of neutrons)
- Large cylindrical neutron-sensitive image plate detector
- Data collection at room temperature

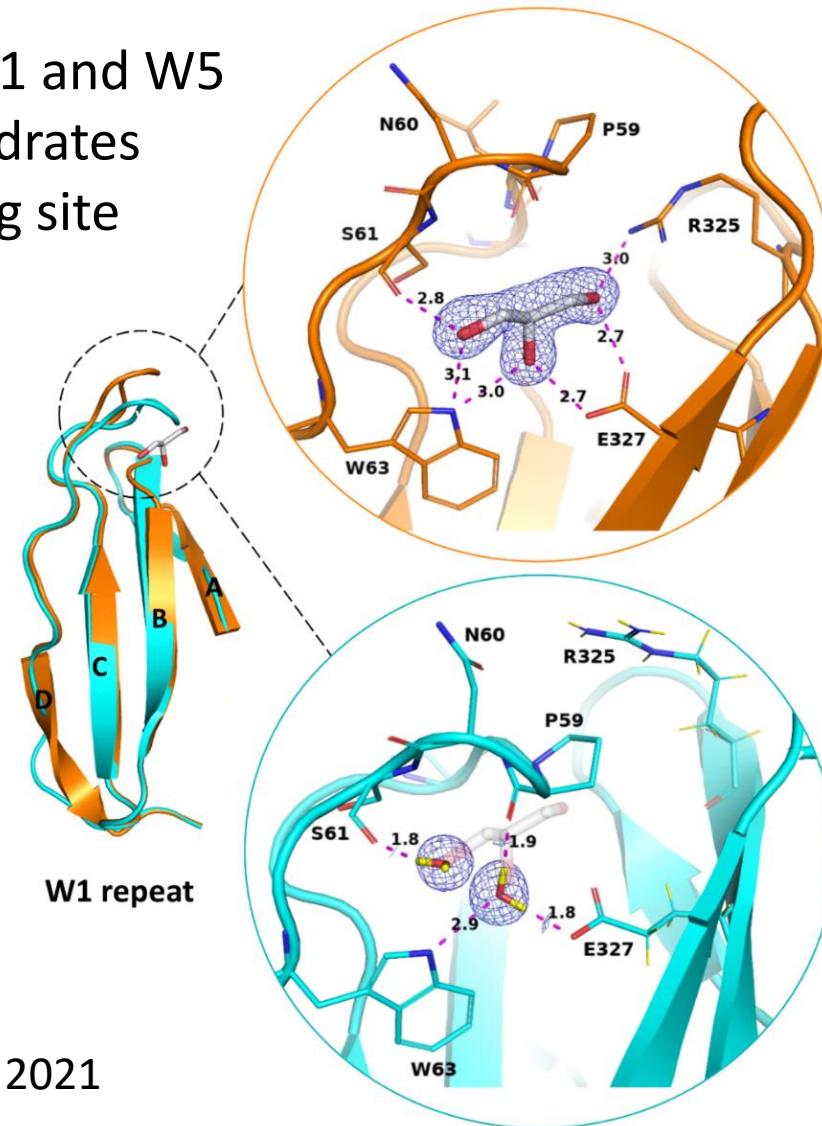


# Room temperature (RT) versus cryo (100K) data

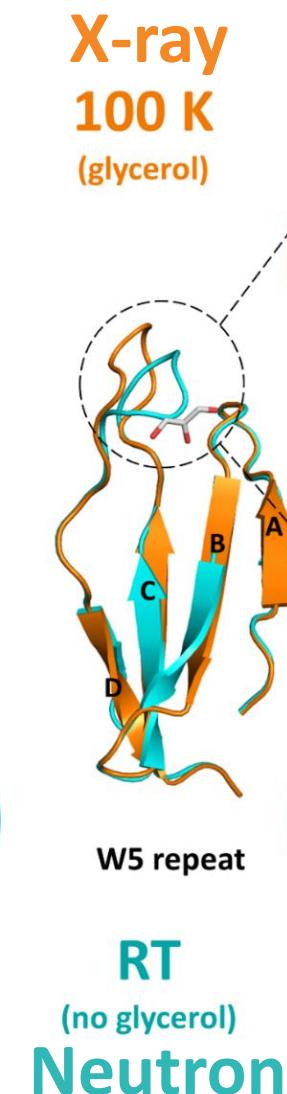
- Differences in repeats W1 and W5
- Glycerol mimics carbohydrates
- Additional fucose-binding site



Gajdos L. et al., Structure, 2021

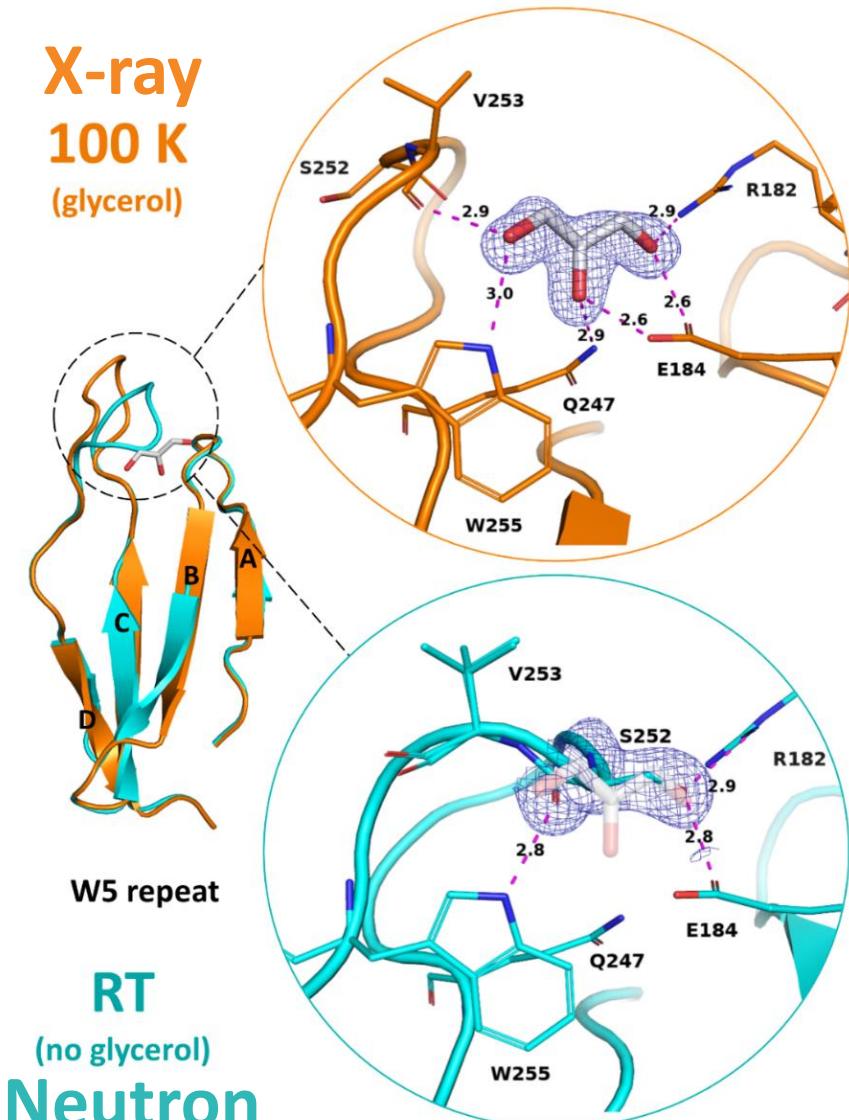


W1 repeat



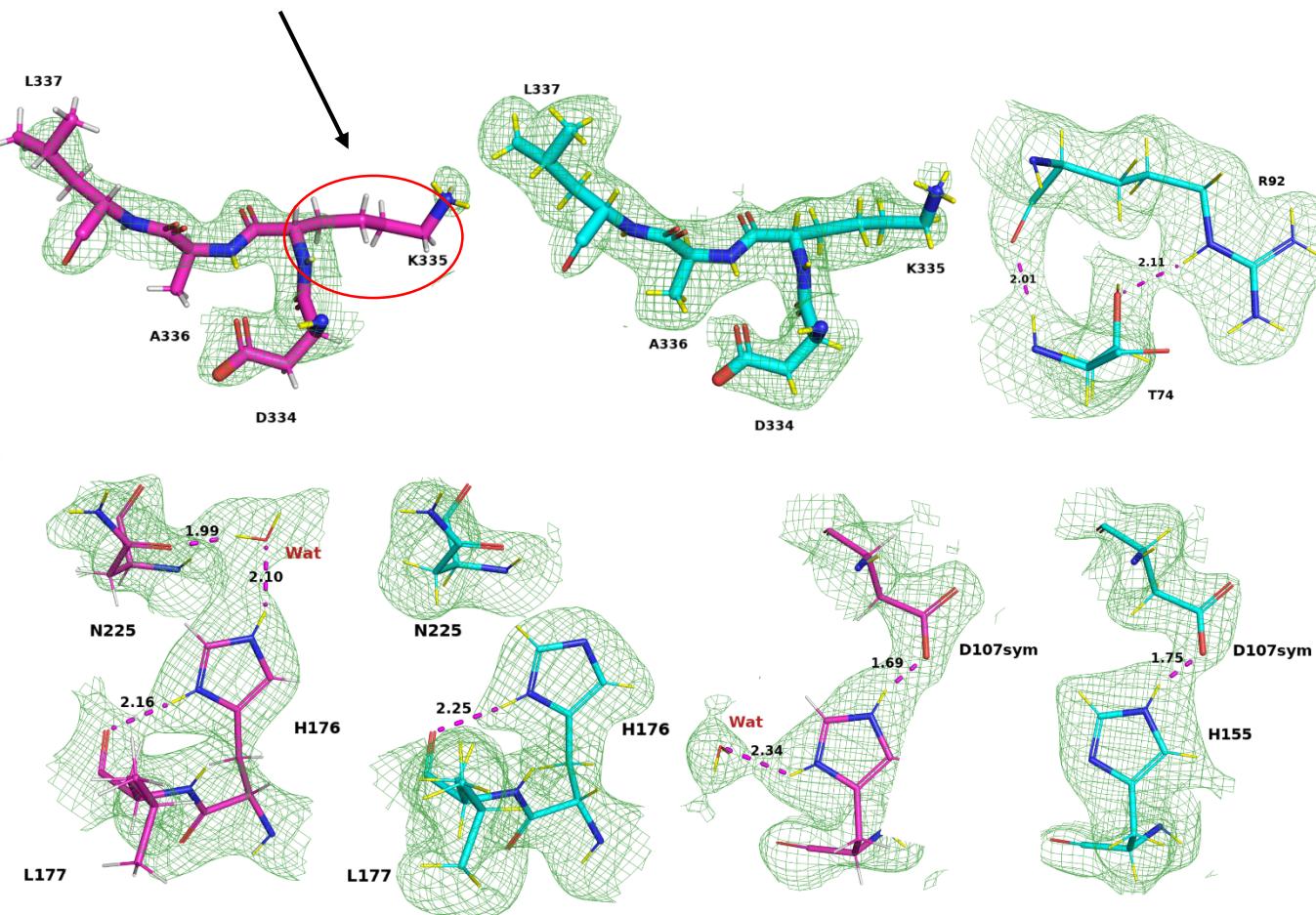
X-ray  
100 K  
(glycerol)

RT  
(no glycerol)  
Neutron

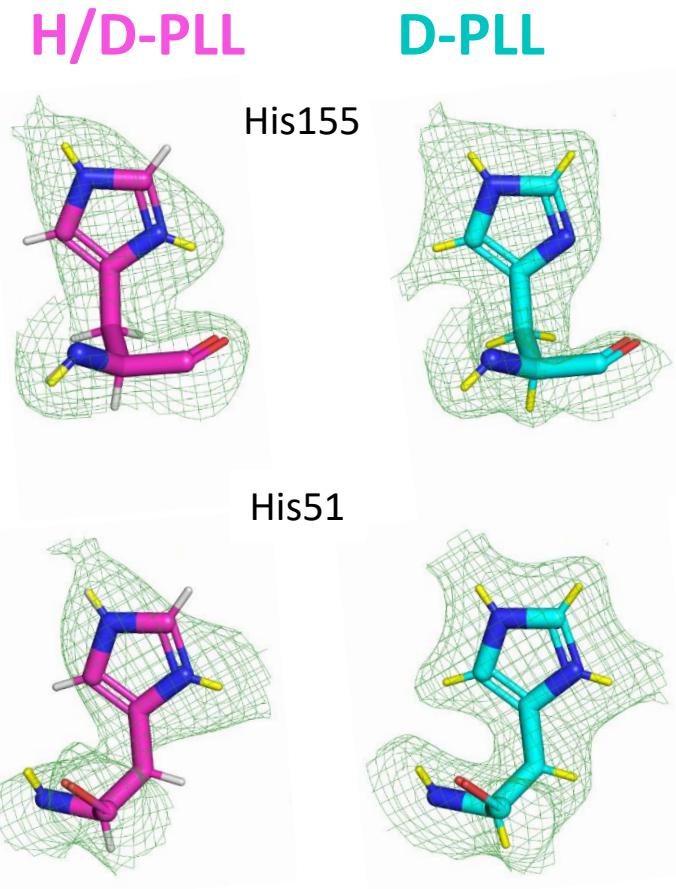


# Advantages of perdeuteration in neutron structures

- H/D-exchanged apo PLL and D-PLL
- Cancellation effects around CH<sub>2</sub> groups

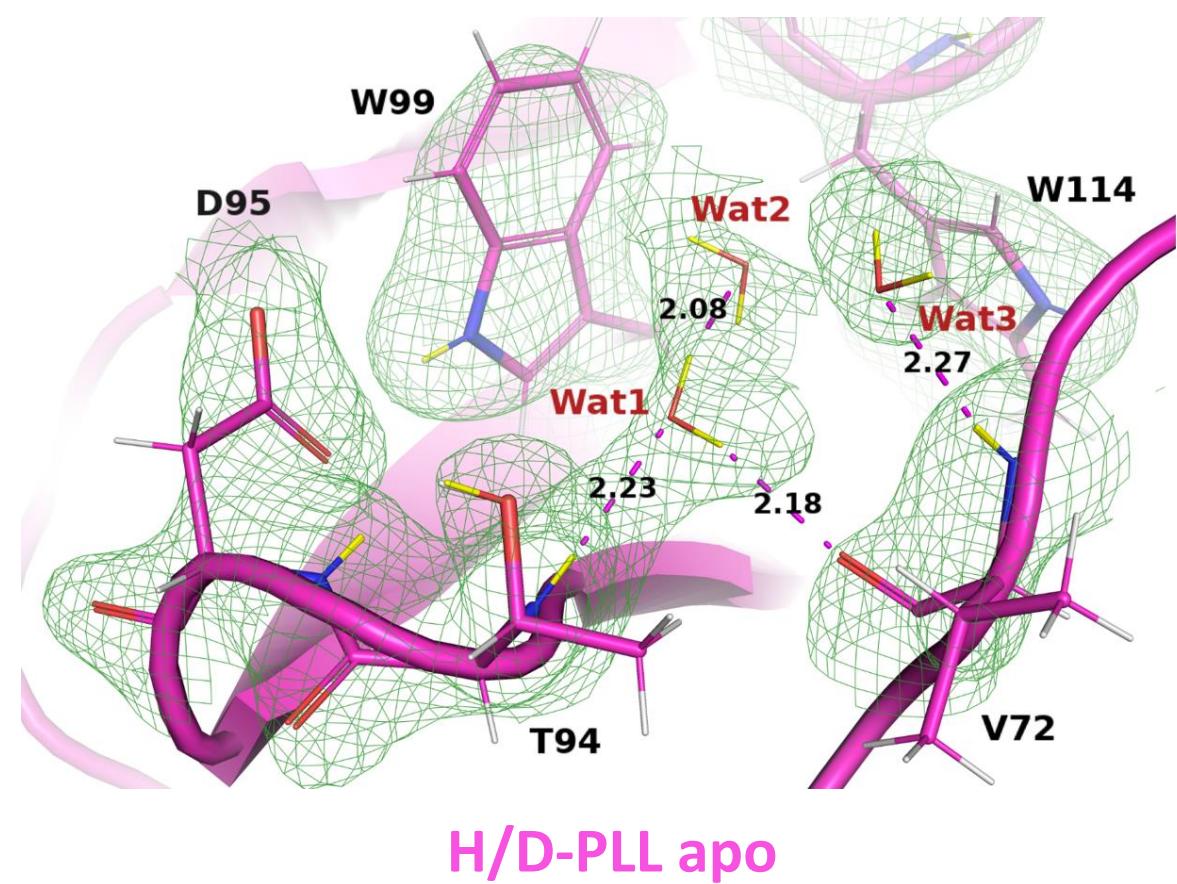
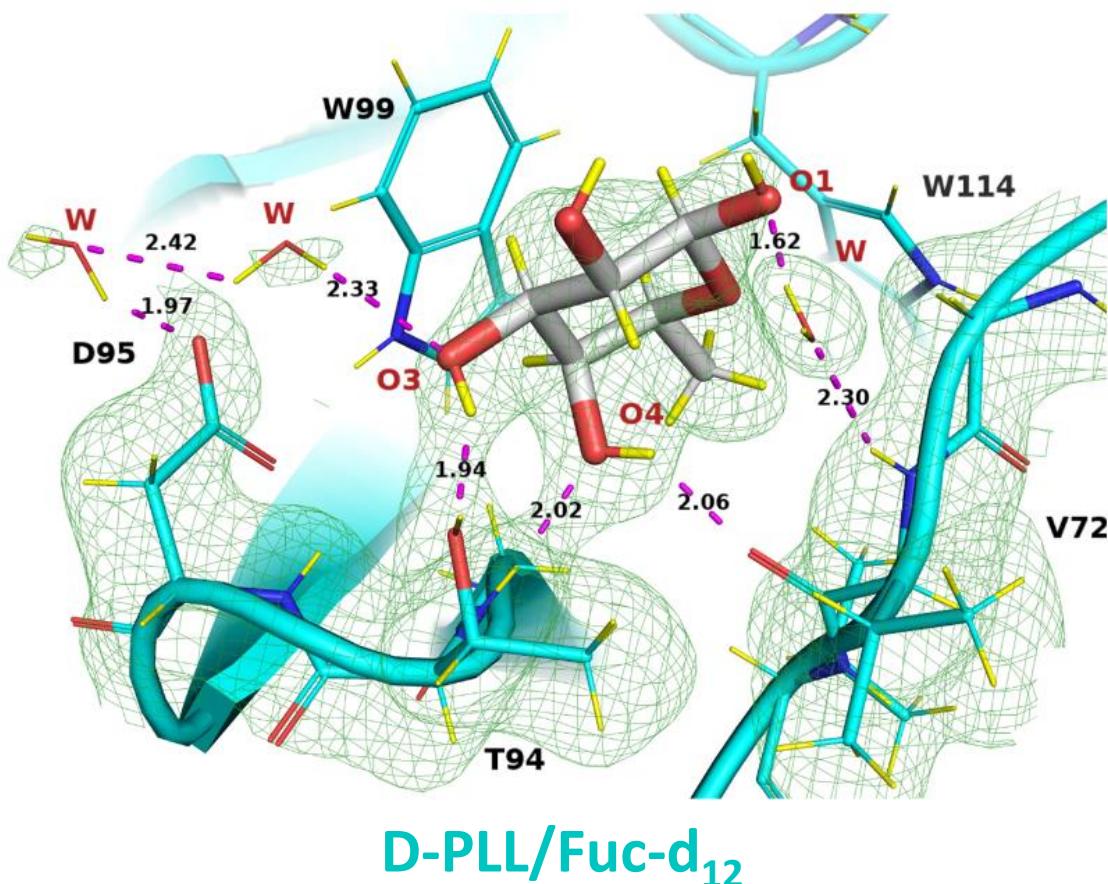


Protonation state of histidines



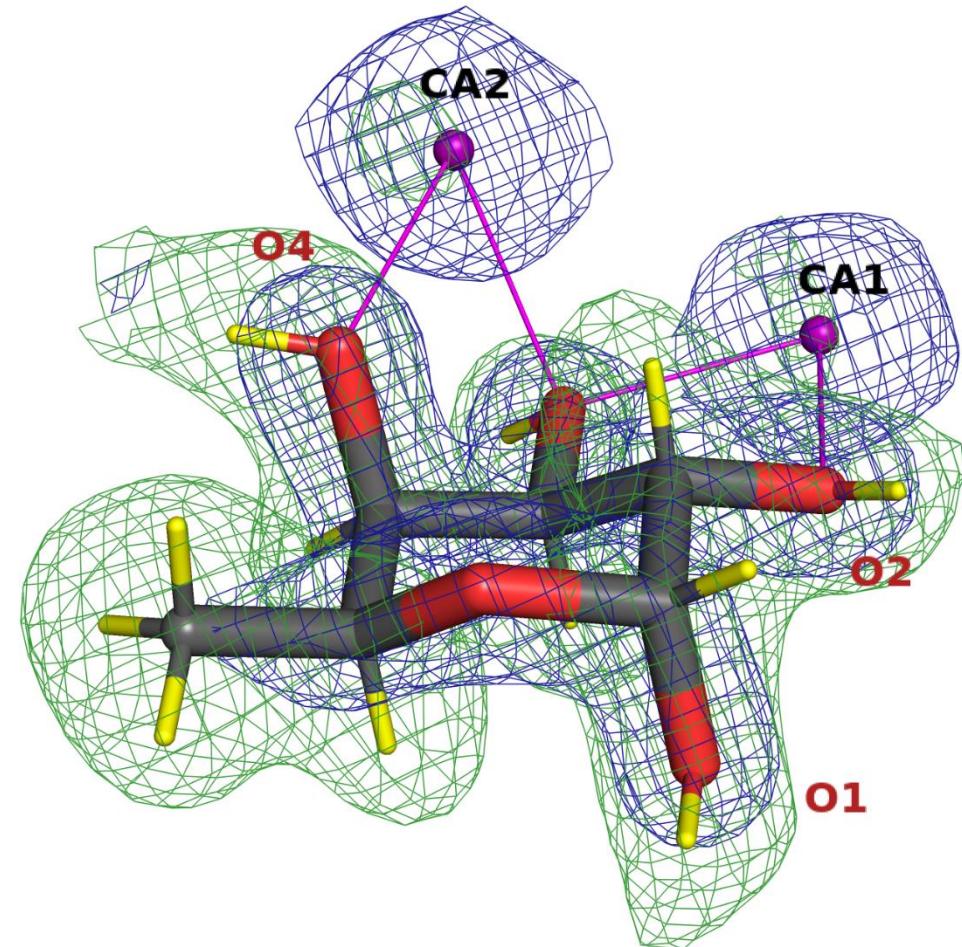
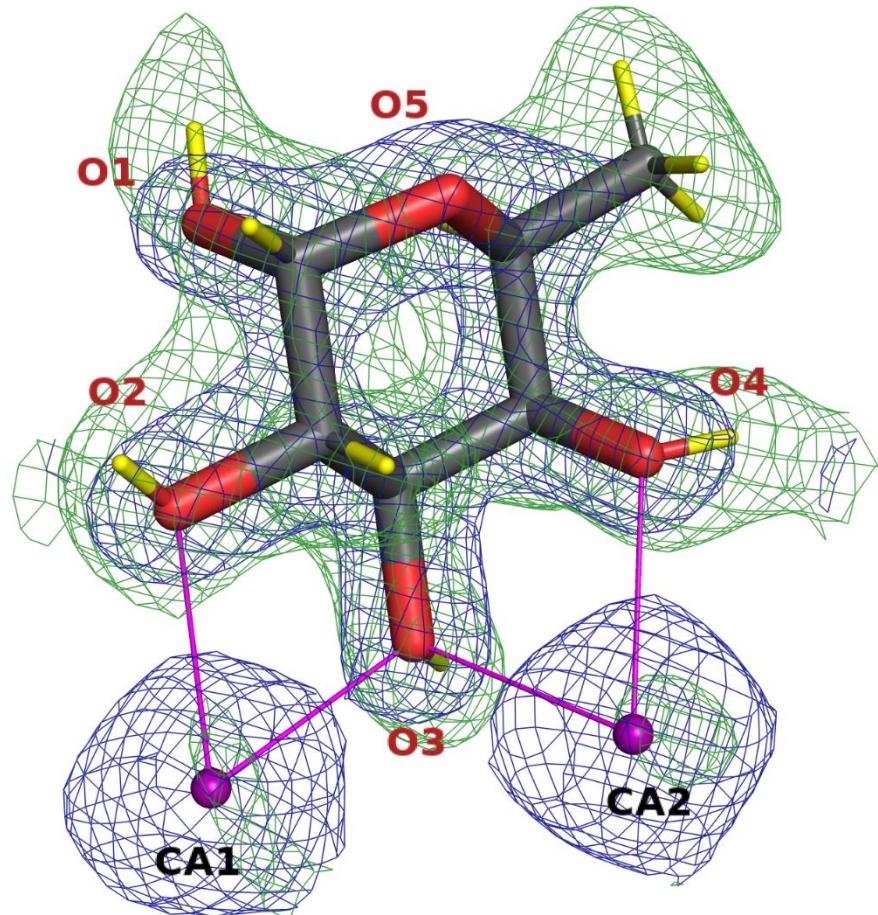
# Binding site details of the D-PLL/Fuc-d<sub>12</sub> complex

- Direct hydrogen bonds
- Water-mediated interactions
- CD-π stacking interaction between the apolar face of fucose and tryptophan residues



# Perdeuterated fucose in the LecB binding site

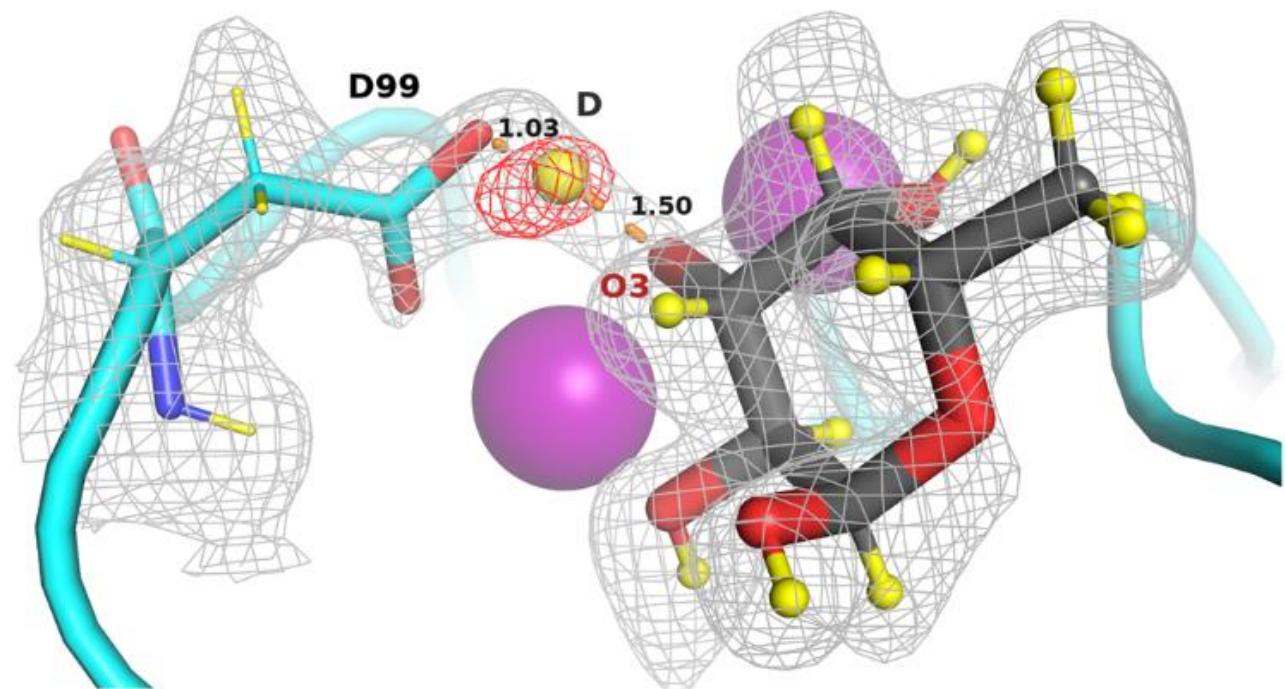
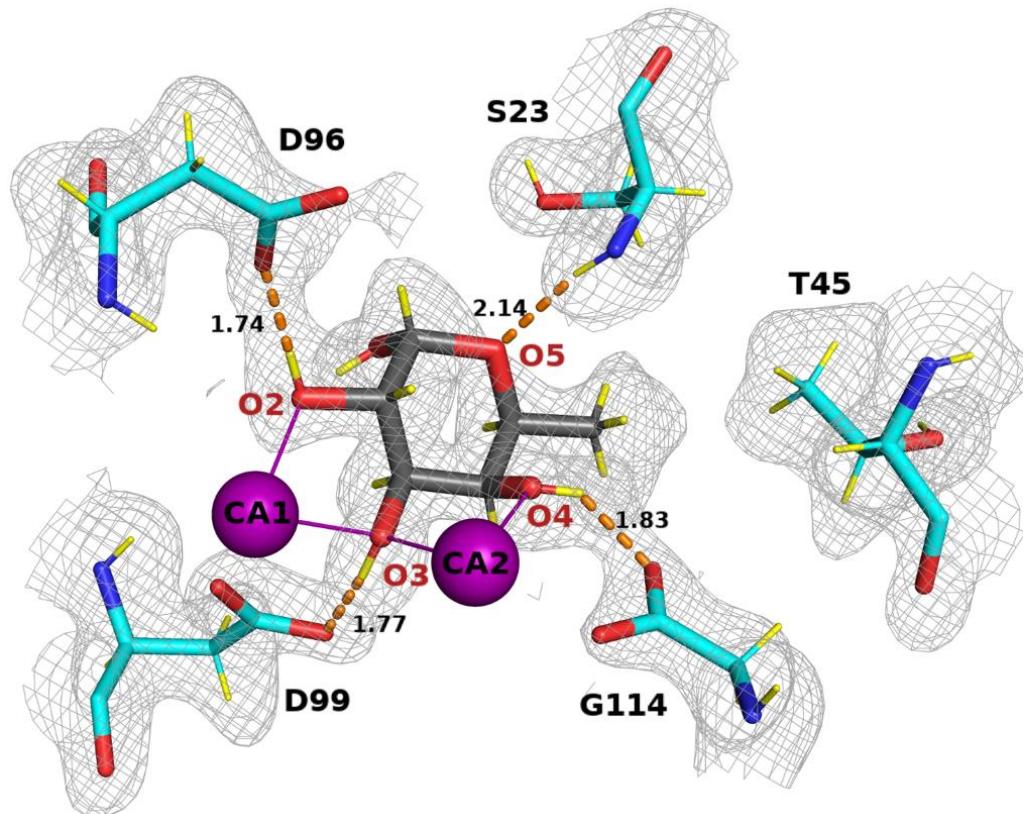
- Electron density ( $1.4\sigma$ )
- Neutron density ( $2.2\sigma$ )



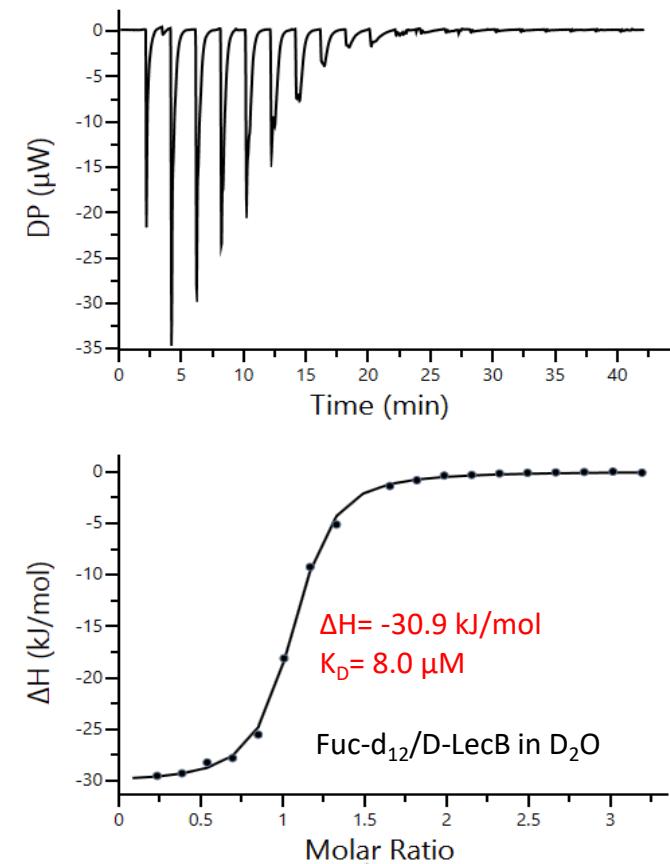
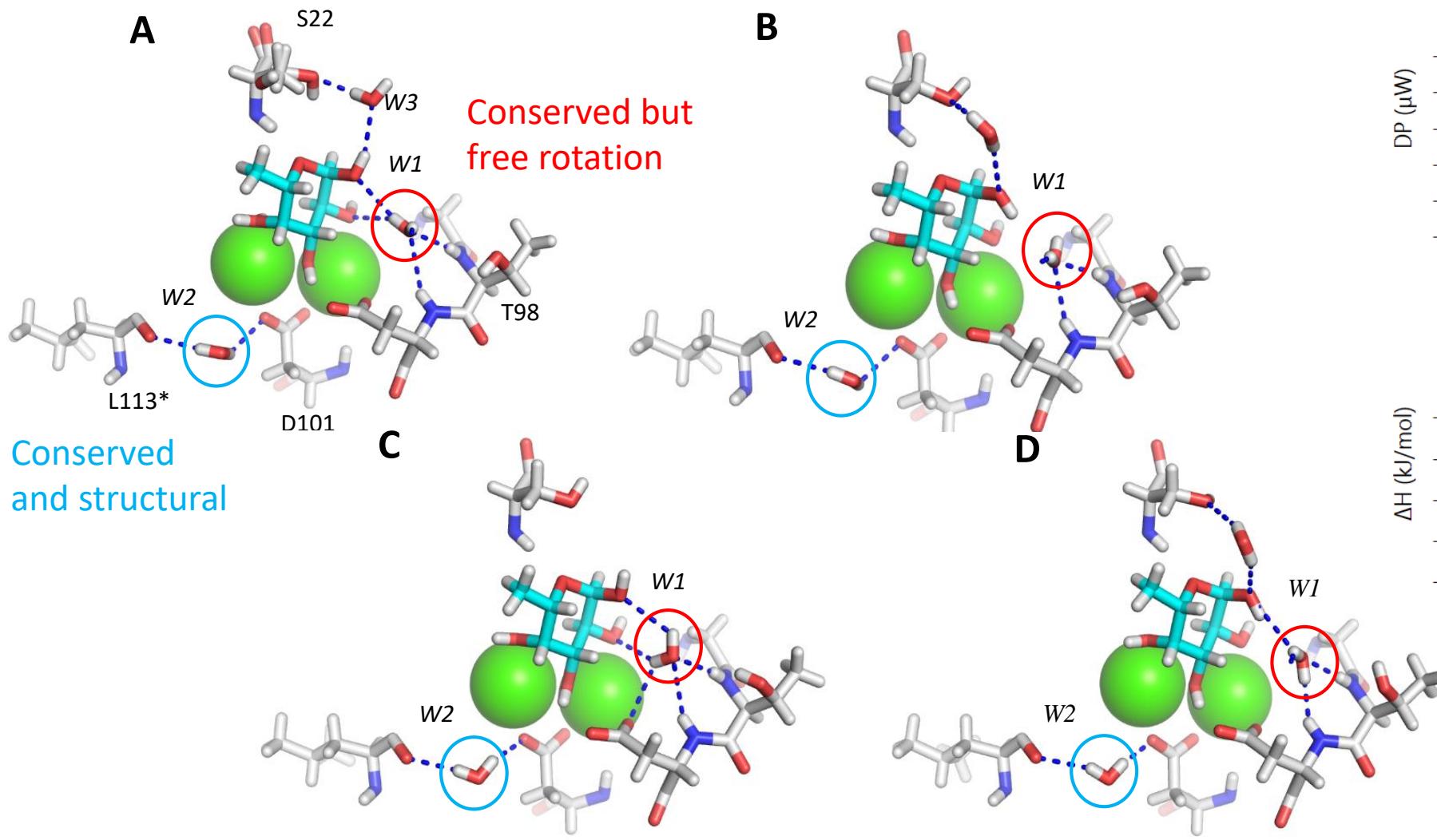
Gajdos L. et al., (2022) *Nat Commun*

# H-bonding network and a low-barrier H-bond

- 4 direct H-bonds (orange dashed lines) with the protein + hydrophobic interaction
- Charged amino acid residues are non-protonated
- Delocalized electrons contribute to the overall net charge -2
- A low-barrier hydrogen bond formed in the proximity of calcium ions

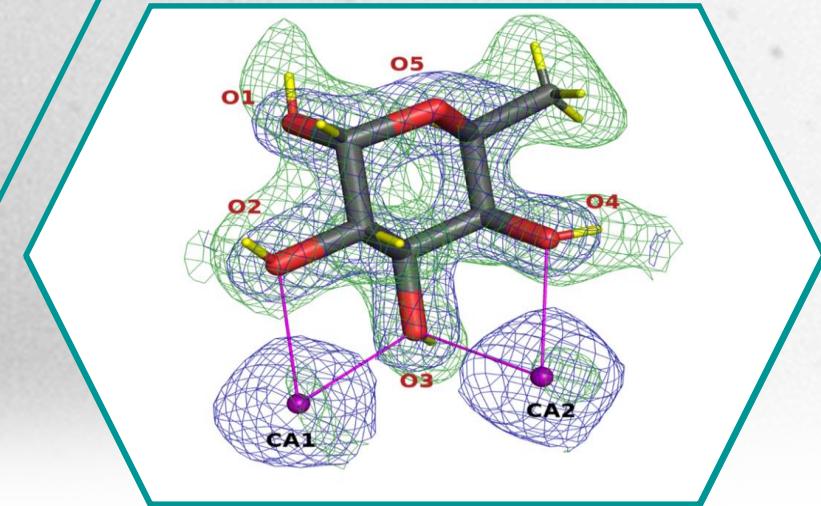
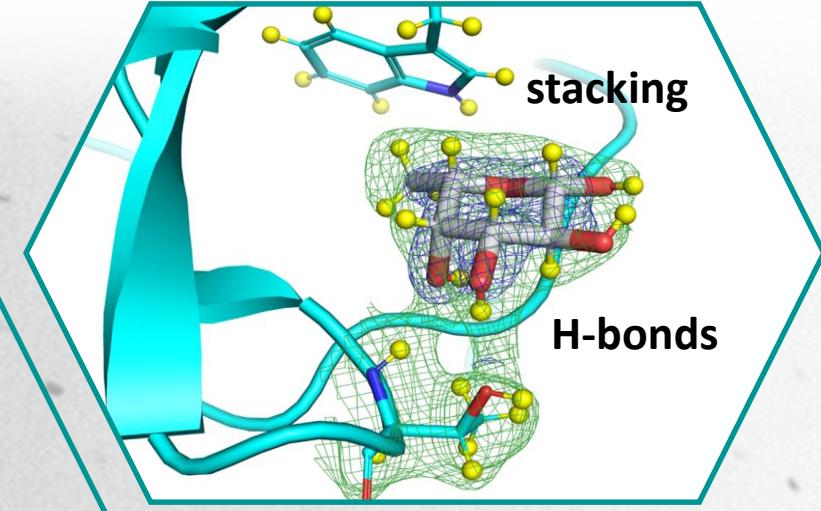
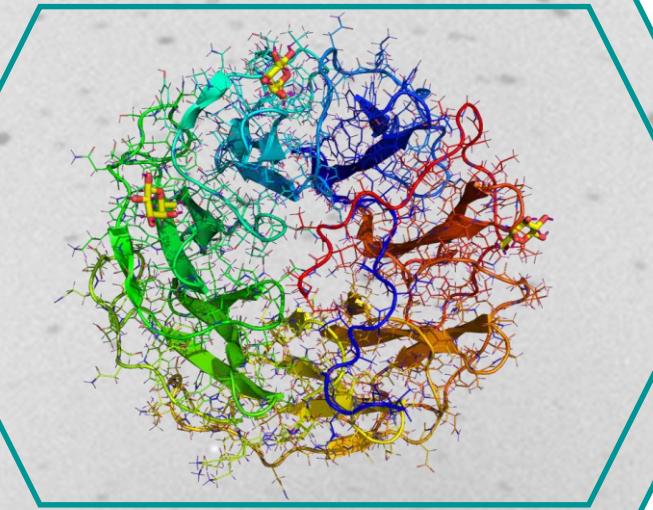
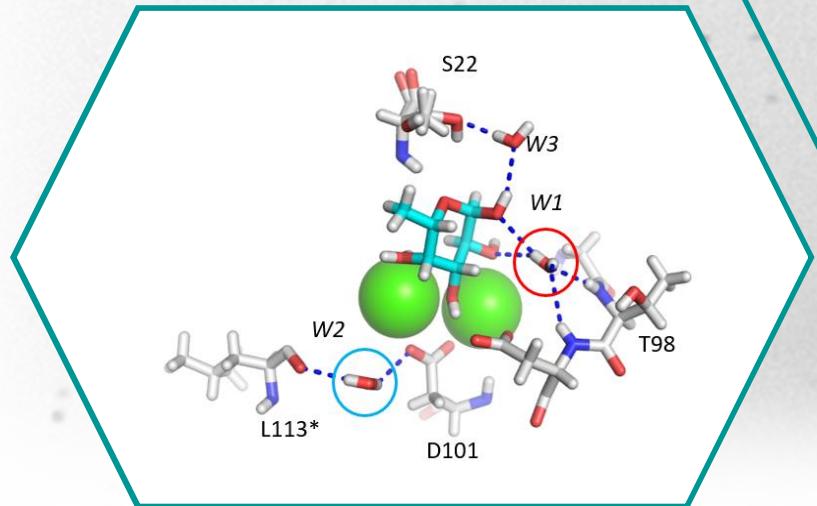
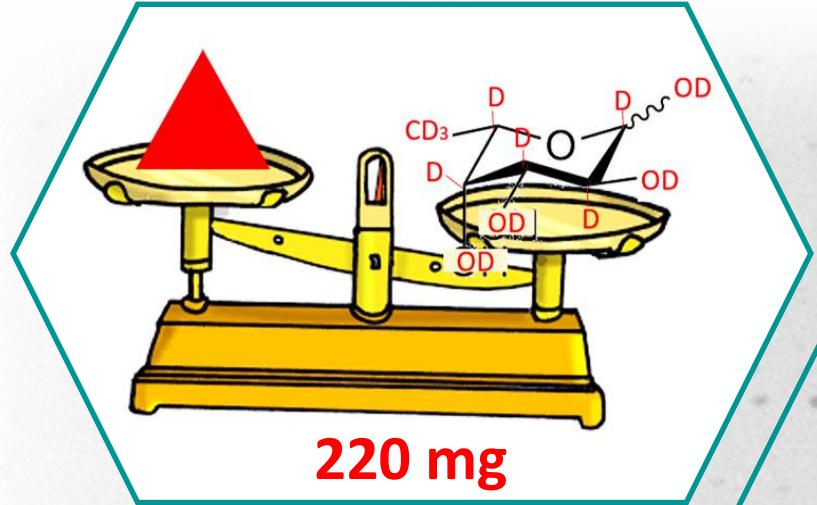


# Water network in the fucose binding site of LecB



Gajdos L. et al., (2022) *Nat Commun*

# Summary



# Acknowledgements

ILL

**Juliette Devos**

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Annabelle Varrot

Eric Samain

Structural and Molecular  
Biology group

CEITEC

Michaela Wimmerová

Atul Kumar

Glycobiology group

