

# Connecting dynamics and phase behavior of proteins: The neutron perspective (and beyond)

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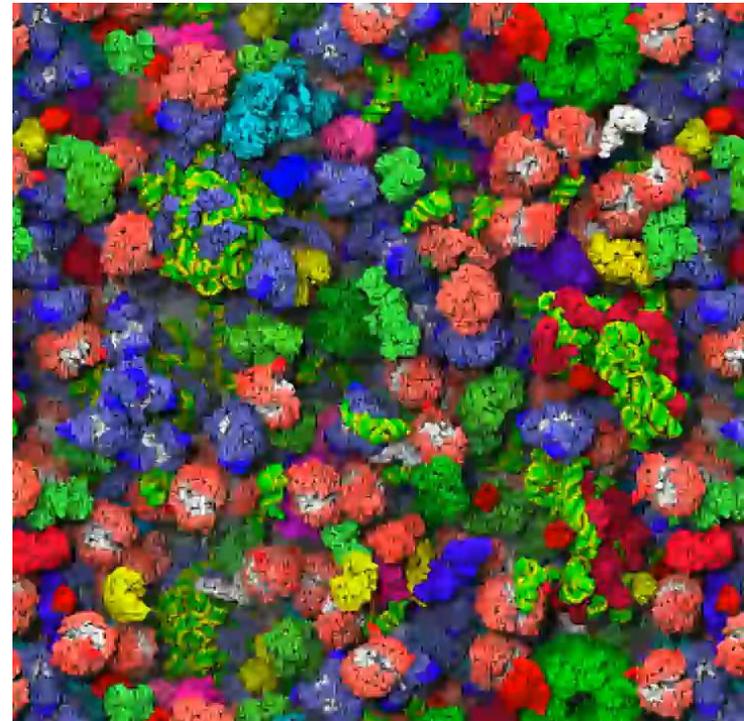
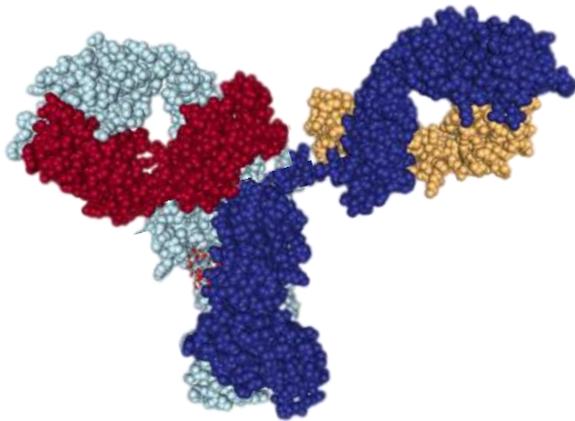
*A. Girelli, N. Begam, A. Ragulskaya, S. Chandran, F. Zhang, R. Roth, M. Oettel (Tübingen)*

*H. Rahmann, C. Gutt (Siegen), J. Möller, M. Reiser, F. Westermeier, M. Sprung (Hamburg), F. Roosen-Runge (Malmö)*

*T. Seydel, C. Beck, O. Matsarskaia, K. Pounot, I. Mosca, T. Forsyth, H. Lopez, J.-L. Barrat, T. Narayanan (Grenoble)*

*R. Jacobs (Oxford), M. Skoda (Rutherford)*

*et al.*

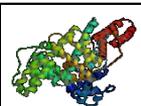
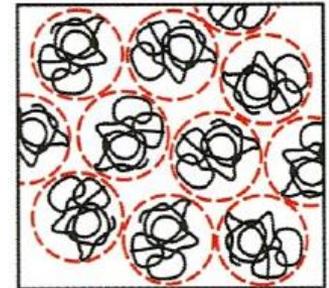
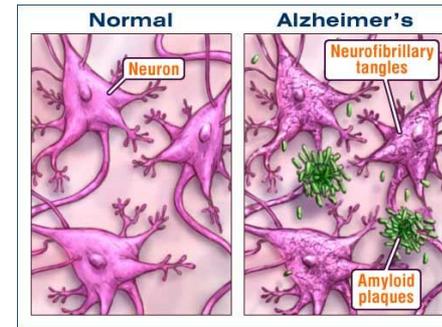
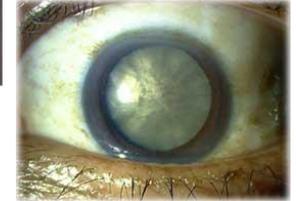
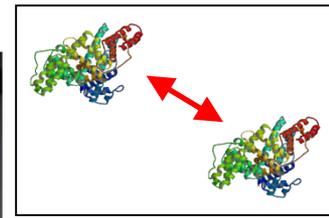
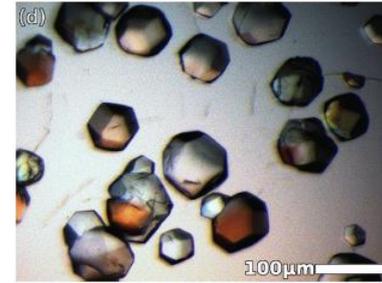


πάντα ῥεῖ

From individual proteins to ensembles

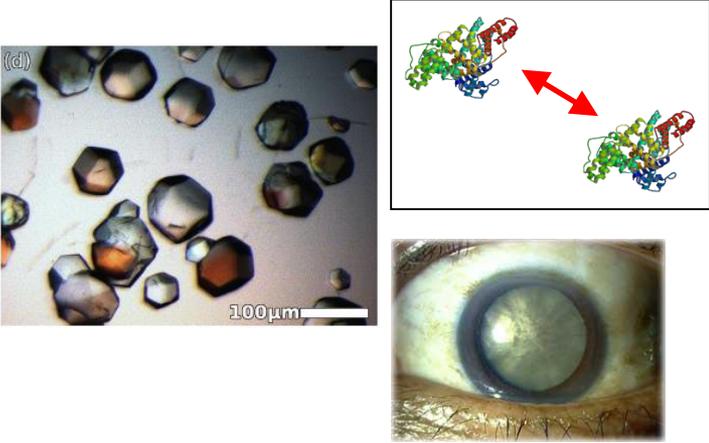
# Interactions and Aggregation

- understanding crystallization (structural biology!)
- understanding phase behavior (stability of pharmaceuticals)
- eye as an example:
  - inside: avoid cataract (LLPS)
  - outside: avoid sticking and infection
- aggregation-related diseases (Alzheimer; Kreutzfeld-Jacob; Parkinson)
- *in vivo*: ionic cloud required (salt)
- crowding (high concentration in cells) (key to understanding of kinetics etc.; Ellis et al.)
- 'extreme conditions'
  - highly concentrated solutions (molecular crowding)
  - high salt concentrations
  - high temperatures (denaturing and network formation)
- note the impact of interfaces (container walls, sensors, implants, ...)
  - adsorption effects
  - nucleation effects (see Fries et al., PRL 2017; Sci. Rep. 2020)



# Interactions and Aggregation

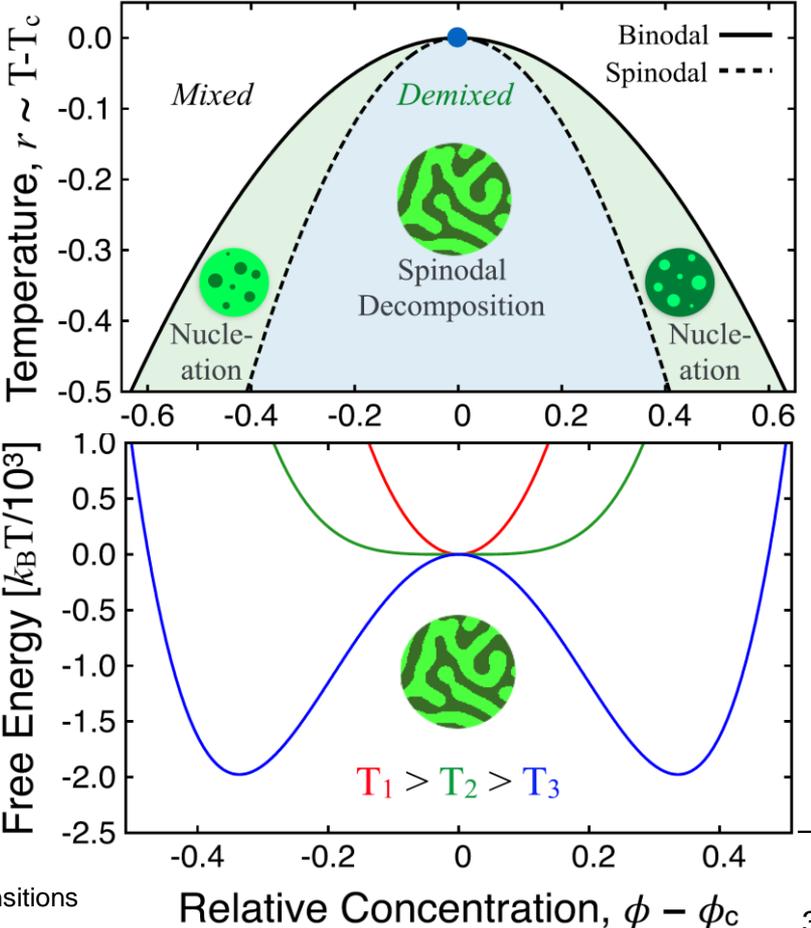
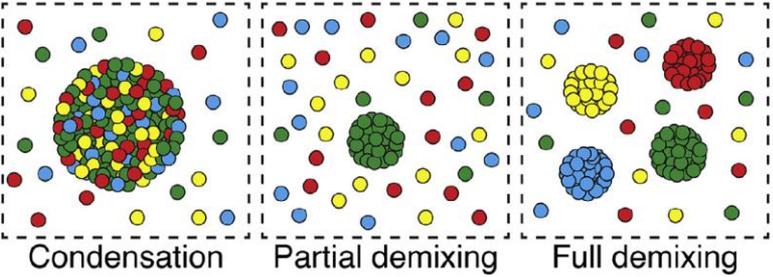
- understanding crystallization (structural biology!)
- understanding phase behavior (stability of pharmaceuticals)
- eye as an example
- ...



## Theory ?

Free energy of a multi-component system

$$f_{RS} = \sum_{i=1}^N \left[ \underbrace{\phi_i \ln(\phi_i)}_{\text{entropy}} + \sum_{j=1 \neq i}^N \underbrace{\frac{\chi_{ij}}{2} \phi_i \phi_j}_{\text{interactions}} + \sum_{j=1}^N \underbrace{\frac{K_{ij}}{2} (\nabla \phi_i) \cdot (\nabla \phi_j)}_{\text{interfaces}} \right]$$

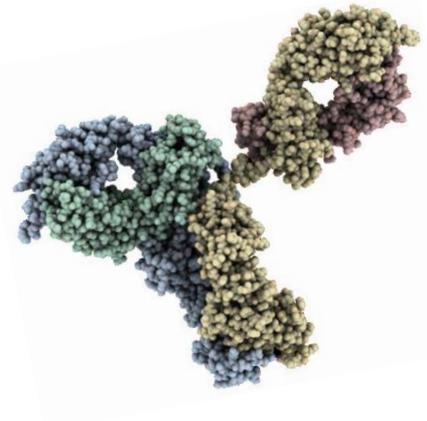


# Protein Dynamics: Going beyond "Static" Phase Behavior

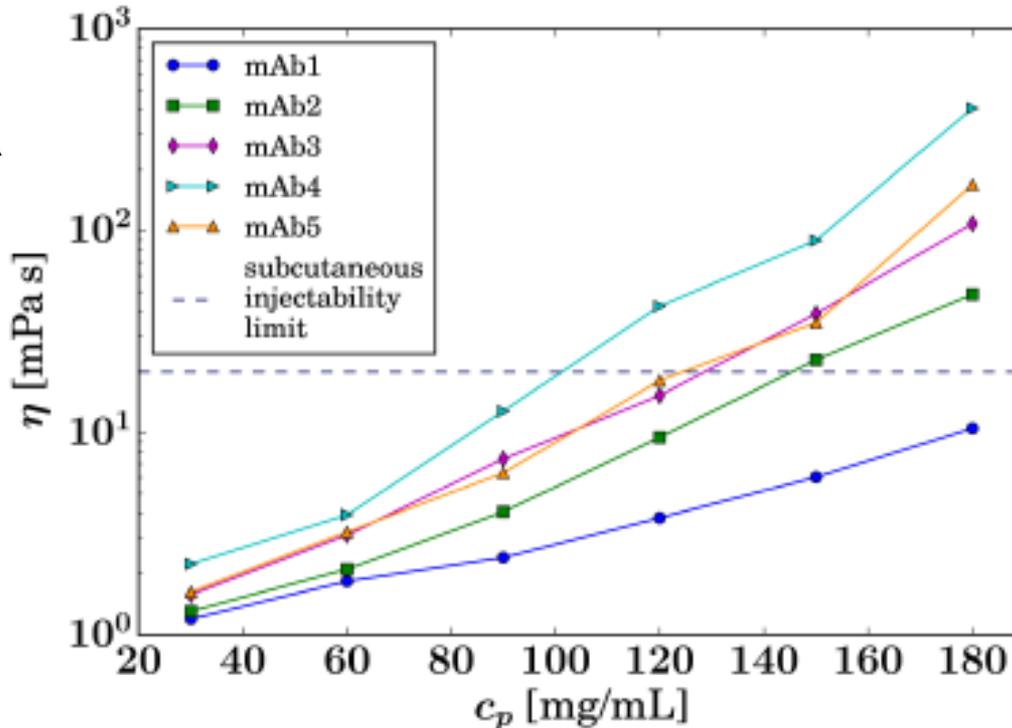


# Protein Dynamics: Macroscopic Viscosity $\eta$

Viscosity  $\eta$  has to stay below threshold for injection !



Viscosity  $\eta$



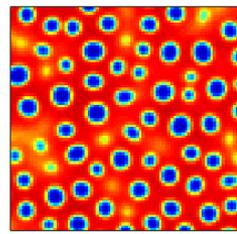
Diffusion  $D \sim 1 / \eta$   
(Stokes-Einstein)

See also talk 16:15  
by Ilaria Mosca

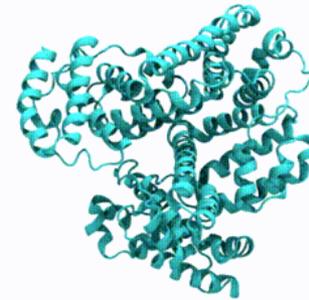


# Protein Dynamics: Types of Motion

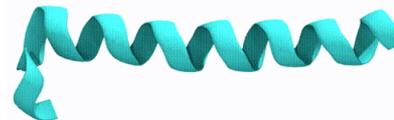
If you want to understand function,  
you have to understand structure  
*and dynamics!*



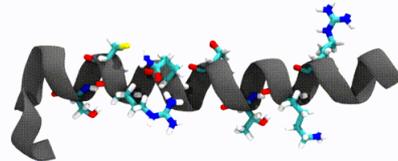
diffusion;  
collective  
motion



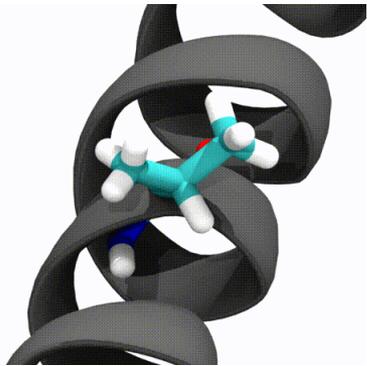
entire protein



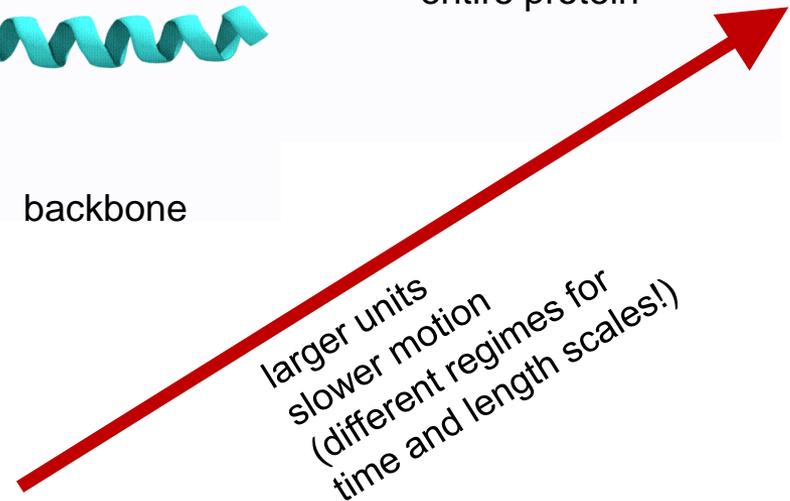
backbone



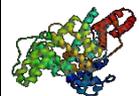
sidechains



methyls



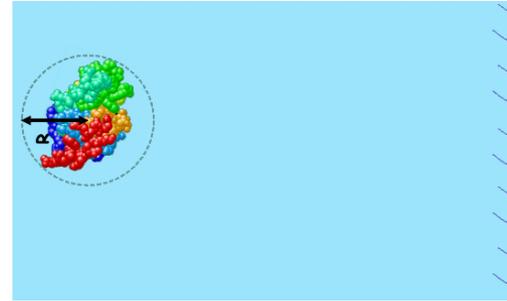
larger units  
slower motion  
(different regimes for  
time and length scales!)



# Protein Dynamics: Control Parameters

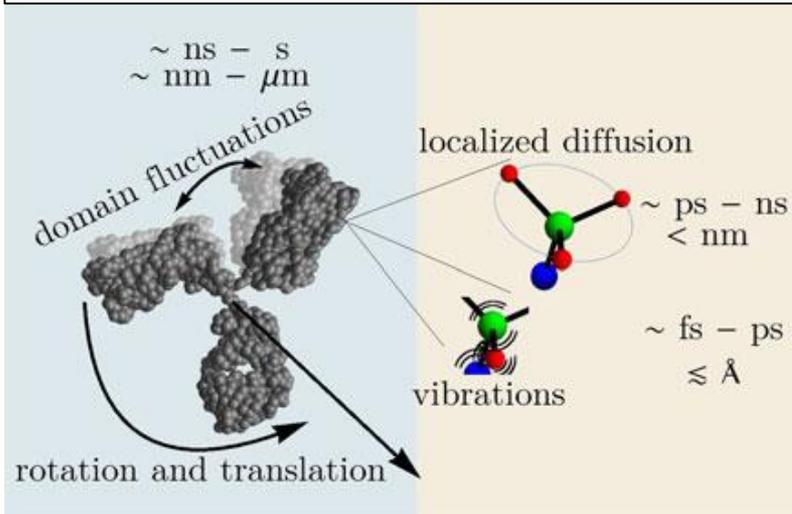


# Protein Dynamics

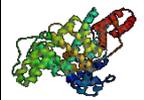
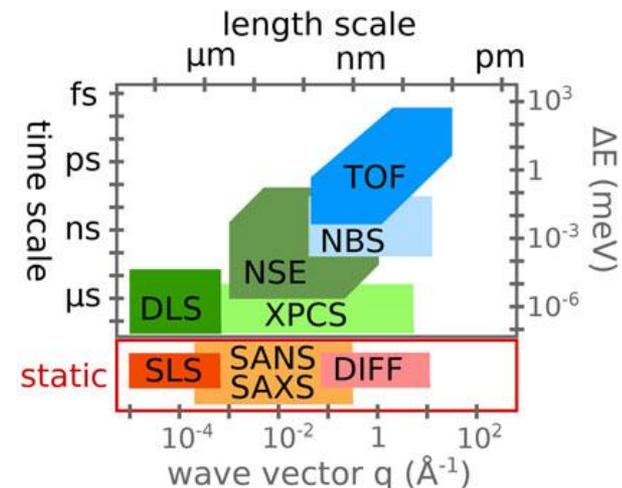


- Effect of  $c_p$  ((self-)crowding) ?
- Effect of  $c_s$  ( $Y^{3+}$  ions near  $c^*$ ) ?
- Effect of lysate (natural crowding) ?
- Effect of crystallization ("dynamics during kinetics") ?
- Effect of  $T$  (denaturation / gel-like network) ?

Generally complex task  
with many different time and length scales



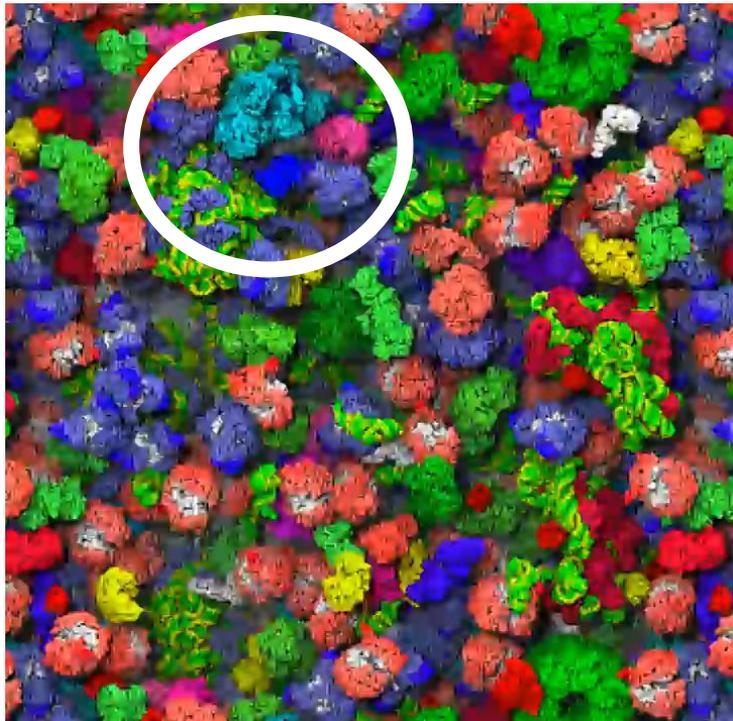
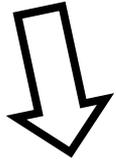
Requires advanced instrumentation  
for different time and length scales



# Protein Dynamics: Crowding

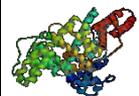
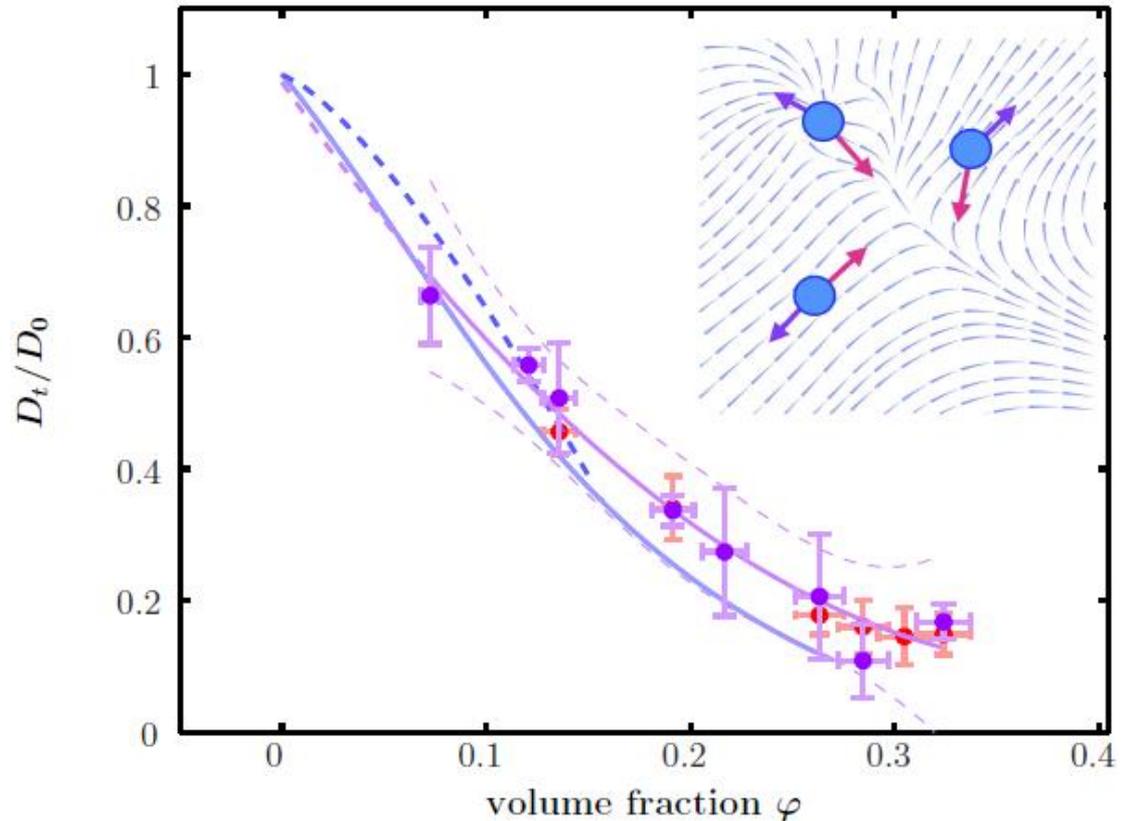
Now targeted deuteration of environment

→ Only proteins of interest visible

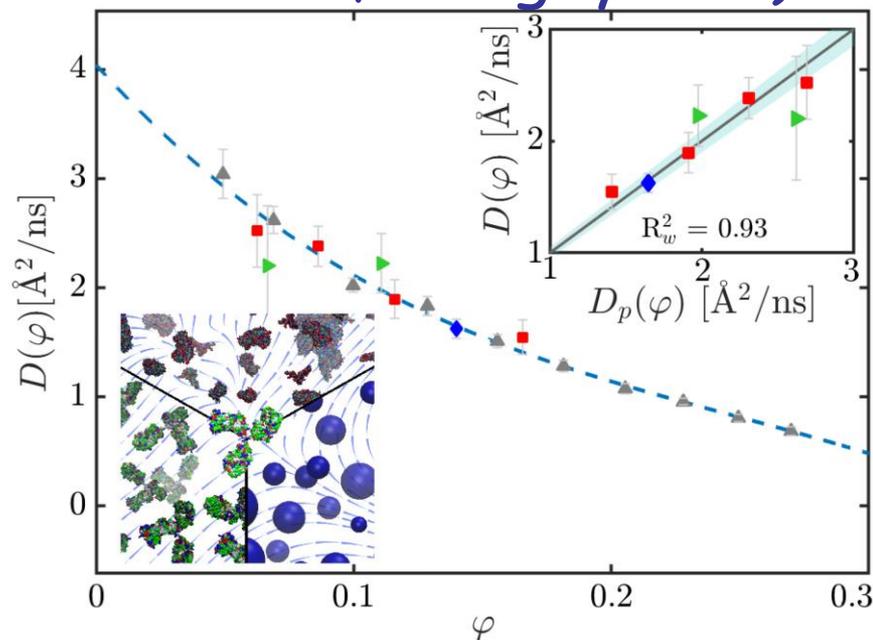
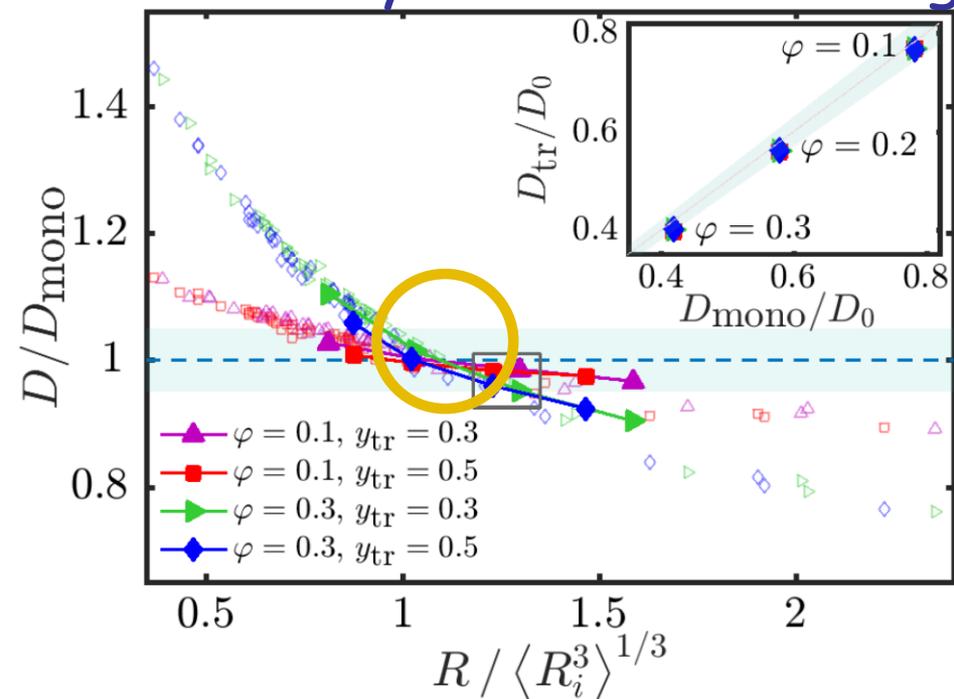


Self-diffusion probed by QENS

- Hydrodynamic interactions important
- 0.3 – 5.0 ns probed are short time diffusion
- no direct interactions
- Slower for higher protein concentration



# Protein Dynamics: Crowding (now natural, using lysate)



Simulated diffusion of spheres as a function of their radius in the Ig-lysate mixture normalized by the diffusion of with self-crowding.

Filled symbols: D of tracer of size R

(R (=1.3) of Ig or other hypothetical R).

Empty symbols: D of the lysate for  $R_{\text{tr}} = R_{\text{Ig}}$ .

The rectangle refer to tracers with  $R_{\text{tr}} = R_{\text{Ig}}$

(only incidentally similar to self-crowding “near 1”).

Measured diffusion as a function of the total volume fraction, i.e. the volume fraction of Ig and lysate combined (T = 293K).

Key conclusion:

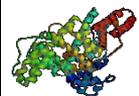
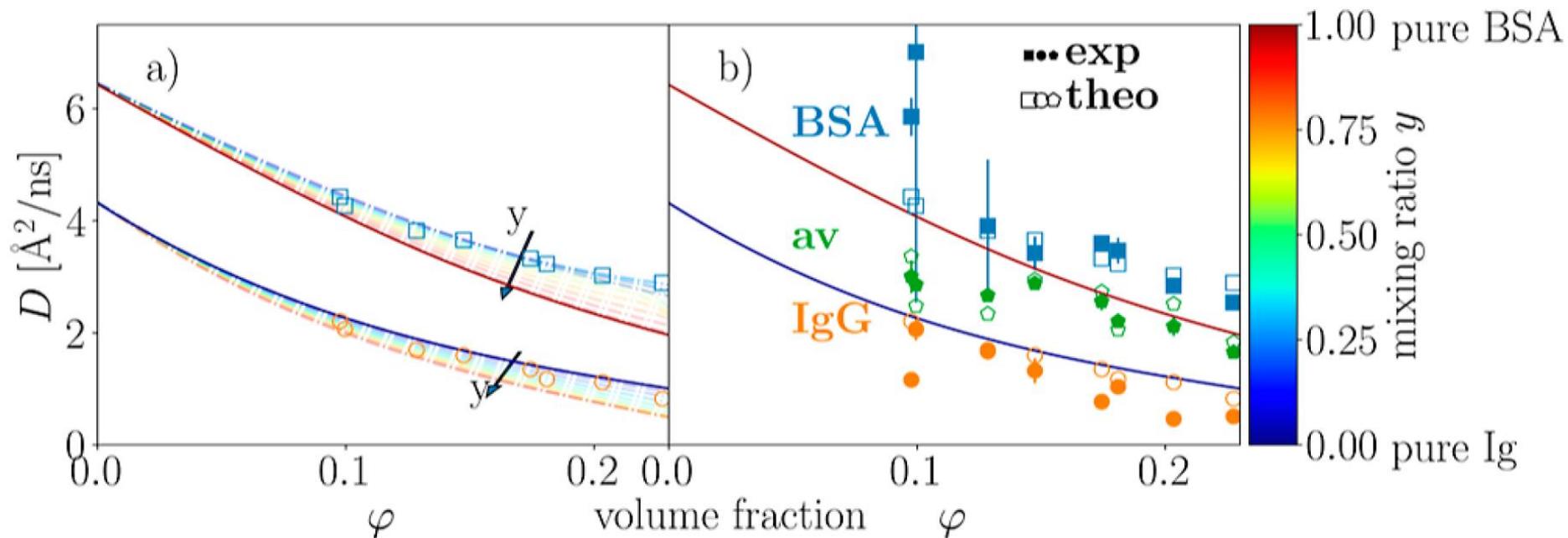
Data for lysate-crowding follow self-crowding.



# Protein Dynamics: Crowding in bidisperse systems

## BSA and IgG mimicking polydispersity in biology

- Diffusion under crowding conditions similar
- Diffusion with mixing consistent (experiment and theory derived from effective hard spheres)
- Diffusion is not simply the average

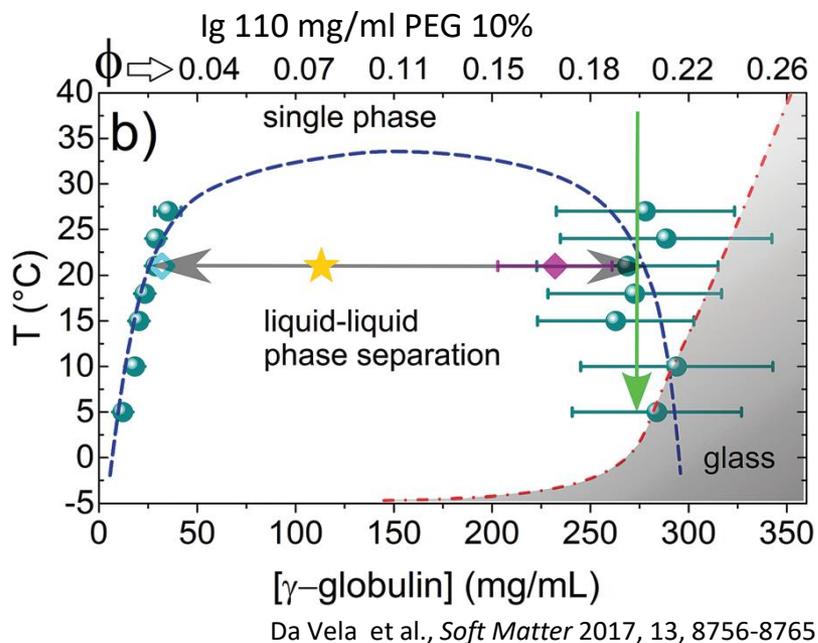
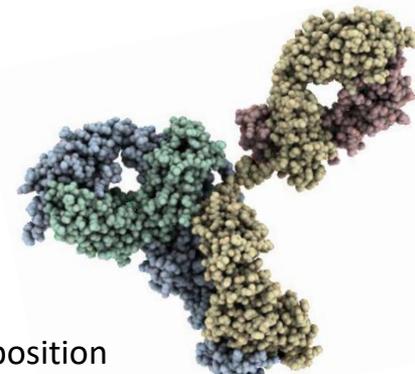


# Protein Dynamics upon Phase Separation

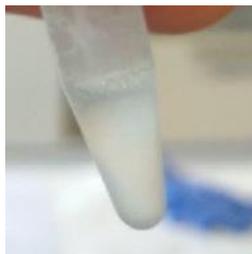
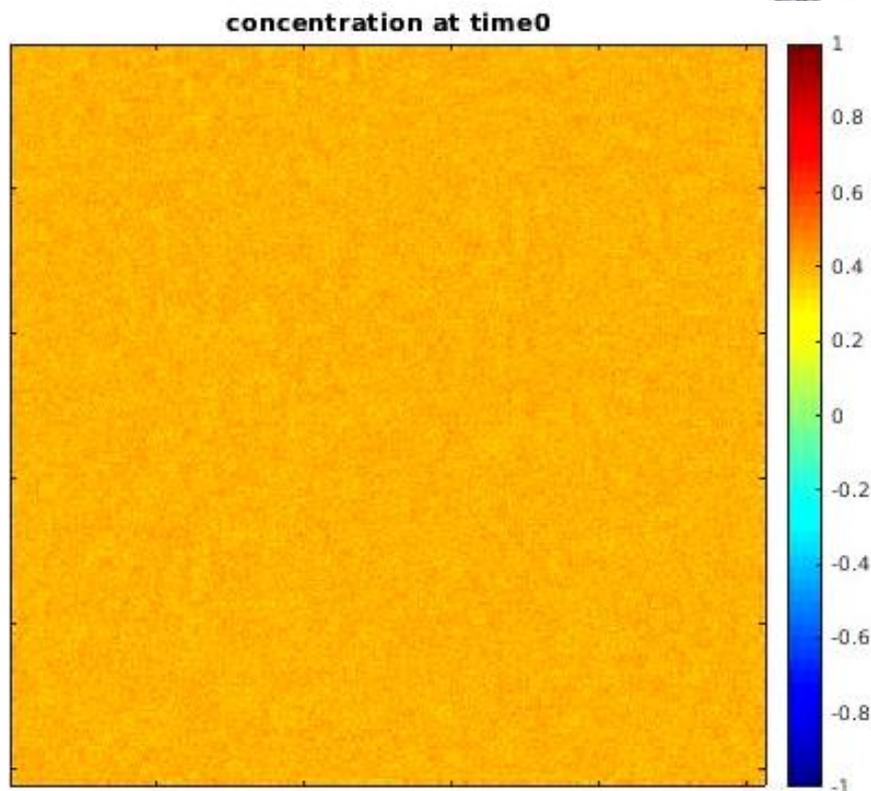


# Protein Dynamics upon Phase Separation

Protein Ig (“ $\gamma$ -globulin”), i.e. an antibody in water, with polyethylene glycol (PEG1000) to increase attractions through depletion

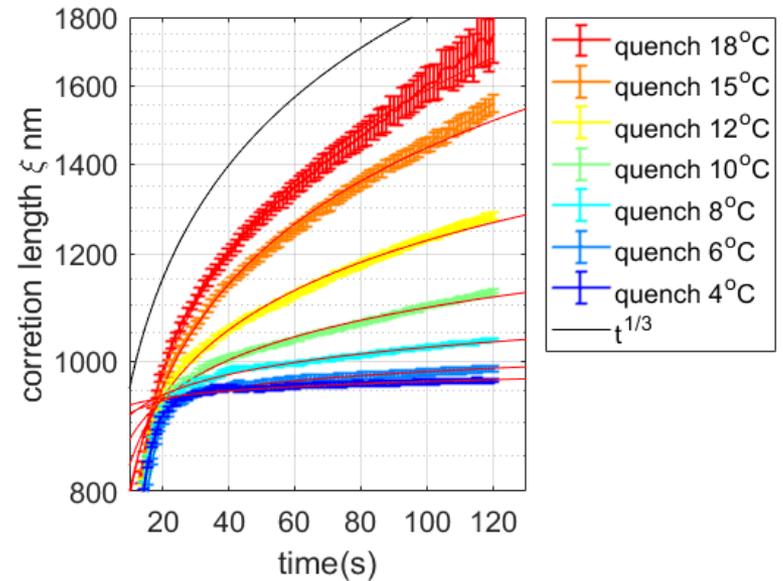
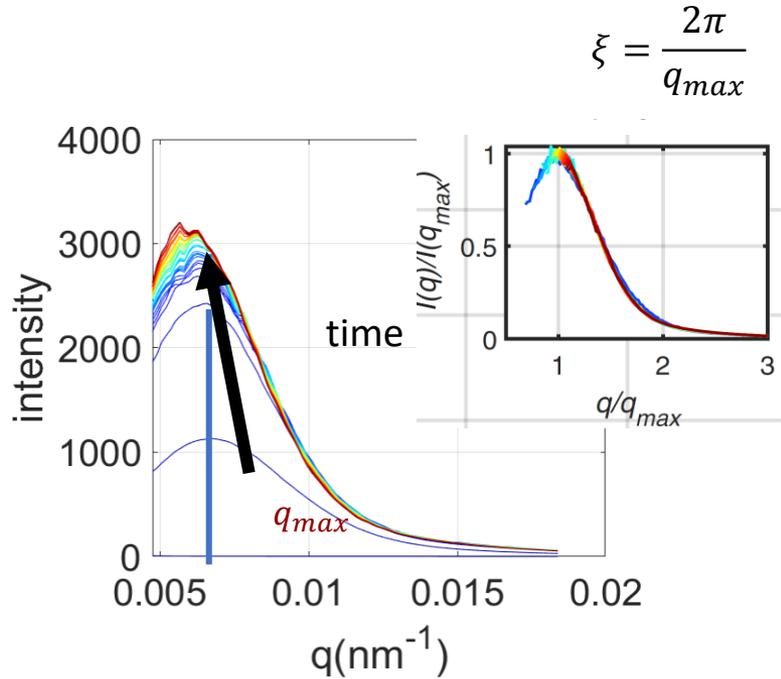


Simulation of a spinodal decomposition



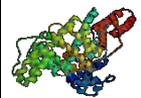
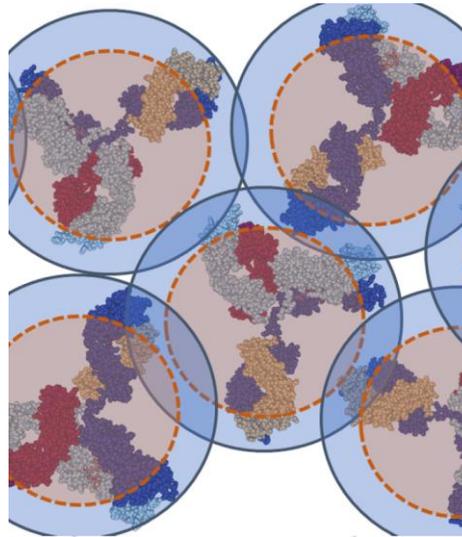
# Protein Dynamics upon Phase Separation

Kinetics of a spinodal decomposition (small angle scattering)



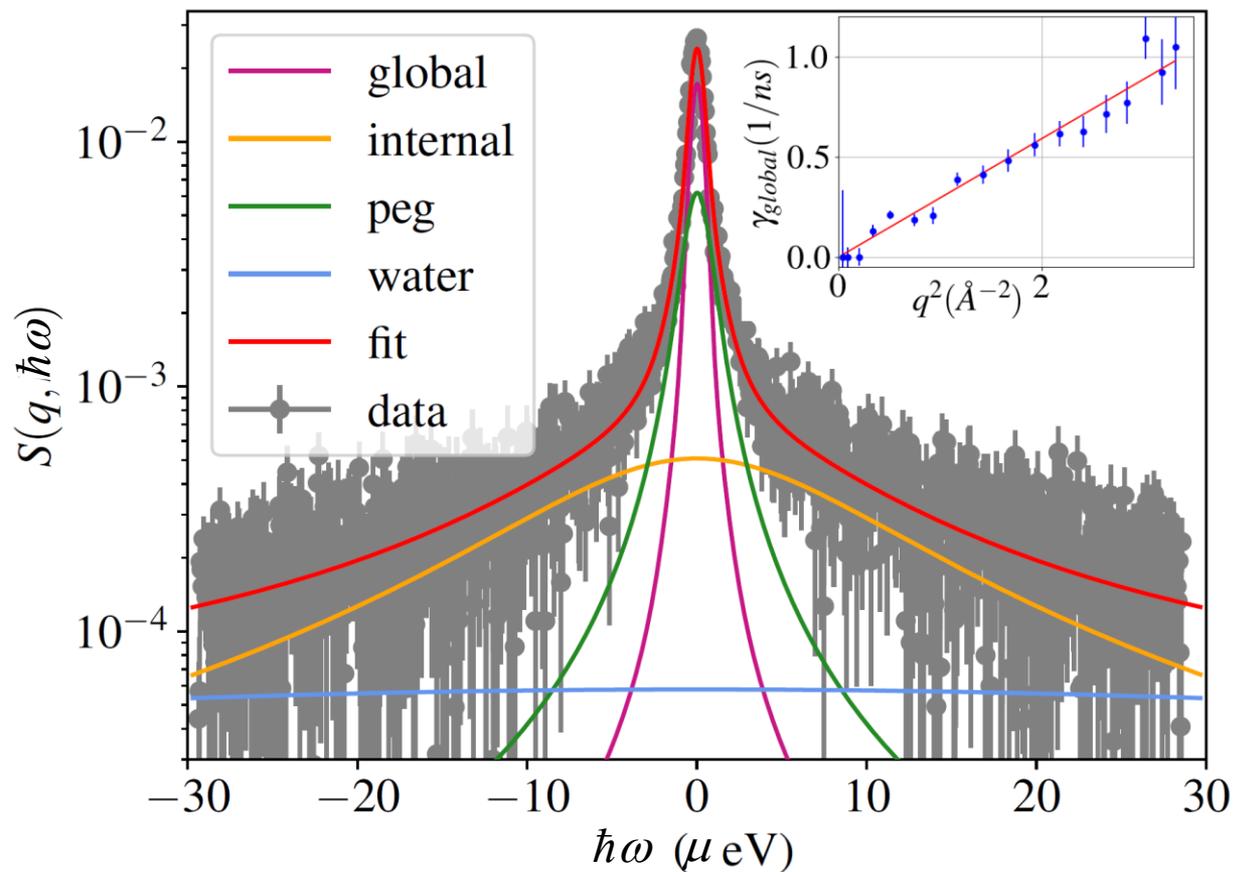
# Protein Dynamics upon Phase Separation

First focus on the dense phase



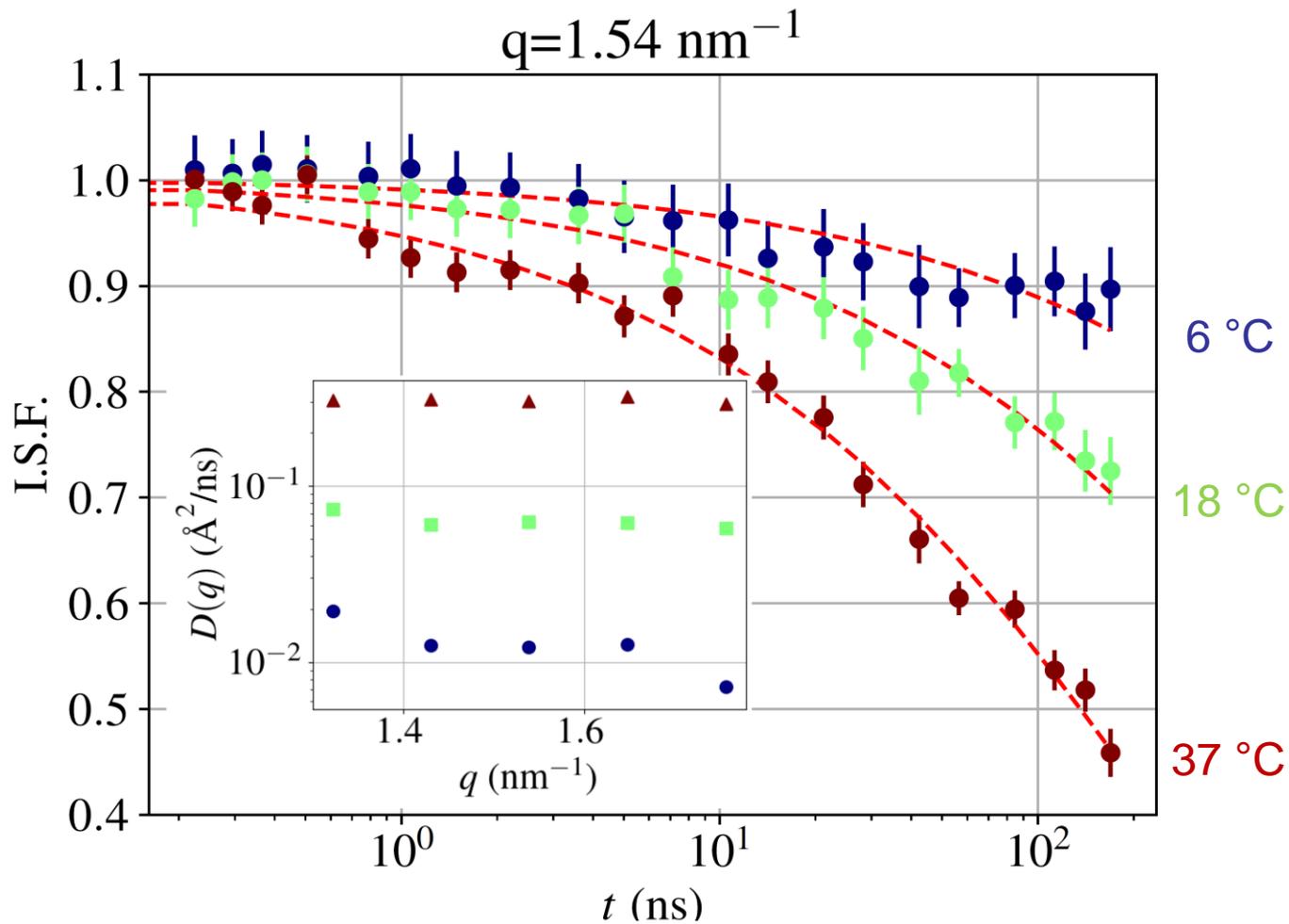
# Protein Dynamics upon Phase Separation

NBS (IN16B) to determine effective diffusion parameter in the dense phase



# Protein Dynamics upon Phase Separation

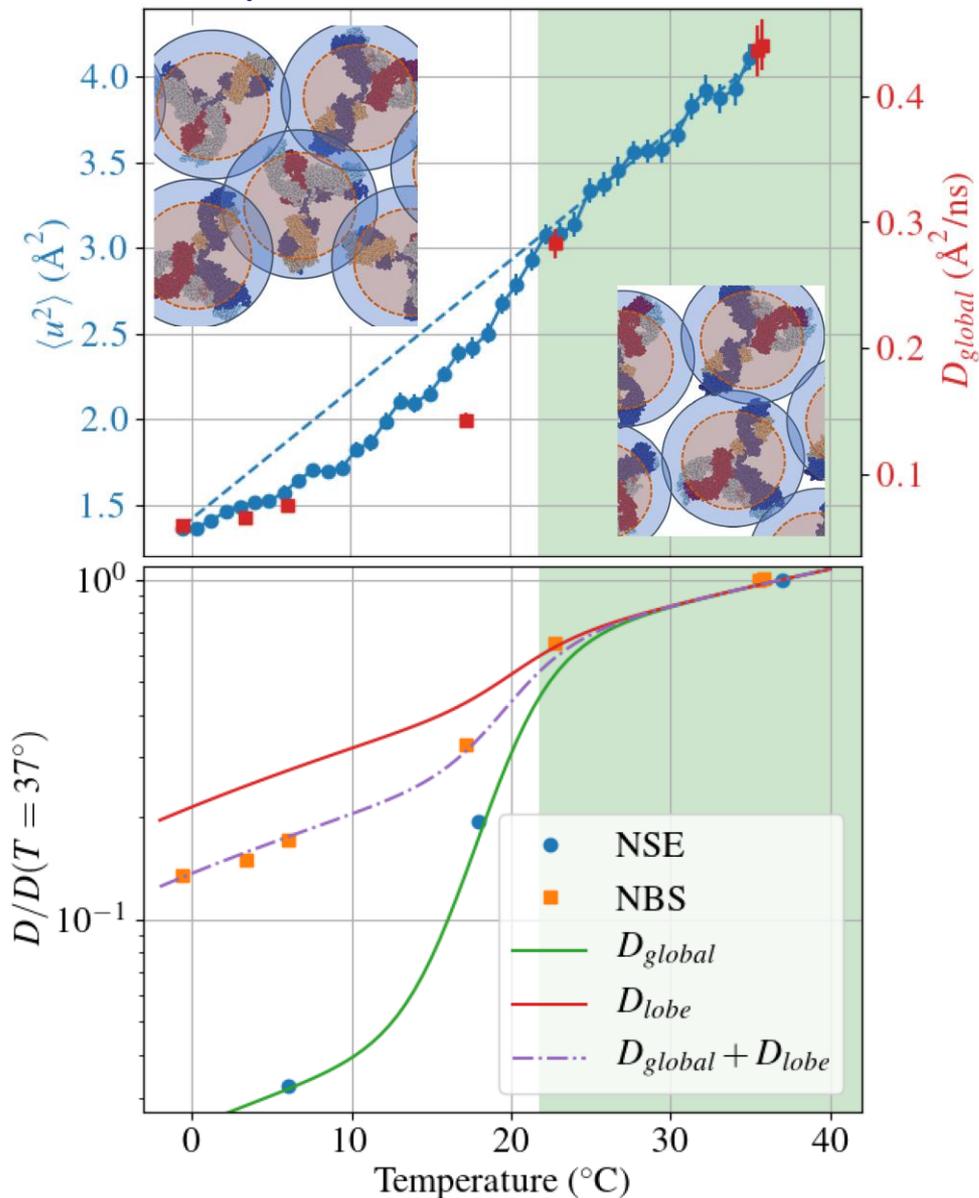
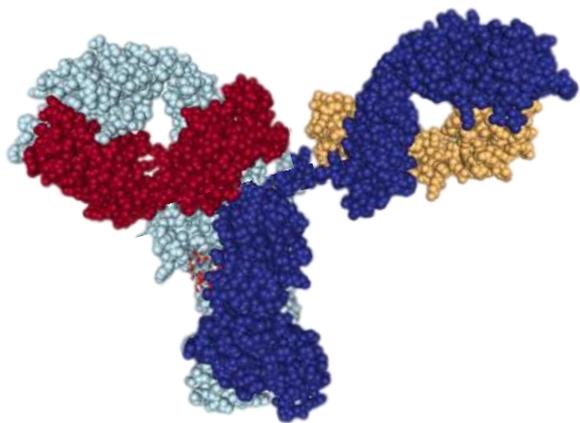
NSE (IN15) to determine effective diffusion parameter  
in the dense phase



# Protein Dynamics upon Phase Separation

From comparison NSE and NBS:

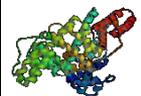
- $D$  slows down with  $T$
- difference NSE vs NBS suggests *local* motion preserved (different length scales; long for NSE, more local for NBS):
- i.e. antibody flexibility in principle preserved even under crowded conditions



# Bonus Track



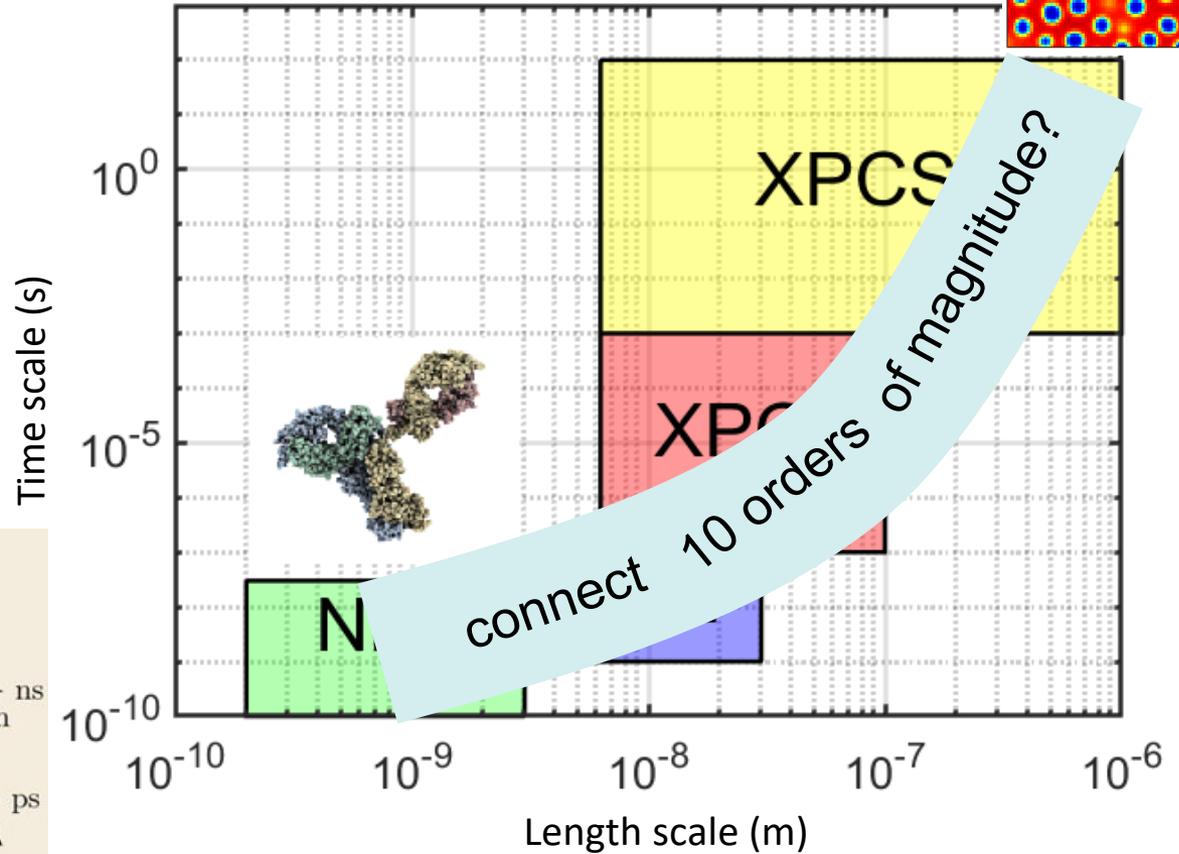
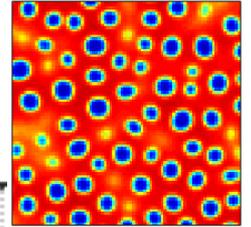
# Protein Dynamics: Time and Length Scales



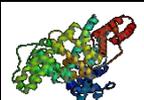
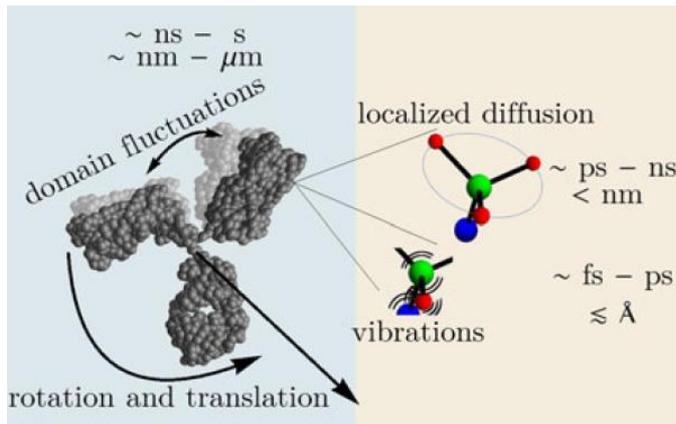
# Protein Dynamics: Time and Length Scales

Generally complex task  
many different time and length scales  
requiring advanced instrumentation

Dynamics of domains ( $\sim \mu\text{m}$ )



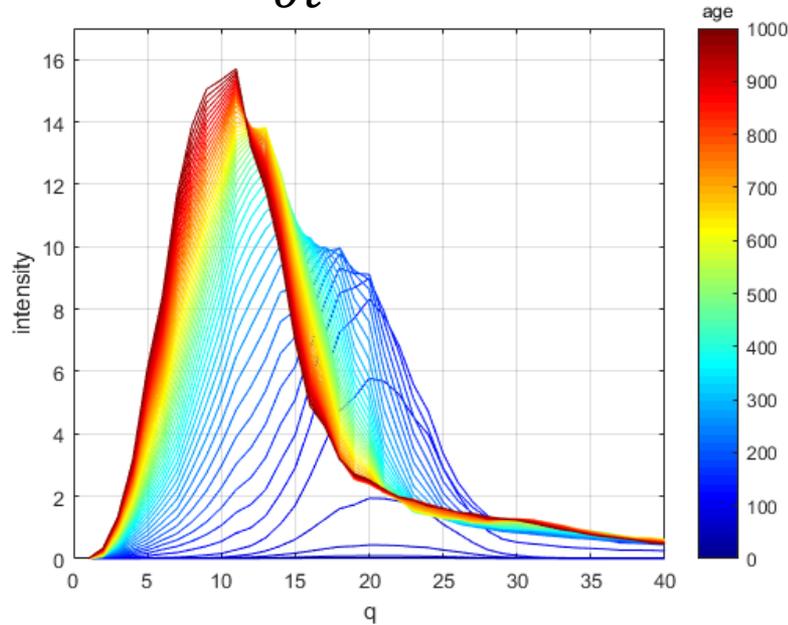
Dynamics at molecular level ( $\sim \text{nm}$ )



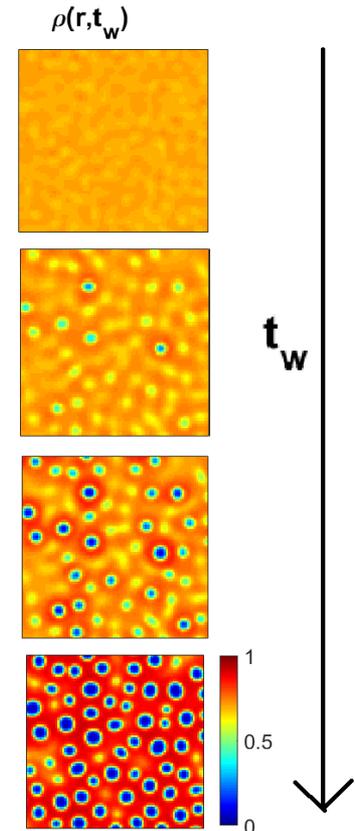
# Protein Dynamics upon Phase Separation

Simulations based on Cahn-Hilliard equation

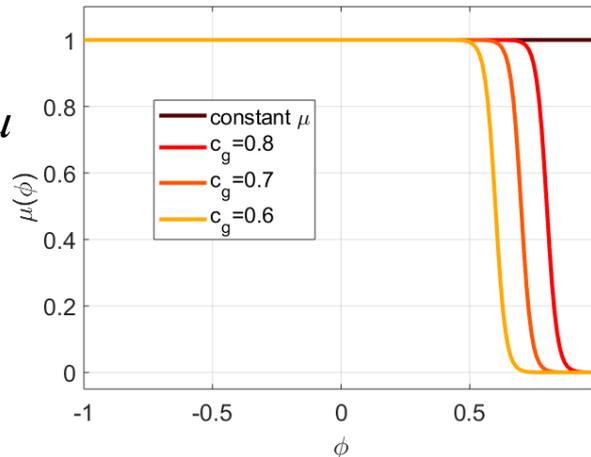
$$\frac{\partial \psi}{\partial t} = \nabla[\mu \nabla(\epsilon \psi - \psi^3 - \nabla^2 \psi)]$$



FT



Mobility parameter  $\mu$  depends on concentration  $\phi$

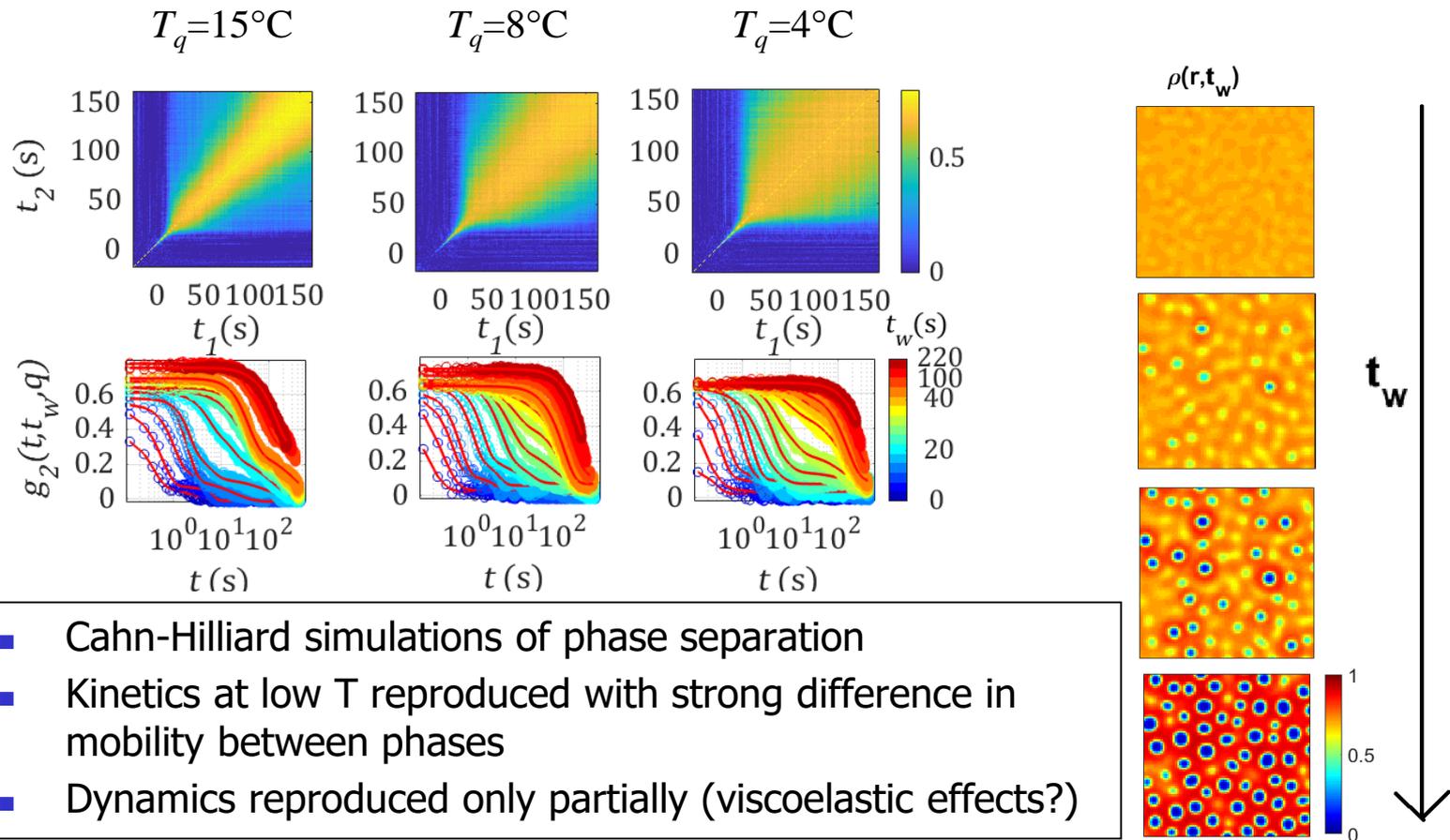


Anita Girelli et al., PRL 2021



# Protein Dynamics upon Phase Separation

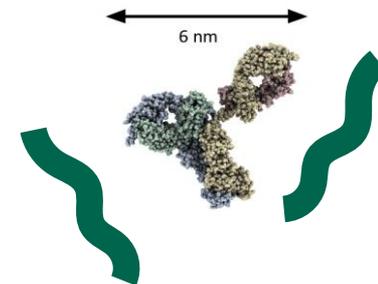
XPCS data @  $q = 0.005\text{nm}^{-1}$



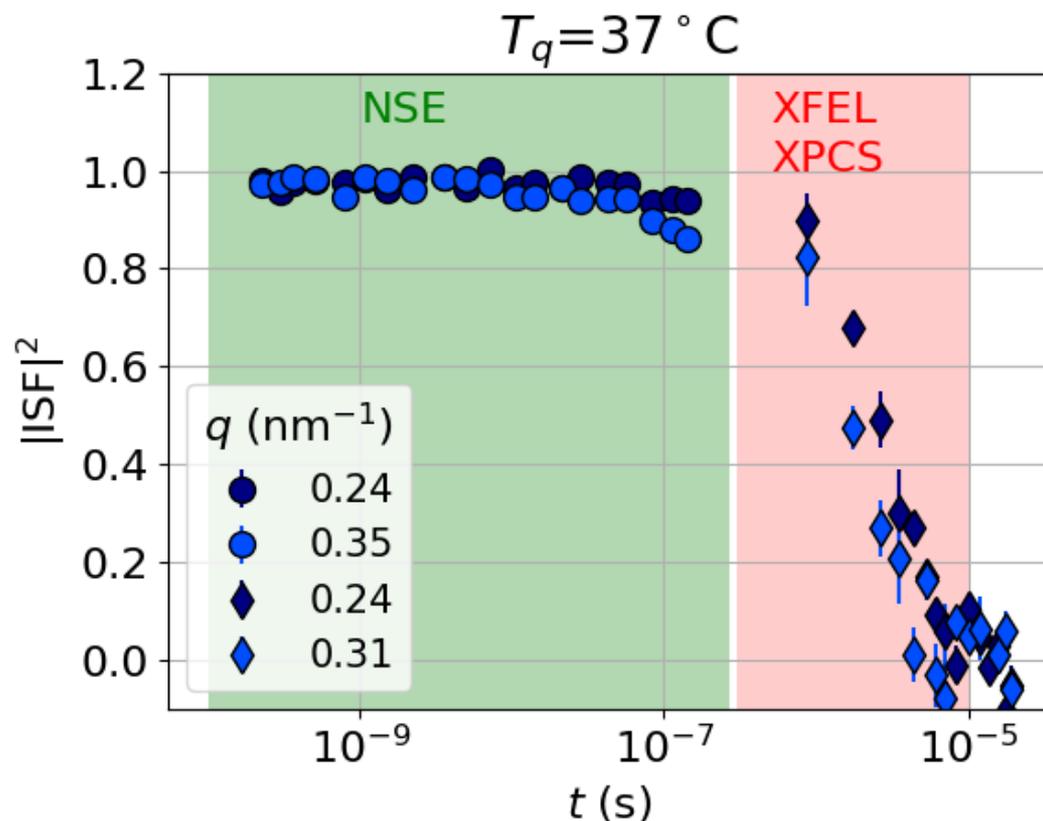
# Bonus Track



# Bonus Track: Connecting NSE with XPCS



now in one-phase region at high  $T$   
for the same  $q$  for XPCS and NSE!



Connecting NSE with XPCS works

- Dynamics resolved with NSE and XFEL-XPCS
- Consistent with Brownian motion



# Conclusions

- For a holistic understanding of complex protein solutions, study structure *and* dynamics
- Dependence on environment and conditions
- Specific example of IgG/PEG
  - Antibody flexibility preserved despite environment
  - Difference in NBS and NSE
  - Short-time self-diffusion not strongly affected by arrest of the phase separation
  - Phase separation described by Cahn-Hilliard with difference in mobility of the two phases
  - Phase separation kinetics and dynamics followable “over 10 orders of magnitude” ?
- For complete picture, need more than one technique, e.g. SANS / NBS / NSE / XPCS / rheo etc.
- SANS & QENS
- Need NSE @ ESS !

