

Dynamic cluster formation, viscosity and diffusion in monoclonal antibody solutions

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1. Introduction

- Monoclonal antibodies (mAbs): structure and functionality
- Pharmaceutical challenges

2. Materials and methods

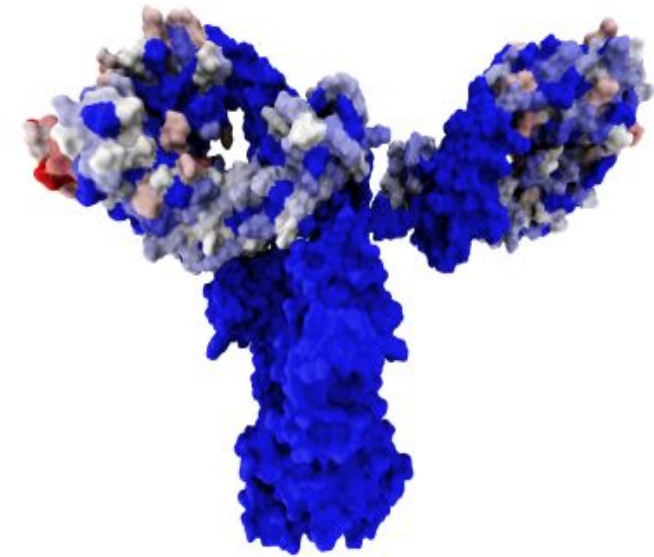
- Lonza mAbs
- Neutron backscattering spectroscopy
- Small angle neutron scattering (SANS)
- Molecular dynamics (MD) simulations

3. Data treatment and analysis

4. First results

- QENS
- SANS

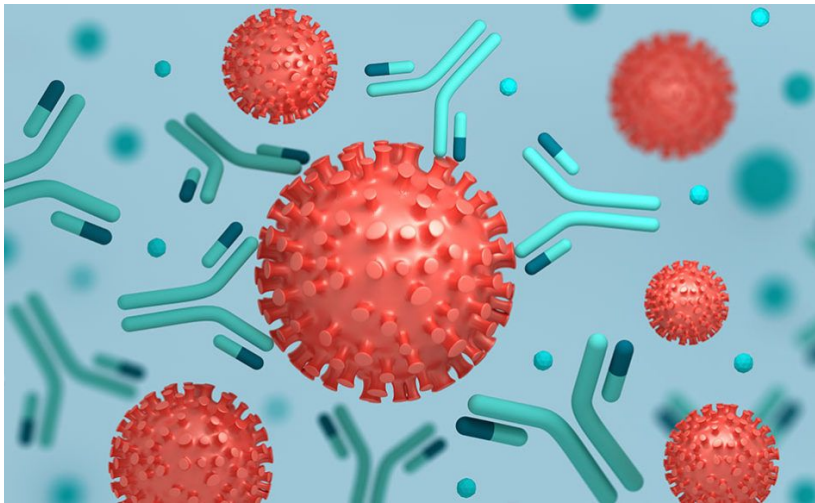
5. Conclusions and future developments



1. Introduction: Monoclonal antibodies (mAbs)

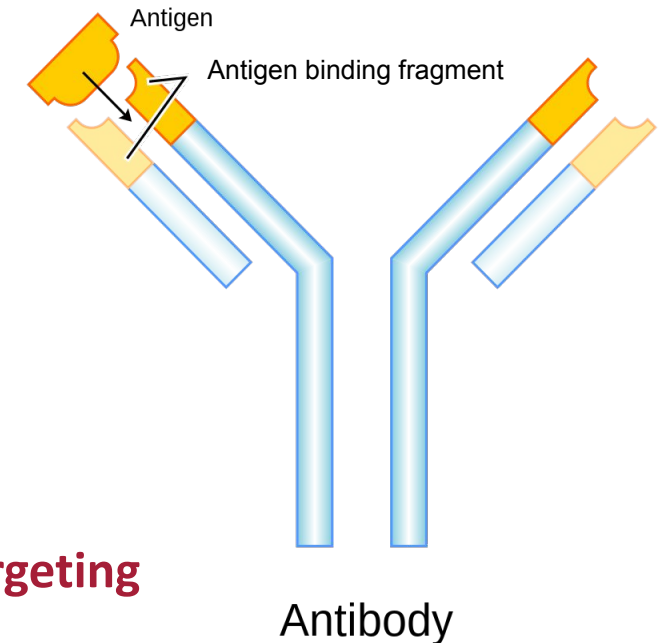
Antibodies: proteins, aka *immunoglobulins* (IgGs), secreted in response to pathogens

- 4 polypeptides, 150 kDa Y-shaped molecule
- Fab = antigen binding fragment → bivalent antigen binding



Monoclonal antibodies recognize only a single type of antigen!

- high **specificity** and **versatility**
- efficiency in **pharmaceutical targeting**



(1) Lipman *et al.*, ILAR Journal, vol. 46, 3, 258-268 (2005) (2) Johnson *et al.*, Mater Methods, 3:160 (2013)

2



1. Introduction: Pharmaceutical challenges

MAB solutions are employed in therapeutics for multiple diseases: cancers, psoriasis, viral infections, ...

Currently:

administration via *intravenous (IV) injection*



high administration frequency
skilled personnel required
less patient convenience

Market need:

administration via *subcutaneous (SC) injection*

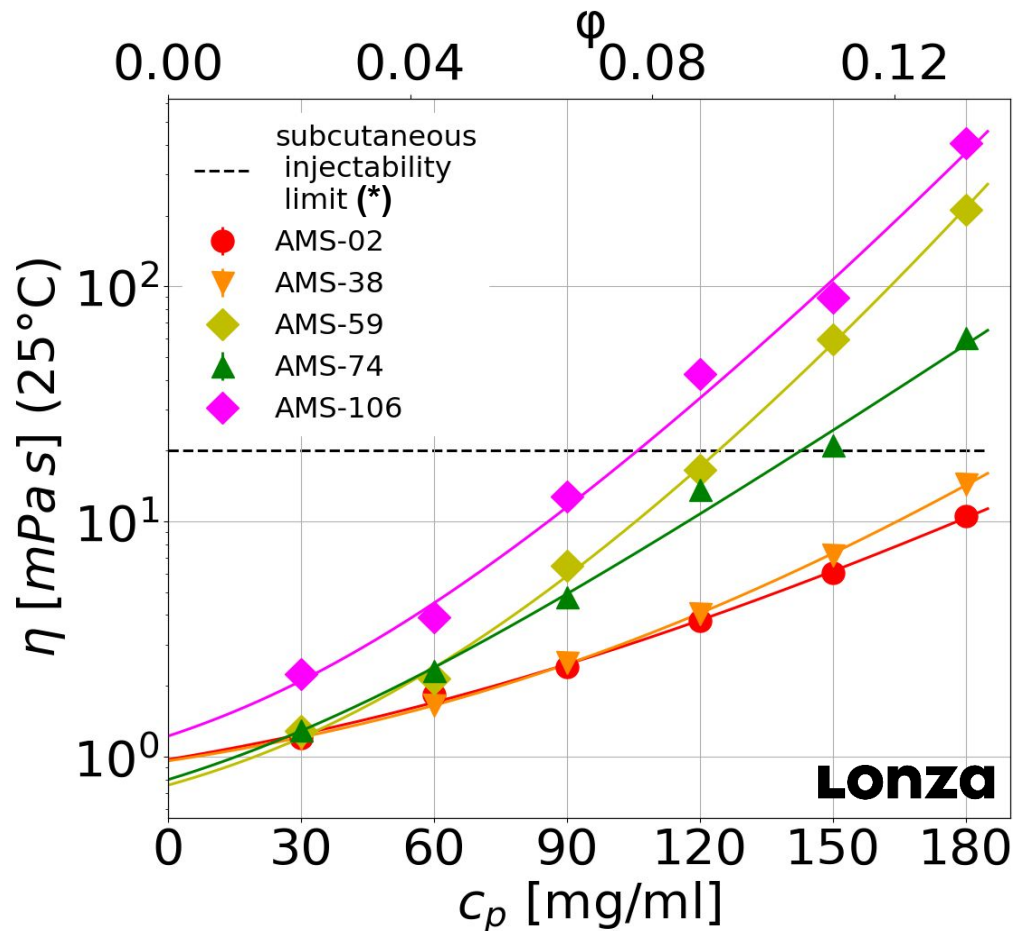
low administration frequency
no skilled personnel required
more patient convenience



SC formulations:

- Low volumes (1-2 mL)
- High concentrations → high viscosity → difficulty to inject
- Viscosity reduction is possible via additives

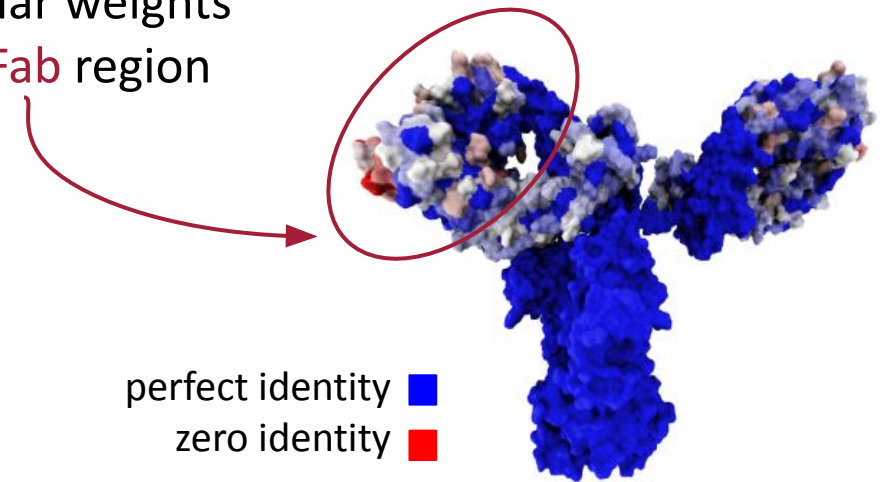
2. Materials and methods: Lonza mAbs



mAb type-dependent viscosity η

Similar molecular weights

Differences in Fab region



→ differences in surface charge and protein-protein interactions

→ aggregation / reversible self-association ^(1,2)

(*) Jolles S., Sleasman J. W., Adv. Ther. 28, 521 (2011) (1) Liu, J., et al., J. Pharm. Sci. 94:1928–40 (2005) (2) Yearley E. et al., Biophys. J. 140:1763-70 (2014)

2. Materials and methods

Neutron backscattering spectroscopy
+ small angle neutron scattering (SANS)

D11 @ILL:

three Lonza mAbs in D2O solution
at $c_p = 80 \text{ mg/mL}$, $T = 18, 21, 37^\circ \text{C}$



IN16B @ILL:

five Lonza mAbs + polyclonal IgG (reference) in
D2O solution, 20mM His-HCl buffer
at different c_p and T (280, 295, 310 K)



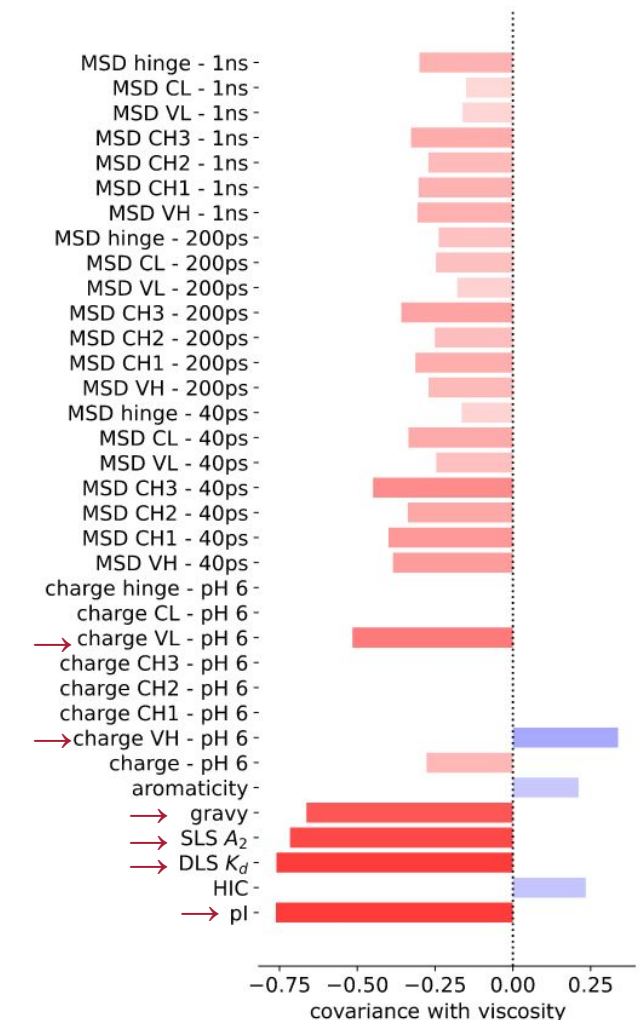
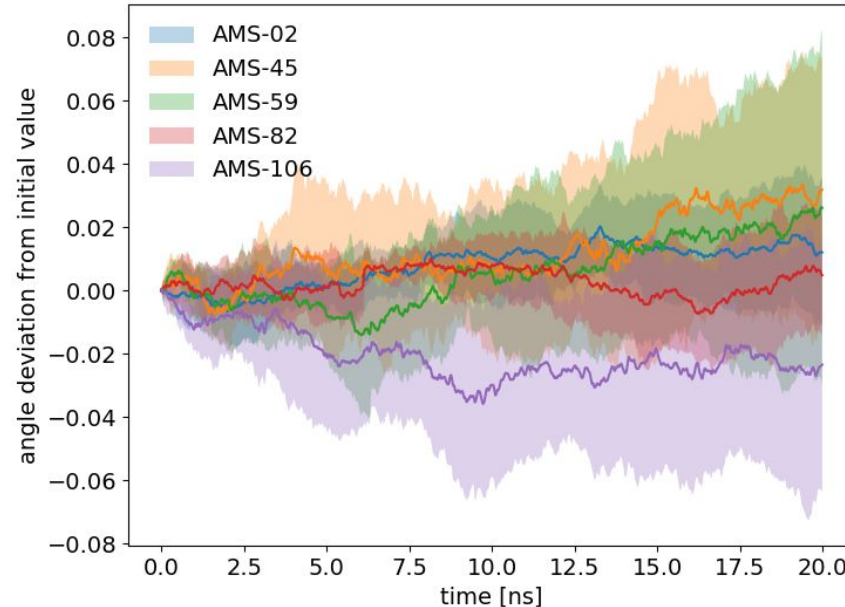
<https://www.ill.eu/users/instruments/instruments-list/in16b, d11>

2. Materials and methods

MD simulations (*K. Pounot*)

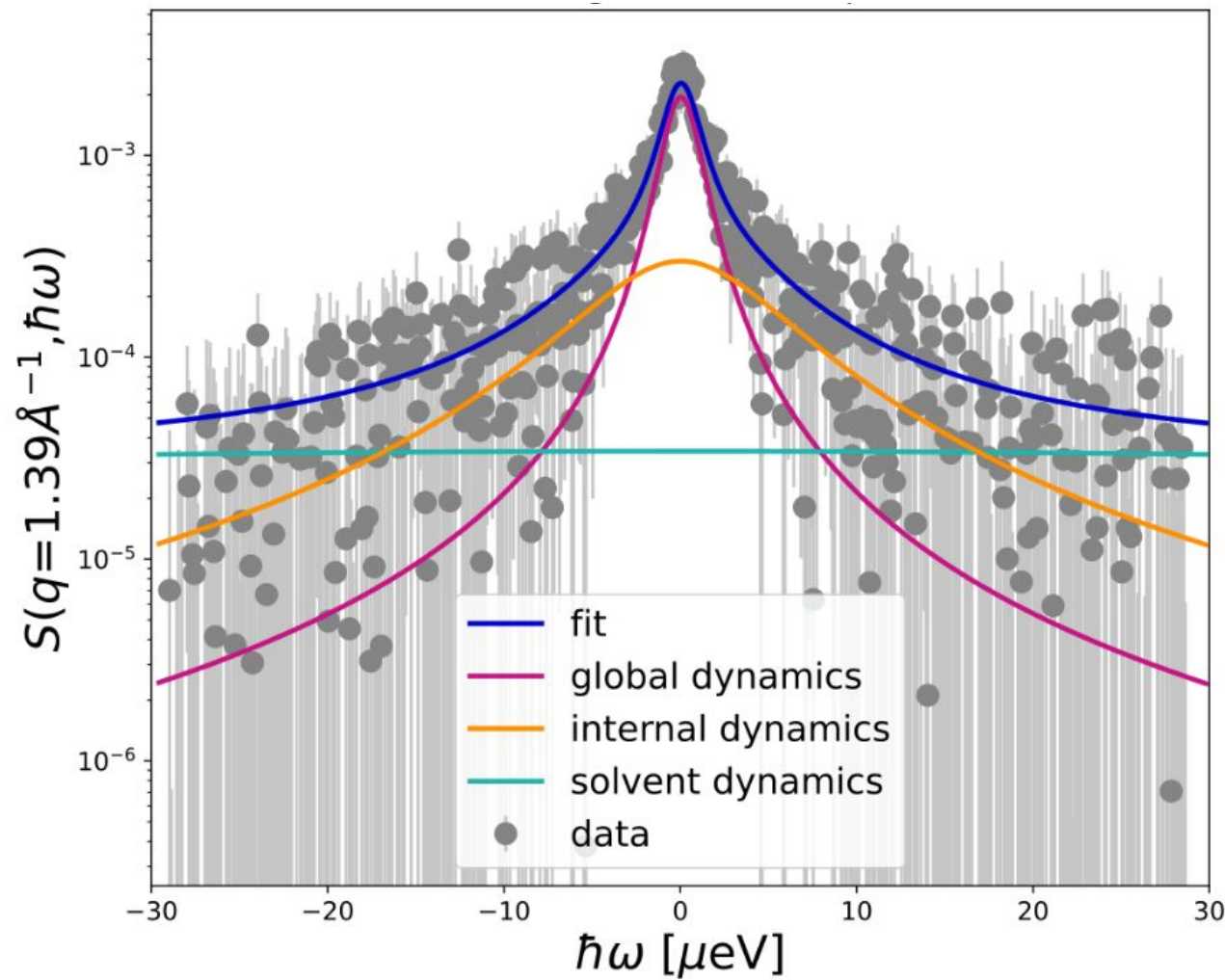
- Single-molecule simulations via *NAMD* using CHARMM36 force field ^(1,2)
- Calculation of MSDs, angles between the lobes, internal diffusion
- Determination of the charge of different domains (*pdb2pqr*) ⁽³⁾
- Calculation of isoelectric point, hydrophobicity, aromaticity via *BioPython*

→ internal dynamics does not strongly depend on mAb type
→ V domains charge and PPI parameters correlate with viscosity



(1) Huang J. *et al.*, J. Comput. Chem., 34:2135-45 (2013) (2) Huang J., *et al.*, Nature Methods, 14, 1:71-73 (2017) (3) Jurrus E. *et al.*, Prot. Sci., 27:112-128 (2018) 6

3. Data treatment and analysis



Sample: mAb **AMS-59**, by **Lonza**

Buffer: 20mM His-HCl in D2O

$c_p = 105$ mg/mL

$T = 280$ K

3. Data treatment and analysis

Data reduction using Mantid, empty cell subtraction
 Fitting and analysis using Python (q -wise and global fits)

$$S(q, \omega) = R(q, \omega) \otimes \left\{ \beta \left[\underbrace{A_0(q) \mathcal{L}_{\Gamma_{glob}}}_{\text{purple}} + \underbrace{(1 - A_0(q)) \mathcal{L}_{\Gamma_{int}}}_{\text{orange}} \right] + \underbrace{\mathcal{L}_{\Gamma_{D_2O}}}_{\text{cyan}} \right\}$$

1. Fickian global + free internal dynamics + parametrized EISF:

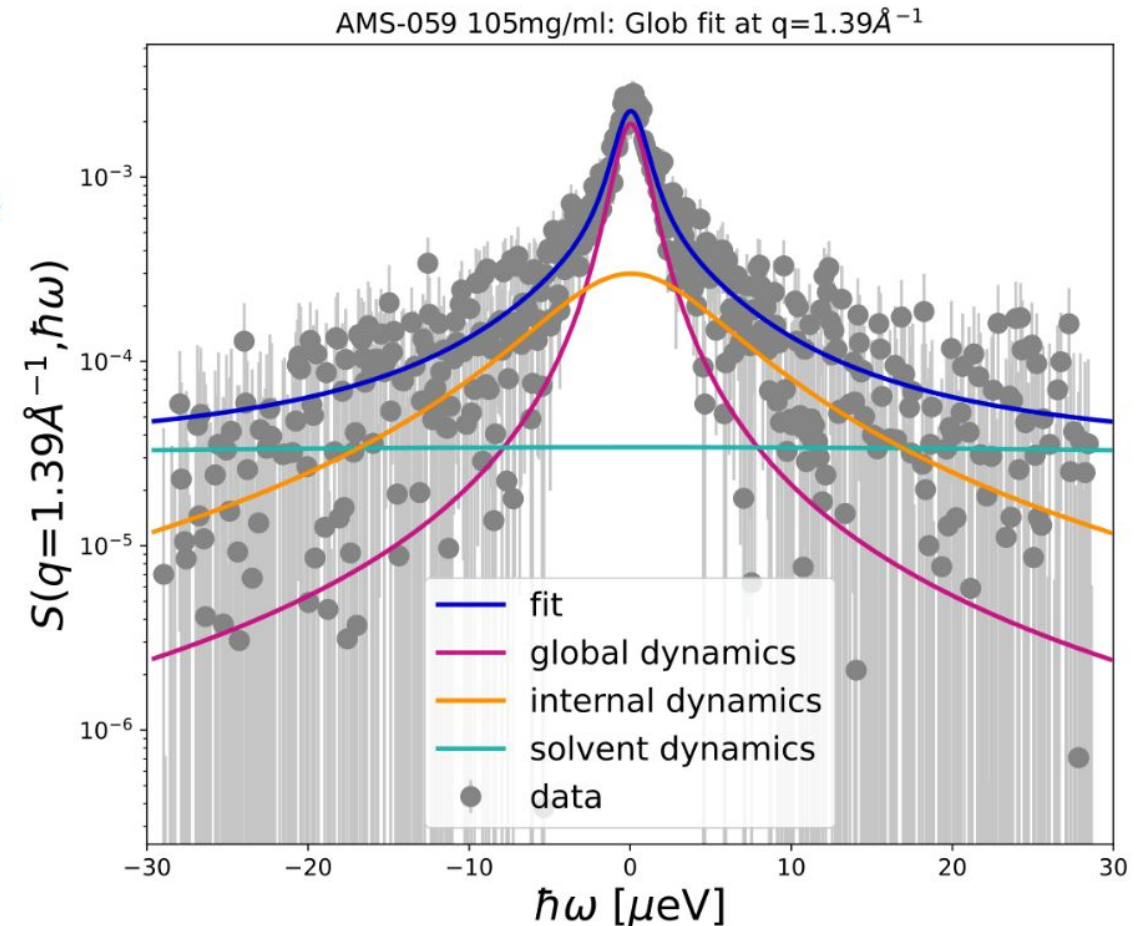
$$\Gamma_{glob} = D q^2$$

$$A_0(q) = p + (1 - p) \left[p_1 A_{3jump}(q) + (1 - p_1) A_{sphere}(q) \right]$$

$$A_{3-jump}(q) = \frac{1}{3} [1 + 2j_0(qa)] \quad A_{sphere}(q) = \left| \frac{3j_1(qR)}{qR} \right|^2$$

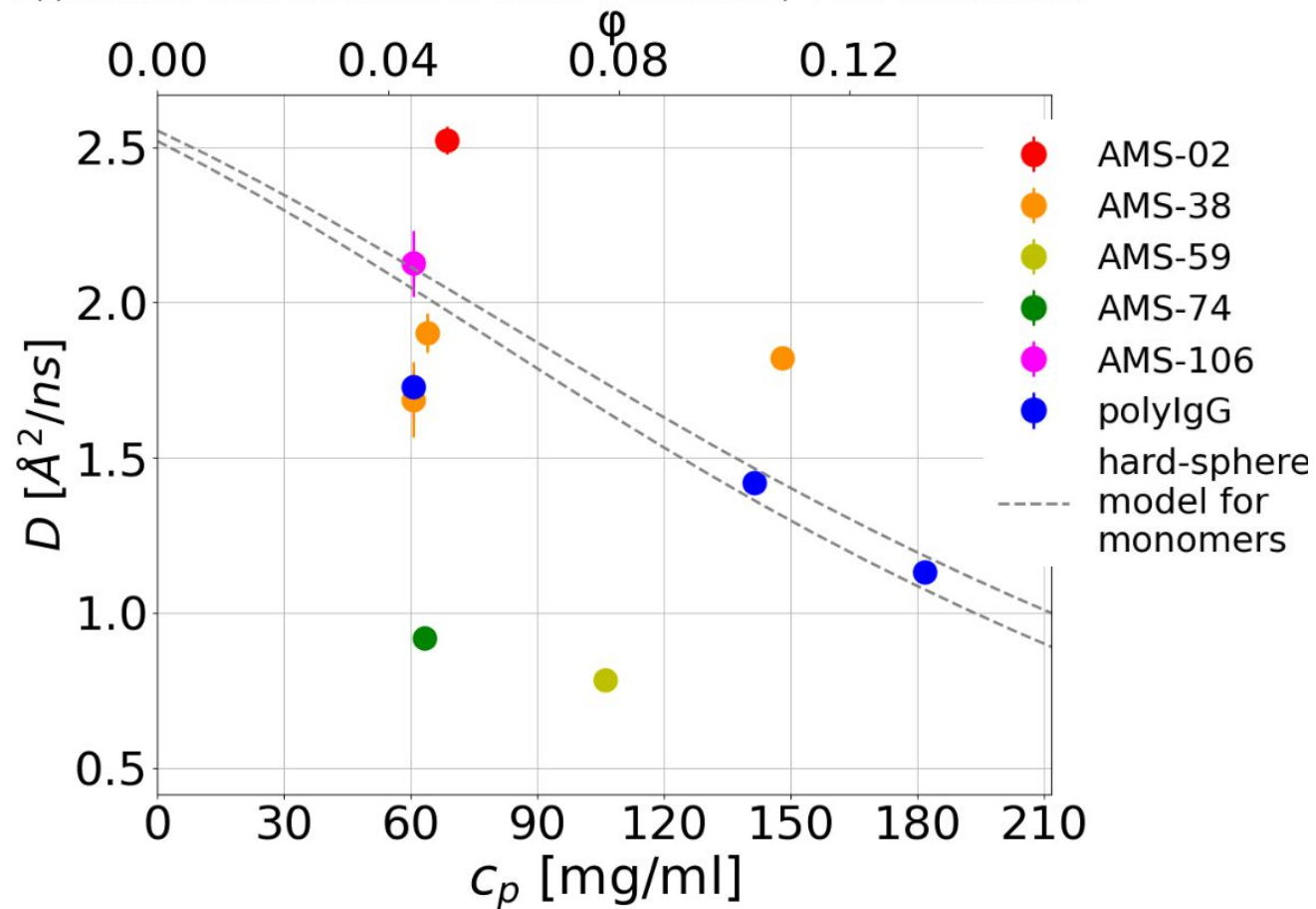
2. Fickian global + jump-like internal dynamics:

$$\Gamma_{glob} = D q^2 \quad \Gamma_{int} = \frac{D_1 q^2}{1 + \tau D_1 q^2}$$



4. First results: QENS

Center of mass self-diffusion (at 280 K)



Theoretical prediction of short-time diffusion coeff. $D = D(Dt(\varphi), Dr(\varphi))$ using hard-sphere model from colloid physics:

$$D_t(\varphi) = \frac{D_{t0}}{1 + L(\varphi)} \quad (1)$$

$$D_r(\varphi) = D_{r0}(1 - 1.3\varphi^2) \quad (2)$$

Effective volume fraction φ_{eff}

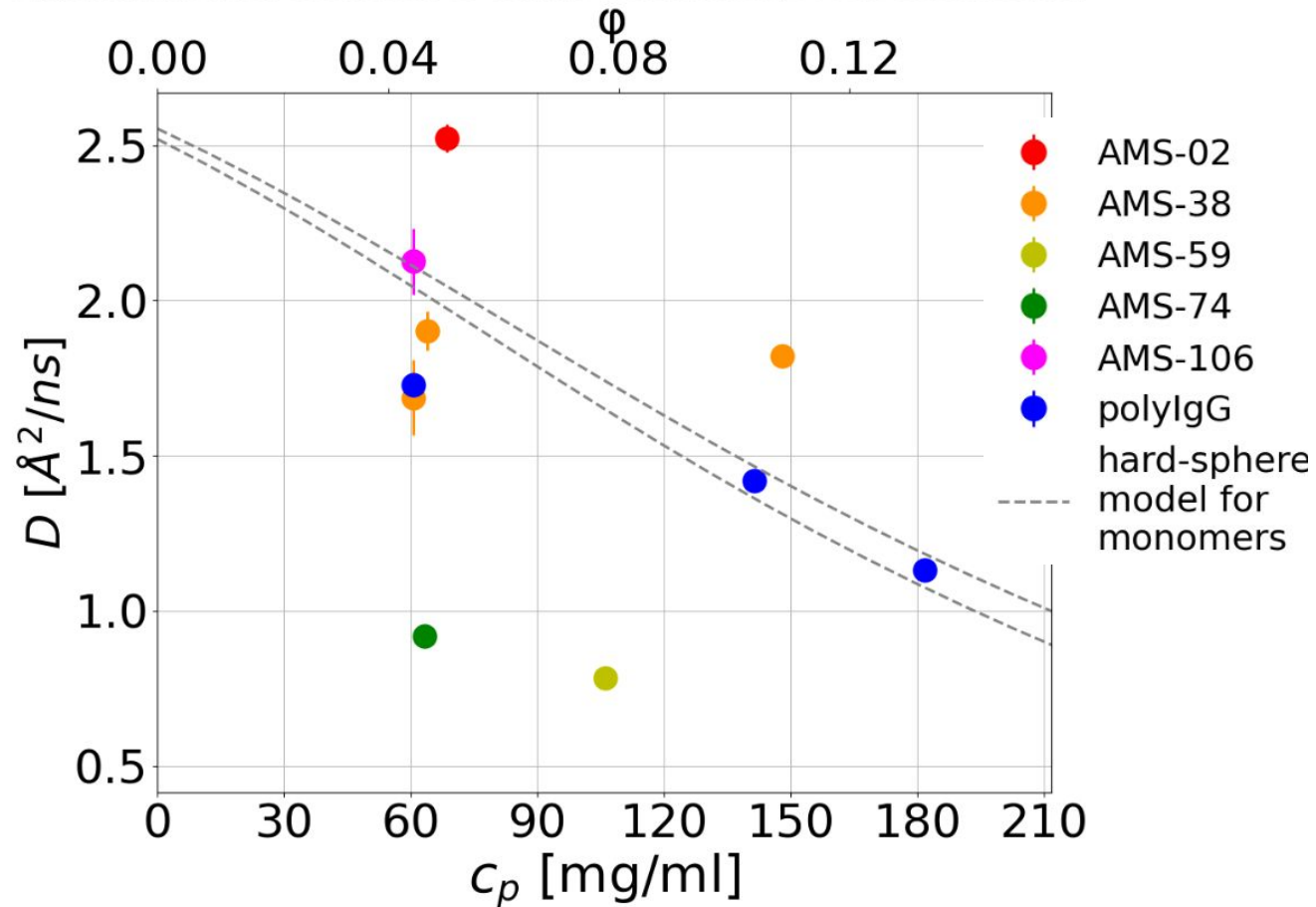
$$\varphi_{\text{eff}} = \varphi \left(\frac{R_h}{R_{\text{dry}}}\right)^3 \quad (3)$$

$D_{t0}, D_{r0}, R_{\text{dry}}$ obtained via HYDROPRO10 ⁽⁴⁾

- (1) Tokuyama M, Oppenheim I., Physica A 216, 85-119 (1995)
- (2) Banchio A., Nägele G., J. Chem. Phys. 128, 104903 (2008)
- (3) Roosen-Runge F., *et al.*, PNAS 108, 29, 11815-11820 (2011)
- (4) Ortega A., *et al.*, Biophys. J. 101, 892-898 (2011)

4. First results: QENS

Center of mass self-diffusion (at 280 K)



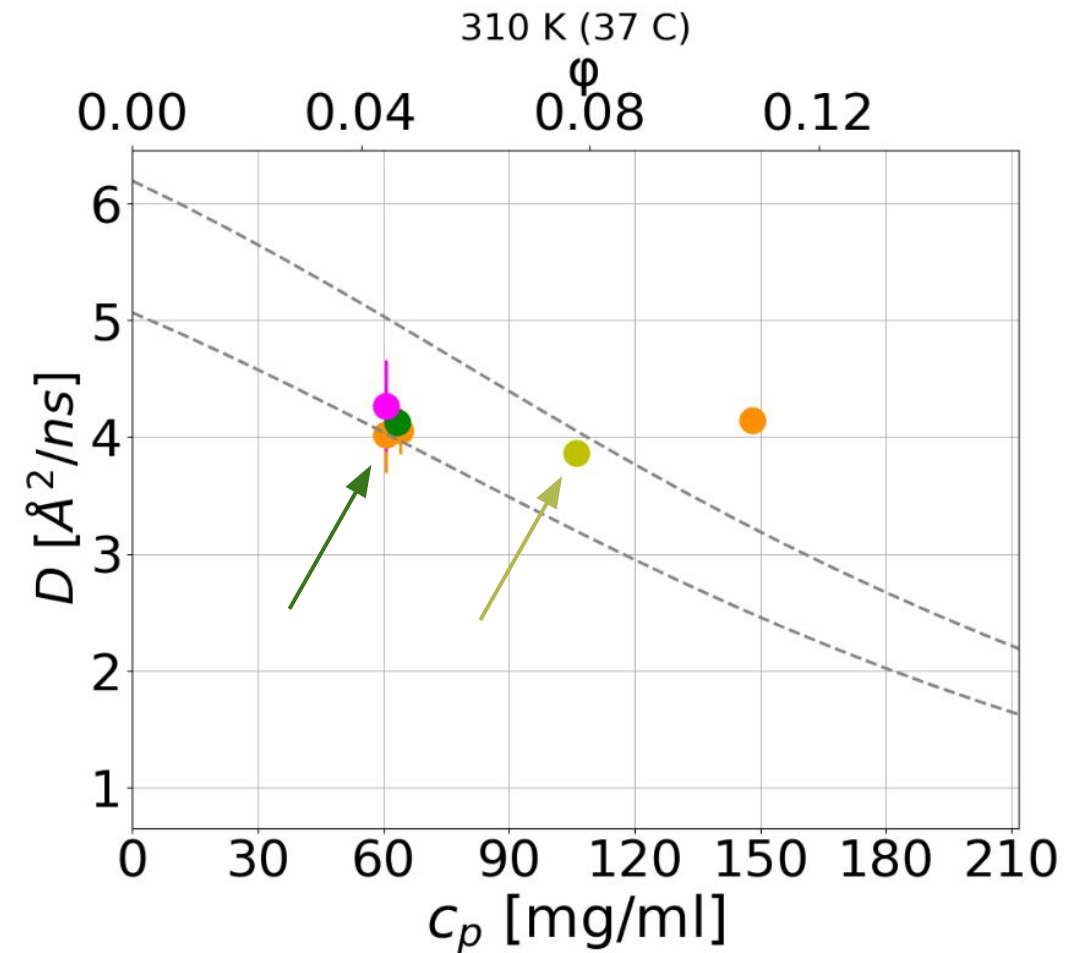
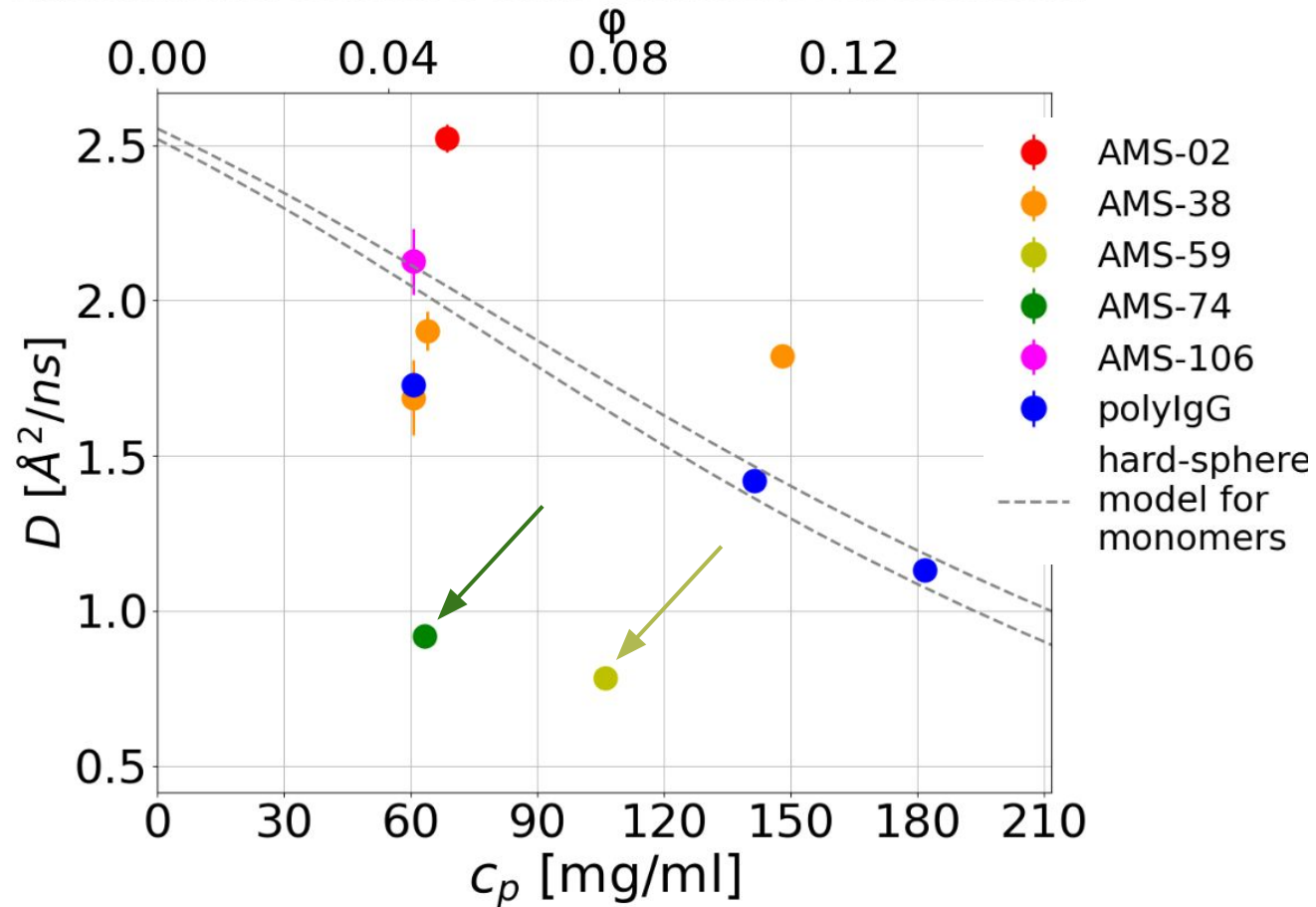
- Points on the lines: monomers
- Points below: clusters
- Points above: possibly heterogeneous samples or phase separation

Possibilities:

1. Incomplete model for heterogeneous situations
2. Cross-talking between global and internal motions
3. Discrepancies between nominal and real protein concentration

4. First results: QENS

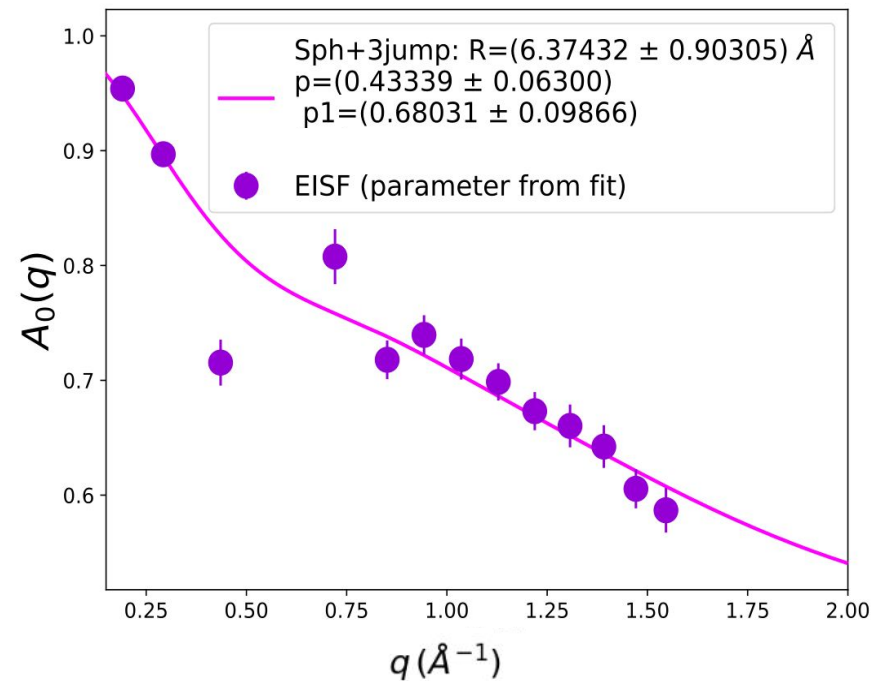
Center of mass diffusion (280K and 310K)



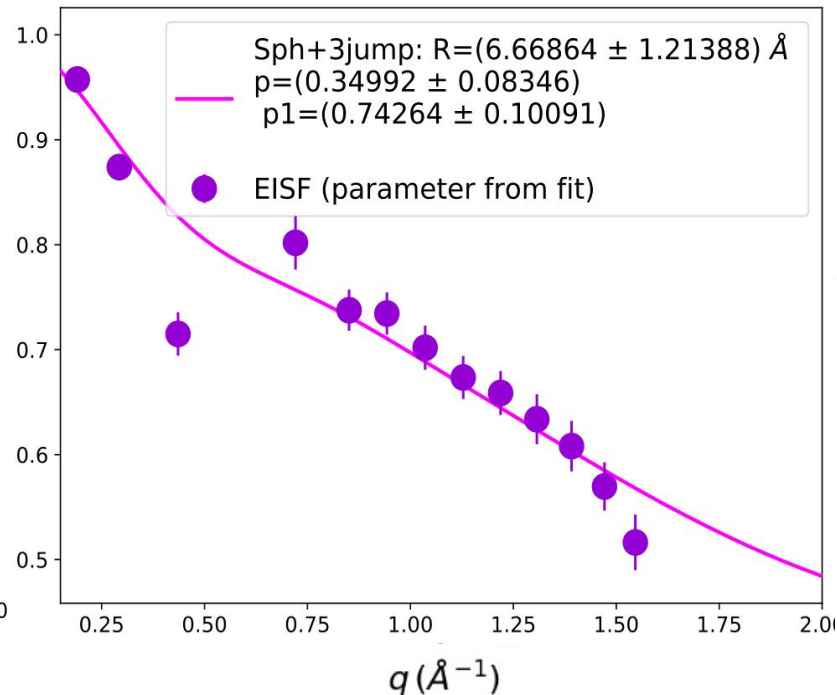
4. First results: QENS

Fitted elastic incoherent structure factor (EISF) $A_0(q)$

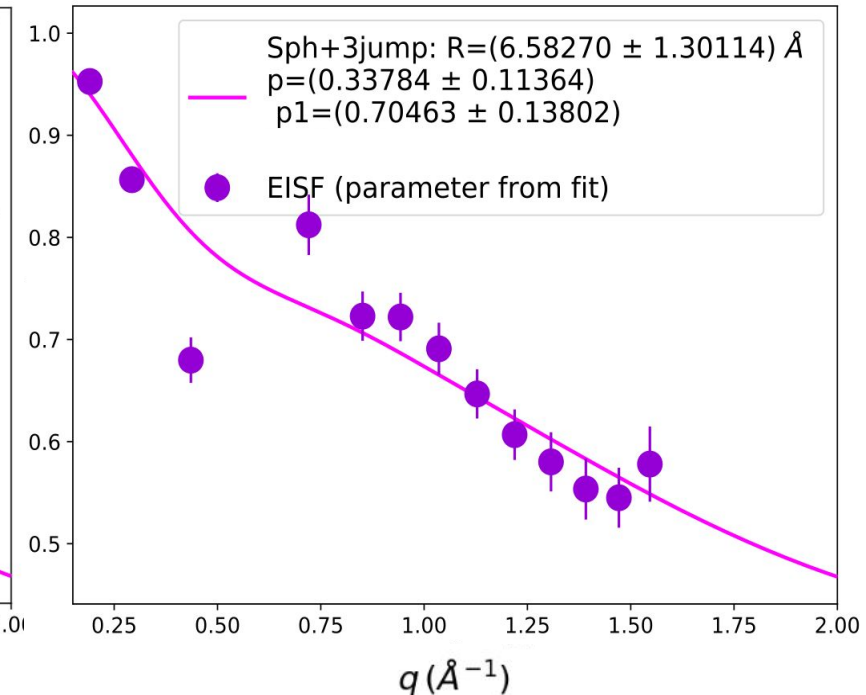
AMS-38 146 mg/mL, 280 K



295 K



310 K



4. First results: QENS

Parameters R , p , p_1 from the EISF

$$A_0(q) = p + (1 - p) \left[p_1 A_{3\text{jump}}(q) + (1 - p_1) A_{\text{sphere}}(q) \right]$$

$$A_{3\text{-jump}}(q) = \frac{1}{3} [1 + 2j_0(qa)] \quad A_{\text{sphere}}(q) = \left| \frac{3j_1(qR)}{qR} \right|^2$$

$a \approx 1.7155 \text{ \AA}$

p independent from c_p

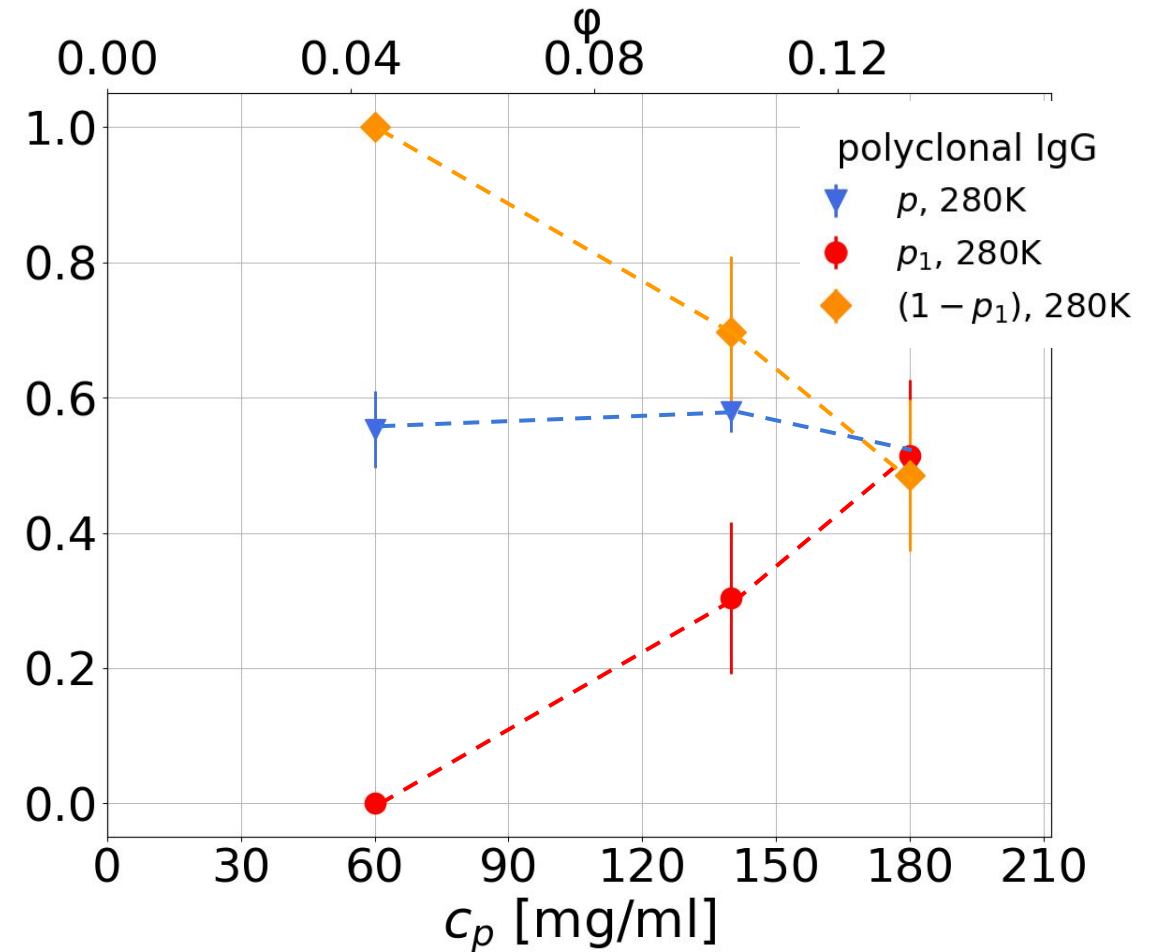
p_1 increases with increasing c_p

$(1-p_1)$ decreases at increasing c_p ⁽¹⁾

R in agreement with previous work on γ -globulin ^(1,2)

(1) Grimaldo M. *et al.*, J. Phys. Chem. B 2014, 118, 7203–7209

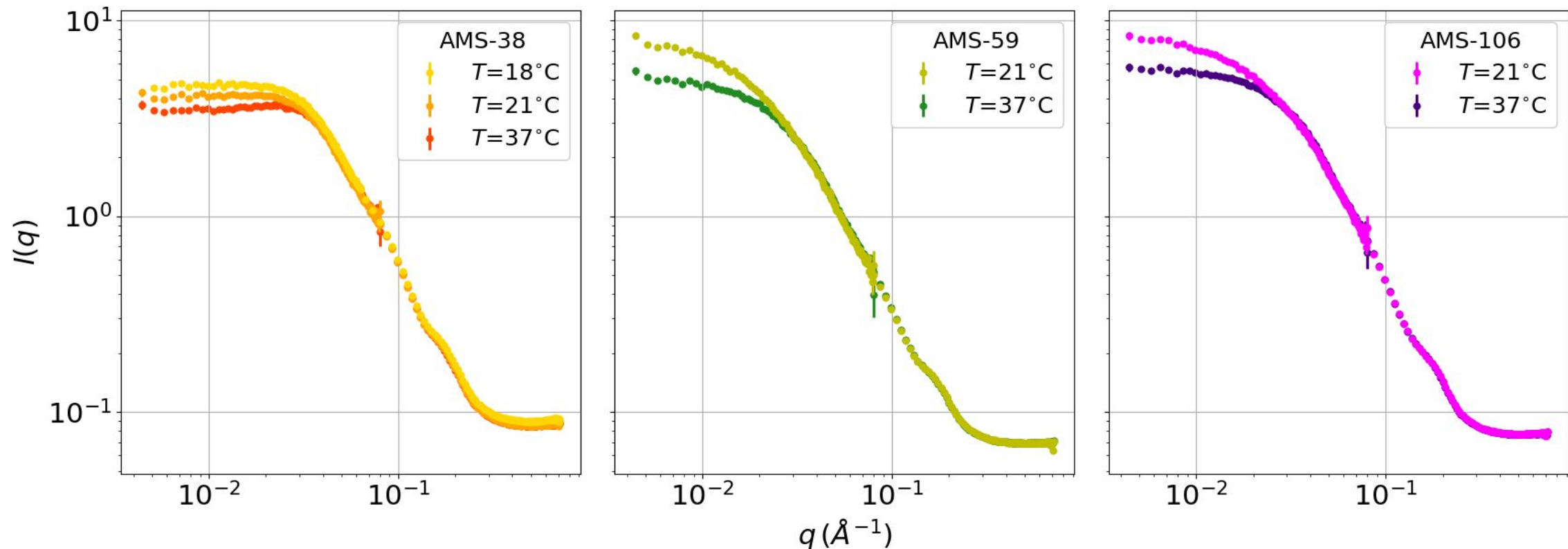
(2) Stagg L. *et al.*, PNAS USA 104:18976–18981 (2007)



4. First results: SANS

SANS curves, temperature dependence

Subset of mAbs: AMS-38, AMS-59, AMS-106; $c_p = 80$ mg/ml, buffered in 20mM His-HCl in D_2O



Rosenbaum, D., *et al.*, Phys. Rev. Lett. 76, 1, 150 (1996)

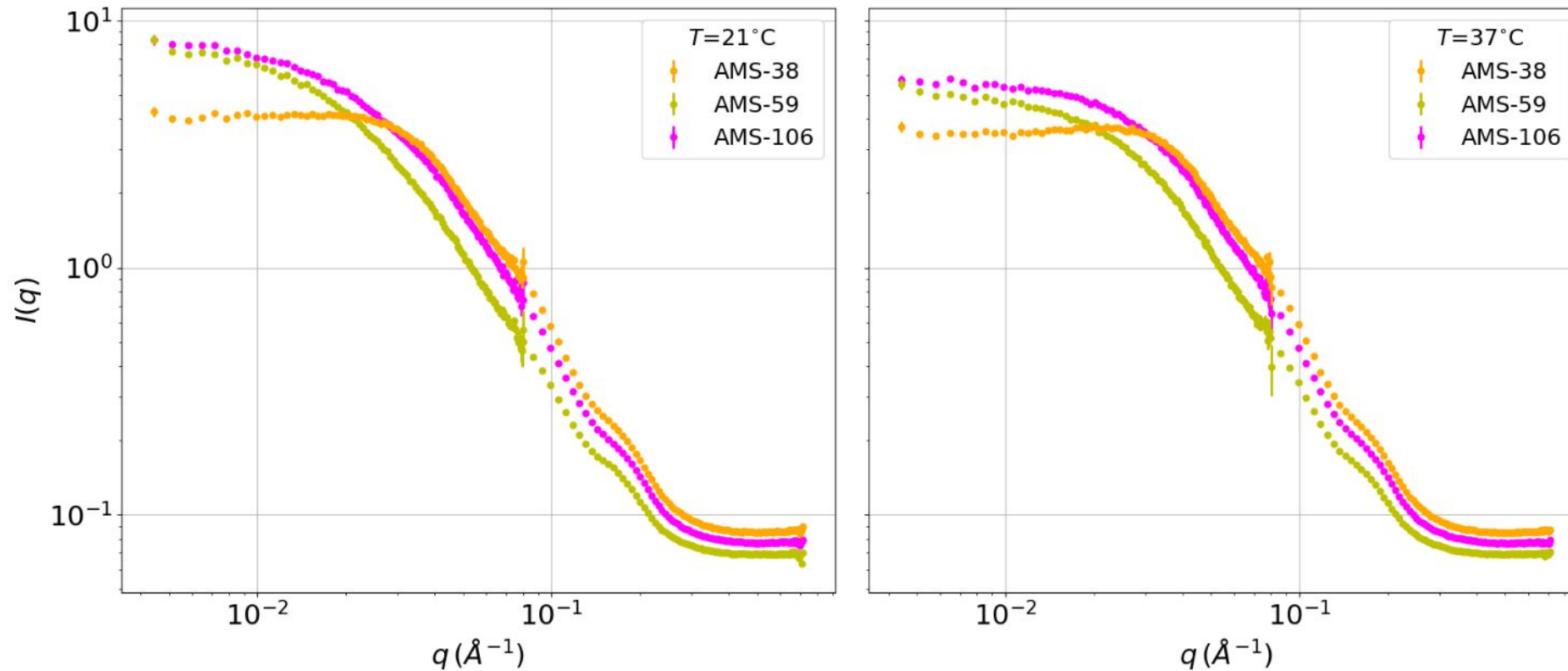
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4. First results: SANS

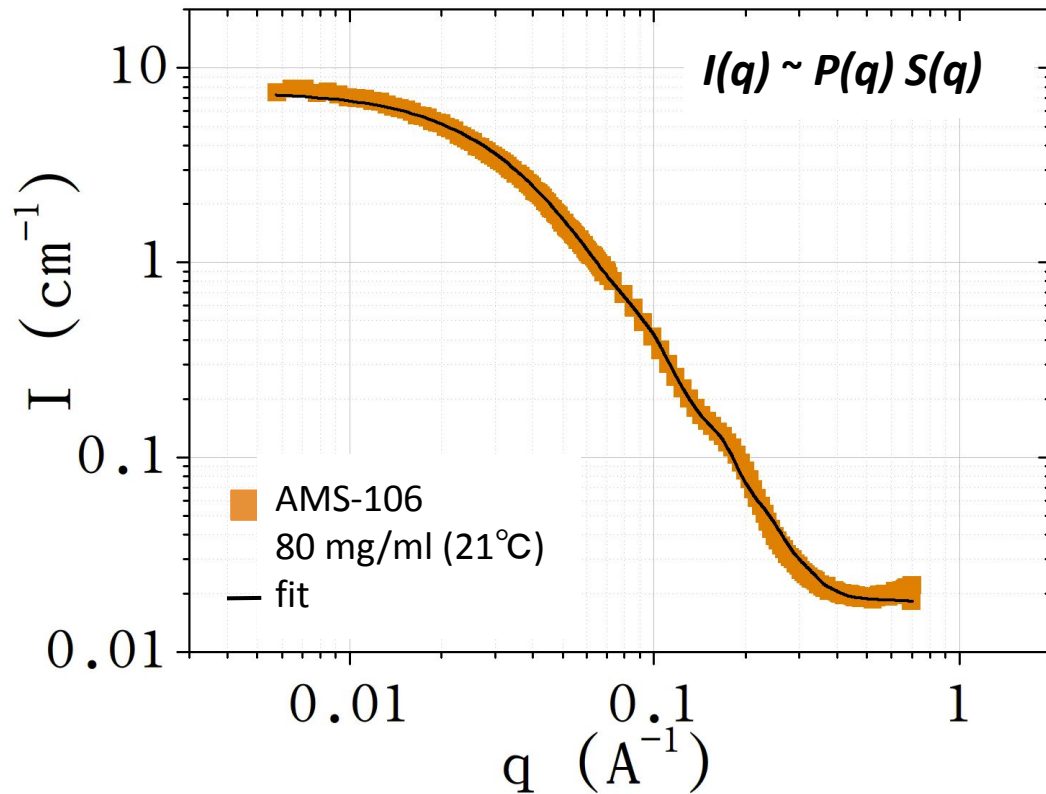
SANS curves, mAb-type dependence

Subset of mAbs: AMS-38, AMS-59, AMS-106; $c_\rho = 80$ mg/ml, buffered in 20mM His-HCl in D_2O



4. First results: SANS

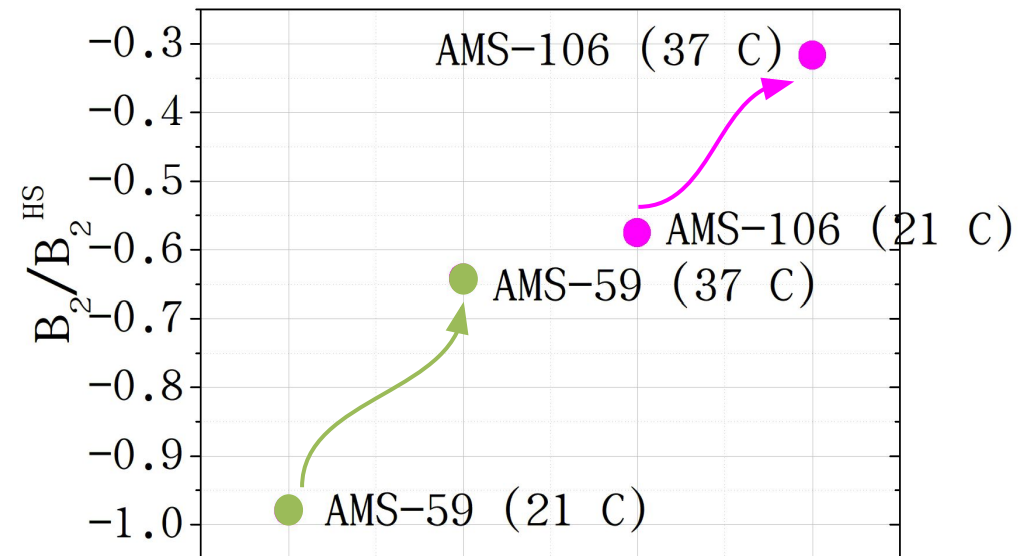
Fit: ellipsoid form factor $P(q)$, sticky hard sphere (SHS) structure factor $S(q)$ ^(1,2)



Calculation of 2nd virial coefficient B_2

$$B_2 = 2\pi \int_0^\infty dr r^2 [1 - e^{\frac{-u(r)}{k_B T}}]$$

→ attraction decreases with increasing temperature

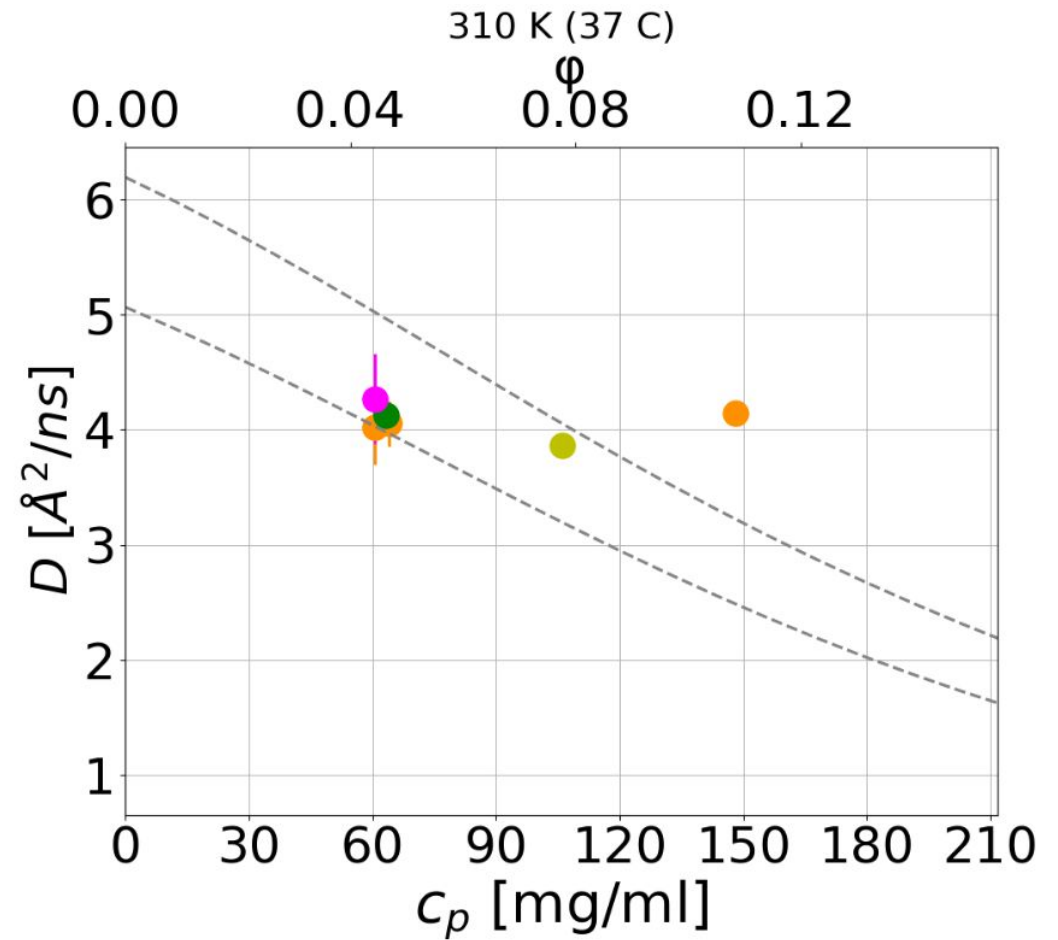
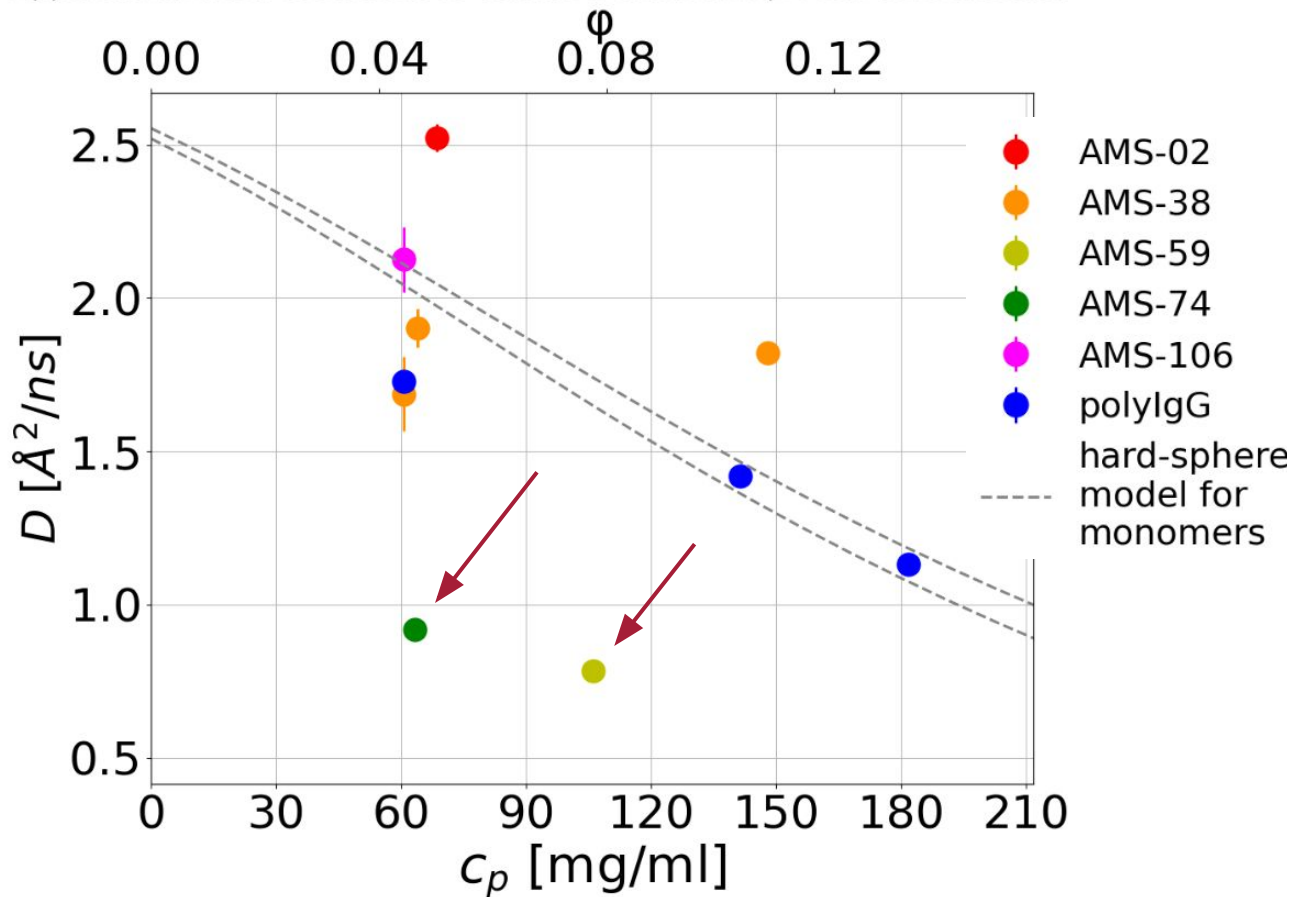


(1) N. Kern, D. Frenkel, J. Chem. Phys. 118 (2003)

(2) G. A. Vliegenthart, H. N. W. Lekkerkerker, J. Chem. Phys. 112 (2000)

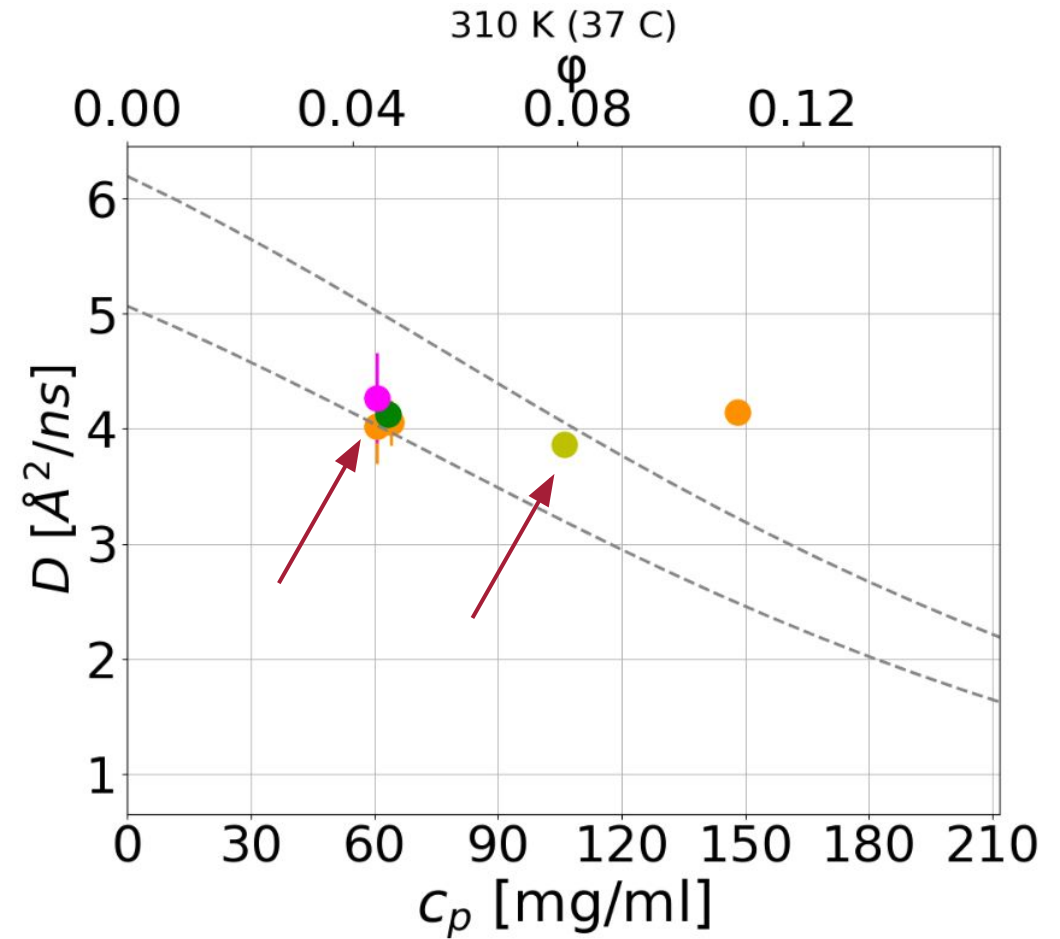
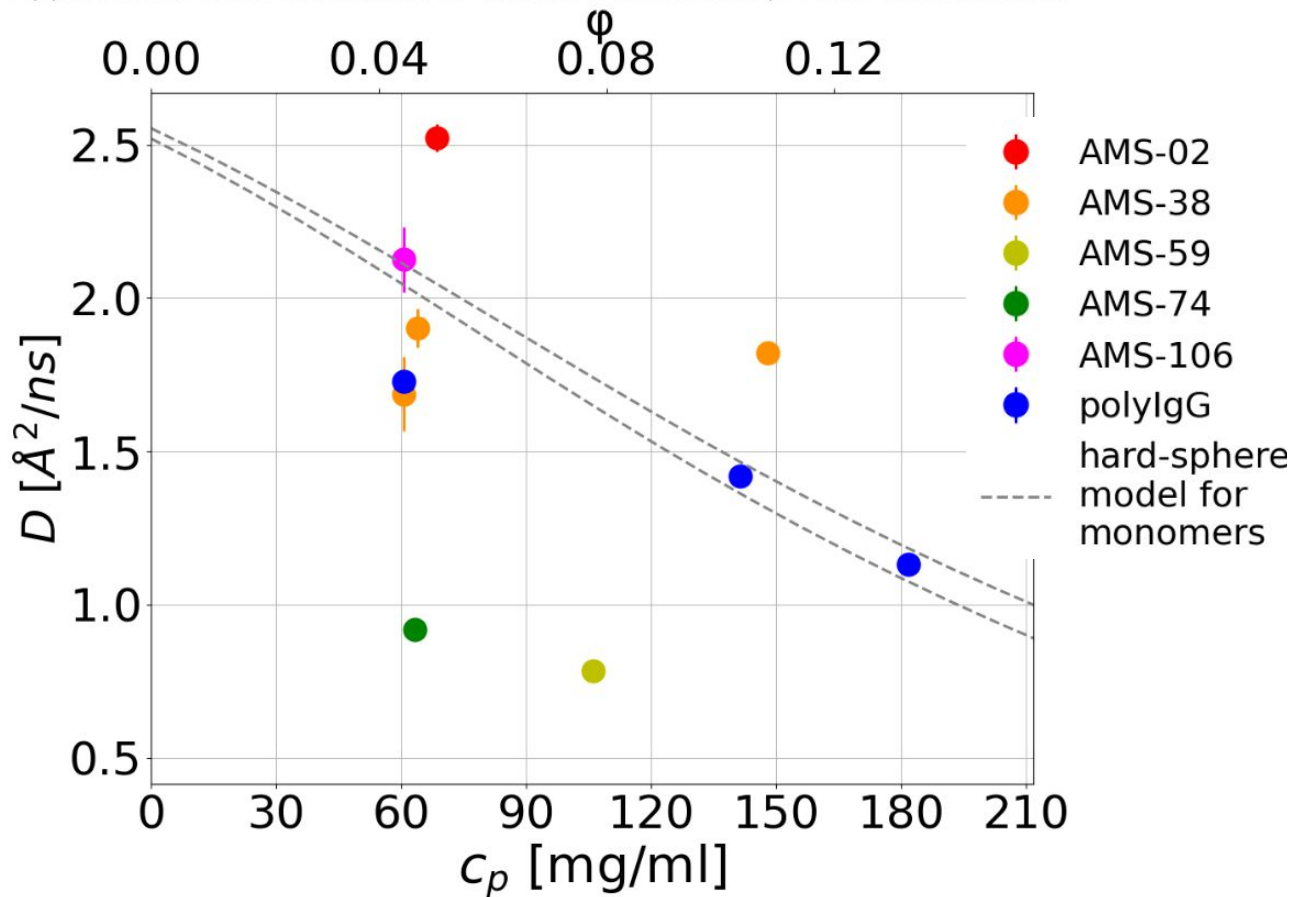
4. First results: QENS

Center of mass diffusion (280K and 310K)



4. First results: QENS

Center of mass diffusion (280K and 310K)



5. Conclusions and future developments

Conclusions

- Different rheology behavior of the mAb variants
- Most of them form clusters at 280K which may dissociate at higher T
- MD simulations indicate correlation between viscosity and surface charge and PPI parameters

Future developments

Analysis of data from:

- QENS @BATS (accurate info on internal dynamics)
- QENS @IN16B & SANS @D11, ILL (mAbs + trehalose)

Upcoming SAXS beamtime @ID02, ESRF: studies of mAb concentration, temperature and additives

