

Structure of soybean oleosomes studied by small angle neutron scattering

B.I. ZIELBAUER¹, M. GHEBREMEDHIN¹, R.K. HEENAN², A.J. JACKSON^{3,4},
S. MAURER^{1*}, L. PORCAR⁵, G. WASCHATKO¹⁺ and T.A. VILGIS¹

¹Max Planck Institute for Polymer Research, Mainz, Germany

²ISIS Facility, STFC Rutherford Appleton Laboratory, Didcot, OX11 0QX, United Kingdom

³European Spallation Source, Lund, 221 00, Sweden

⁴Lund University, Lund, 221 00, Sweden

⁵Institut-Laue-Langevin, Grenoble, 38042, France

* now at Nestlé HealthCare Nutrition GmbH, 67574 Osthofen, Germany

+ now at Cargill, 1800 Vilvoorde, Belgium



Science & Technology Facilities Council

ISIS



EUROPEAN
SPALLATION
SOURCE

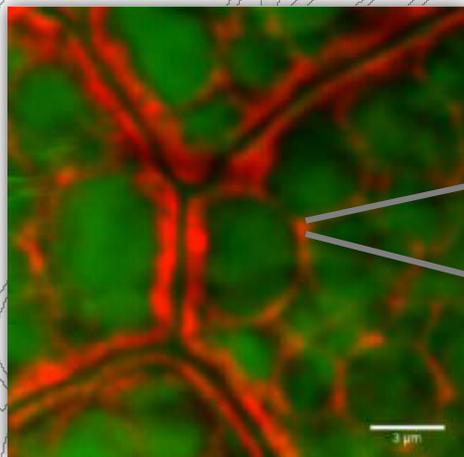


LUND
UNIVERSITY

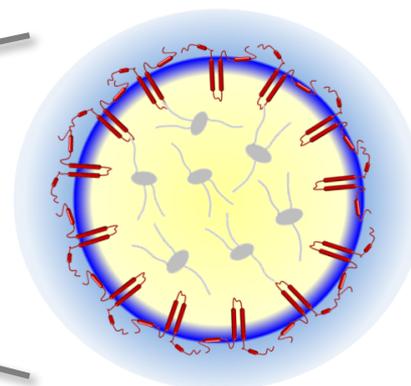
Oleosomes



Oleosomes: Intracellular oil storage particles in plants

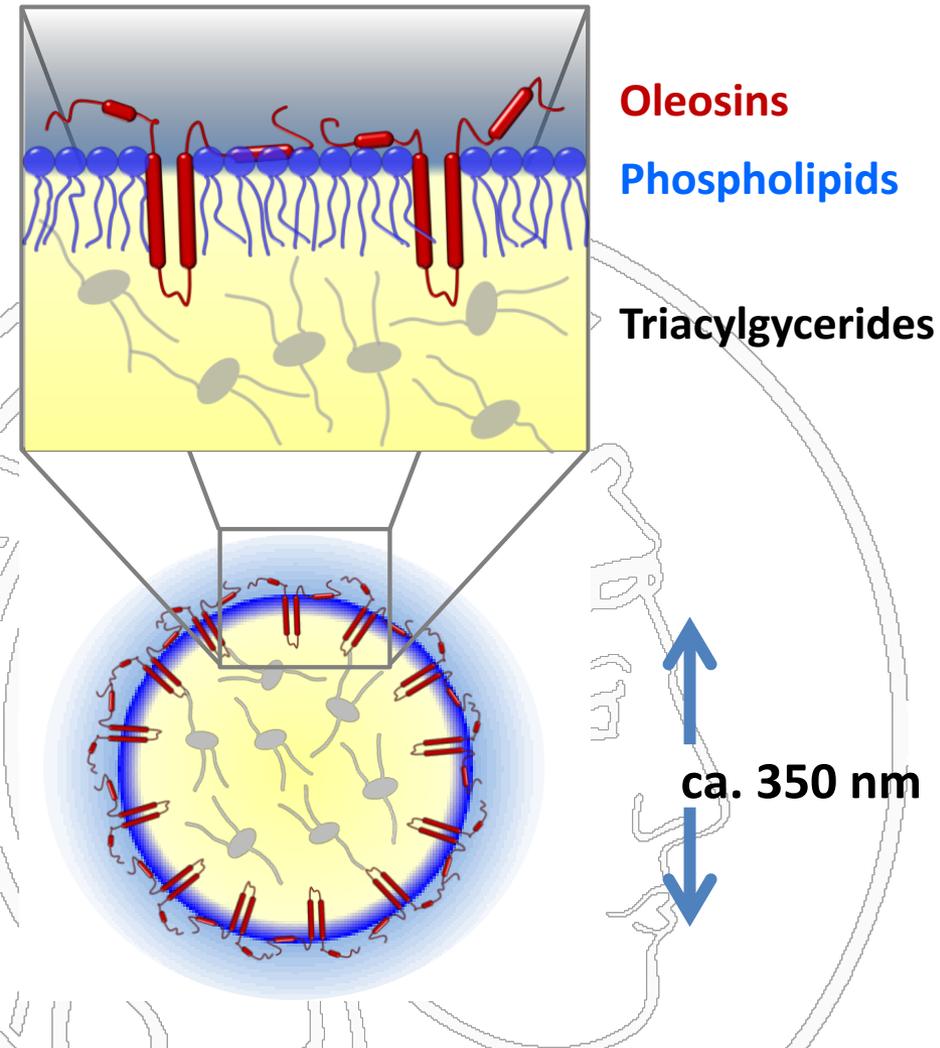


aqueous extraction

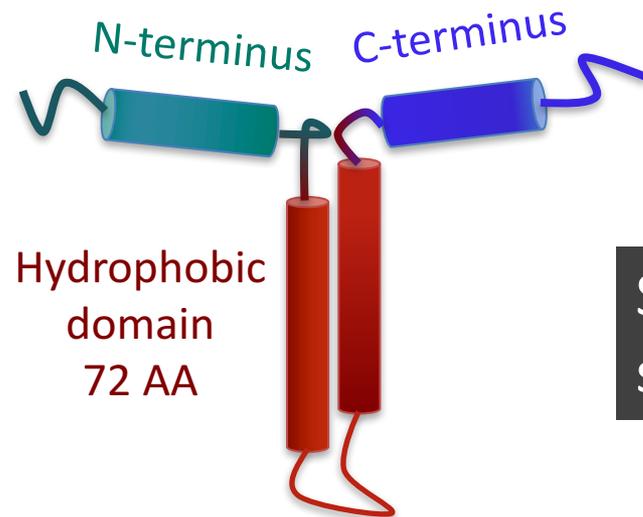


Natural emulsion of highly stabilised oil droplets

Oleosomes



Oleosin



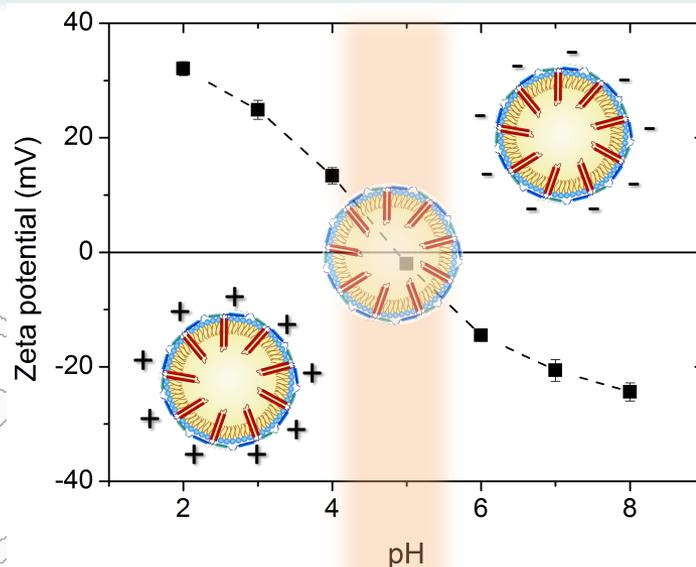
Secondary structure?

- ❖ Crucial for stability
- ❖ Determines oleosome size
- ❖ Acts as lipase acceptor

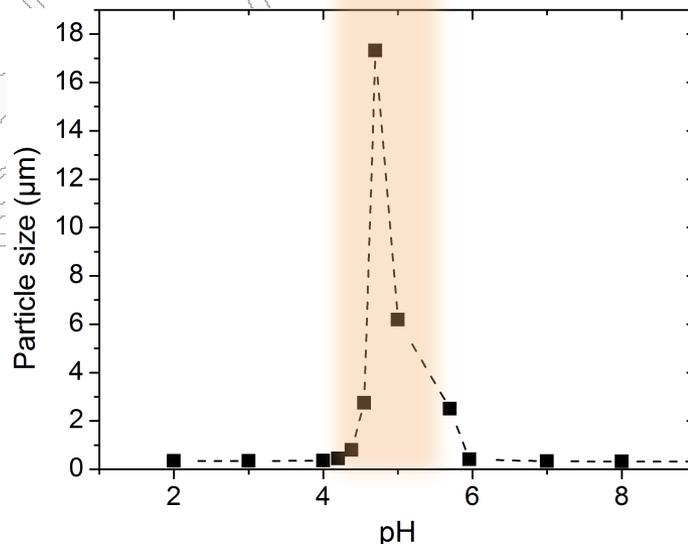
Importance of oleosin



Zeta potential



Particle size



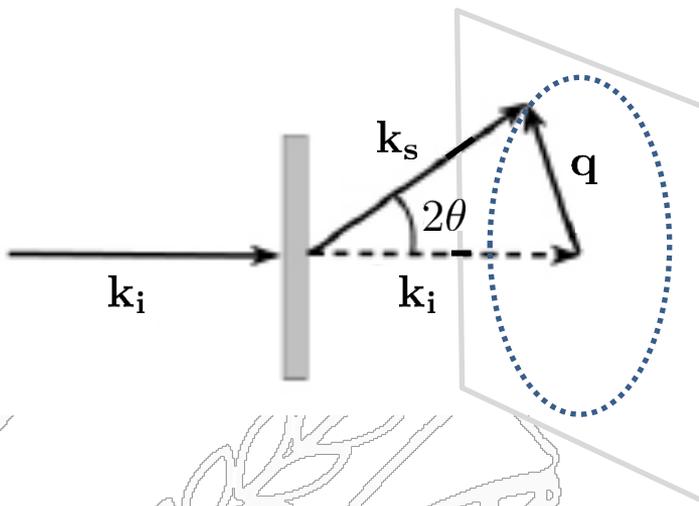
Maurer et al., *J. Phys. Chem. B* (2013) 117



- ❖ Alternative method for size determination
- ❖ Examine oleosin in its native environment – the oil/ water interface
- ❖ Detect early changes in interfacial layer upon environmental changes (e.g. temperature)

Scattering experiment

- Experiments performed at ISIS (Didcot, UK) and ILL (Grenoble, France)



$$\mathbf{q} = \mathbf{k}_s - \mathbf{k}_i$$

$$|\mathbf{q}| = \frac{4\pi}{\lambda} \sin \theta$$

Scattered intensity $I(q) = N(\Delta\bar{\rho}V)^2 P(q)S(q)$

molecules/volume

contrast

form factor:
shape

structure factor:
interactions

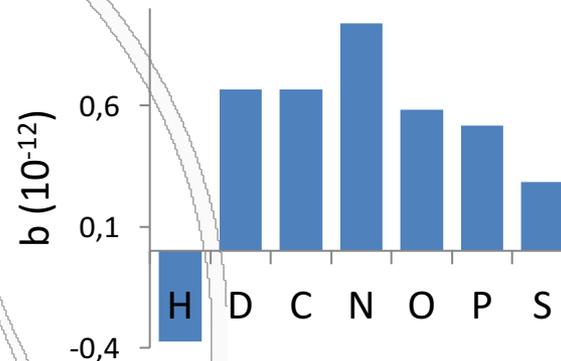
$$\Delta\bar{\rho} = \bar{\rho}_{particle} - \bar{\rho}_{solvent}$$

Neutron scattering

Neutrons interact with nuclei → scattering depends on isotope

Scattering length b (10^{-12} cm):

H	D	C	N	O	P	S
-0.3742	0.6671	0.6651	0.940	0.5804	0.517	0.2847

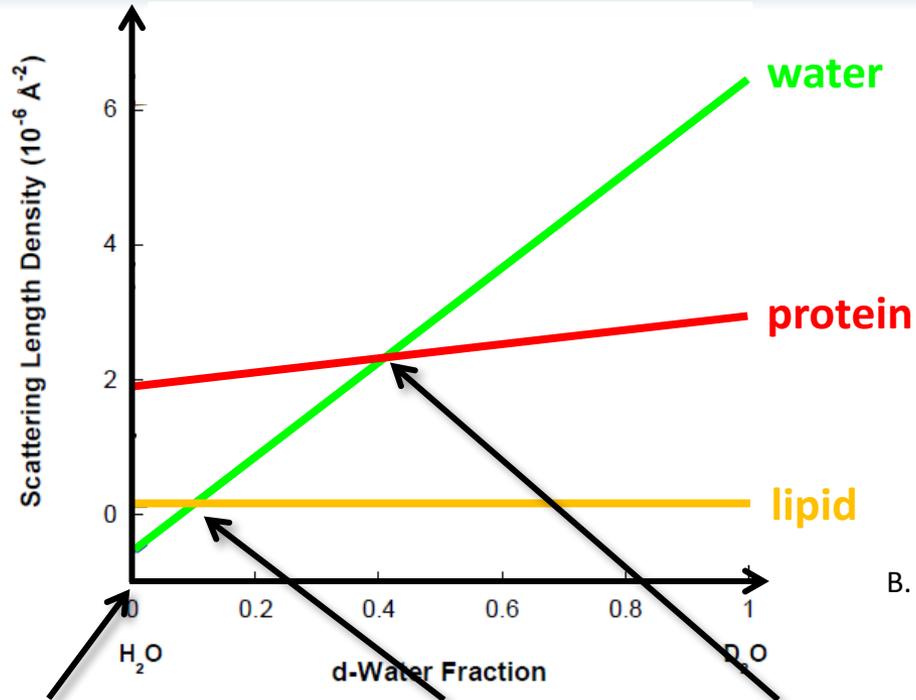


Scattering length density (SLD) ρ :

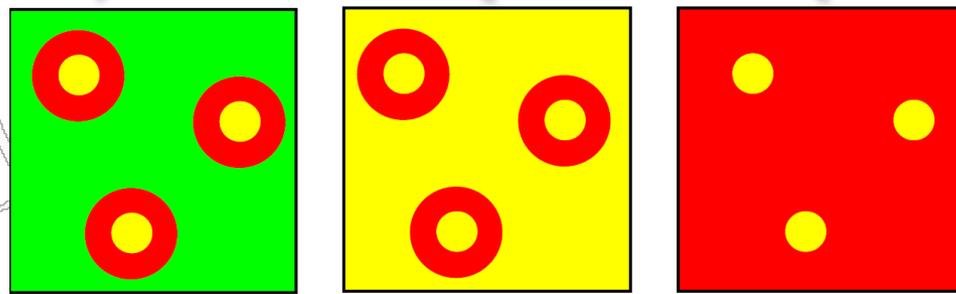
$$\rho = \frac{\sum_{i=1}^N b_i}{V}$$

Contrast variation

Biological systems



B. Hammouda (2008)



Natural contrast

$\rho_{\text{solv}} = \rho_{\text{core}}$
(shell visible)

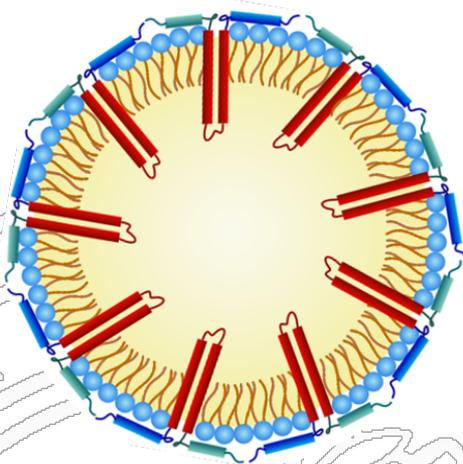
$\rho_{\text{solv}} = \rho_{\text{shell}}$
(core visible)

A. Jackson (2008)

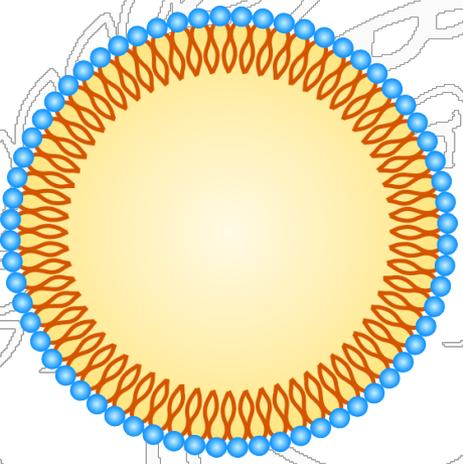
Test contrast variation



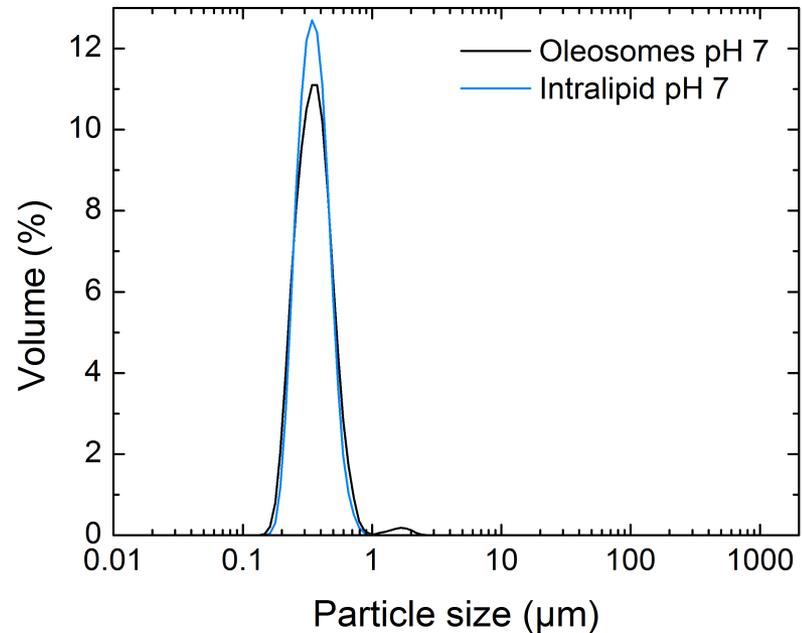
Oleosome



Intralipid



Size comparison

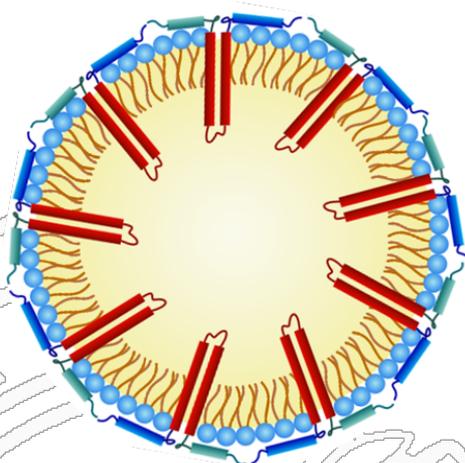


Intralipid:

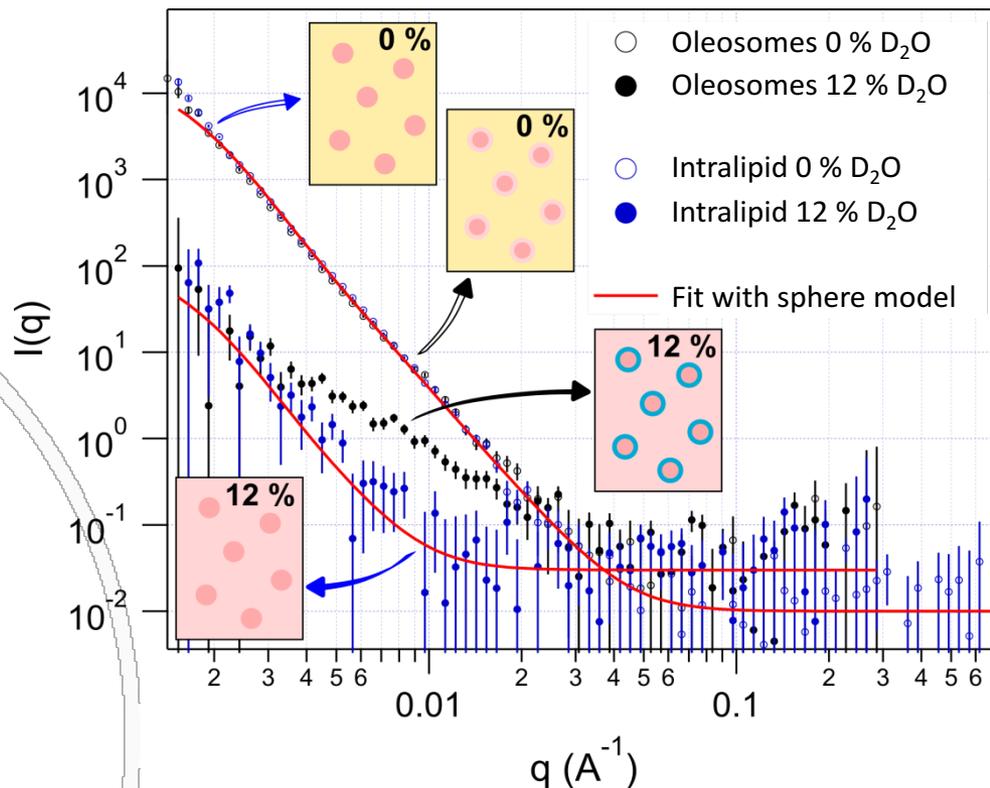
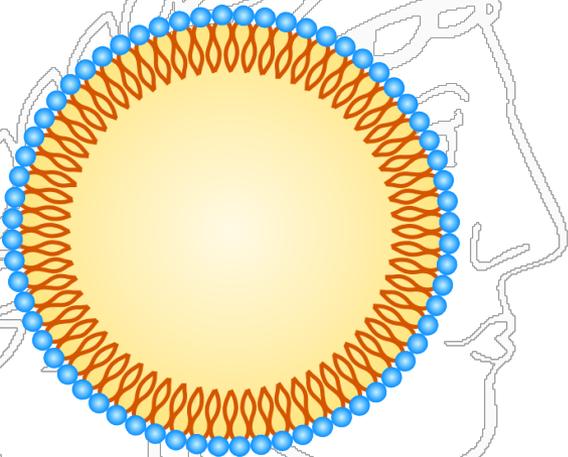
comparable system without protein

Test contrast variation

Oleosome



Intralipid

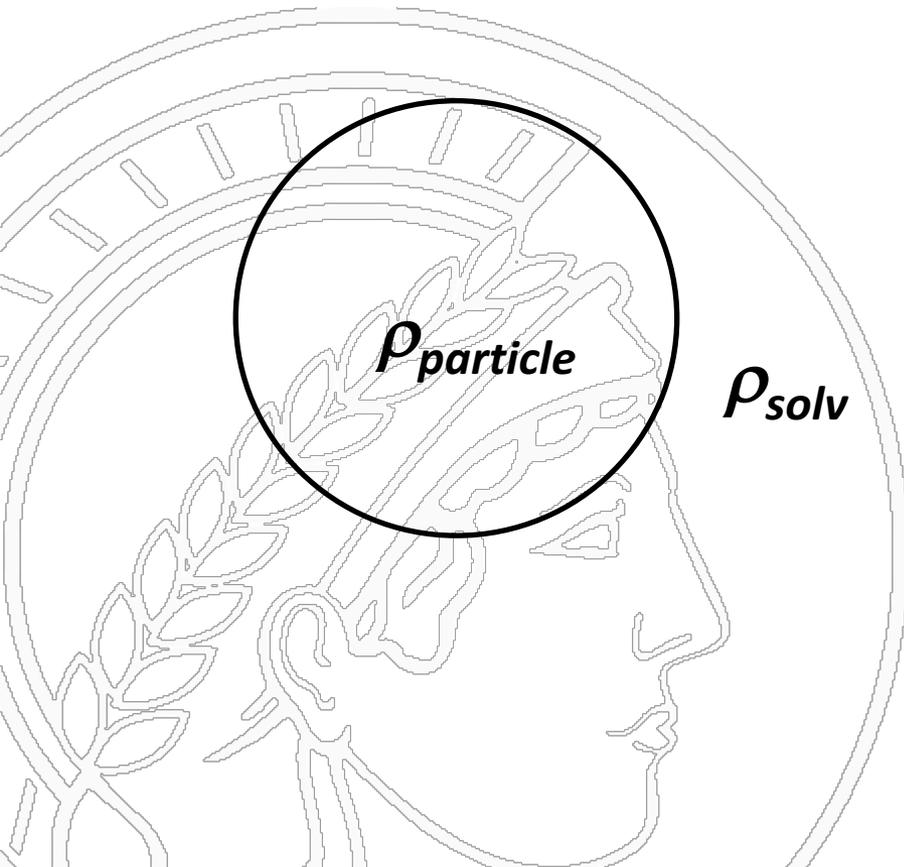


Oleosomes in 12% D₂O (oil core match) can not be fitted with sphere model
→ **signal of protein shell visible**

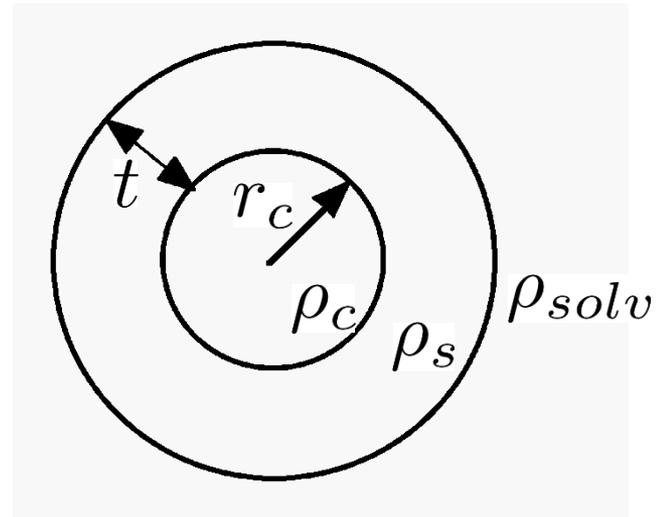
Geometry evaluation



sphere



core-shell particle

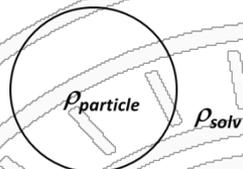


Fitting



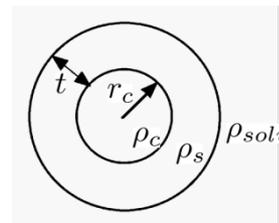
IGOR Pro + NIST SANS analysis macro ("*Reduction and Analysis of SANS and USANS Data Using IGOR Pro*") S. R. Kline, J. Appl. Cryst. 39 (2006) 895900

Sphere model



- polydisperse spheres

Core-shell model

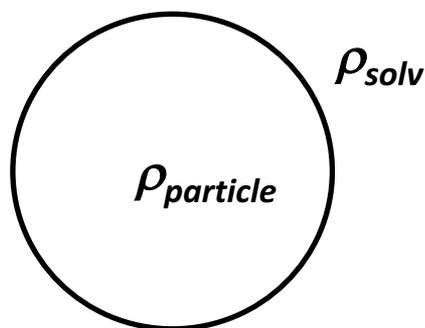
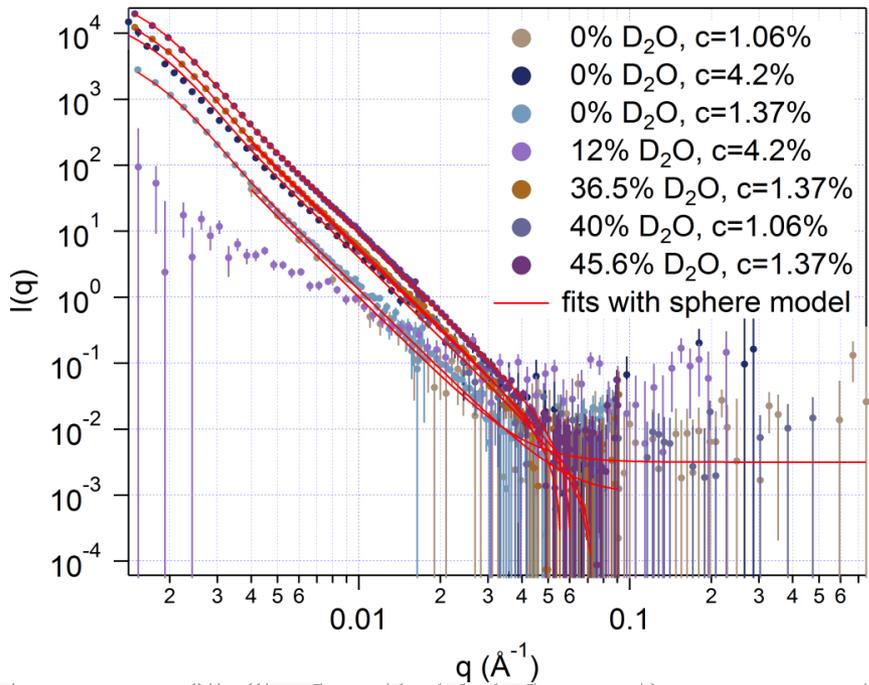


- polydisperse core
- constant shell thickness

Results from fits

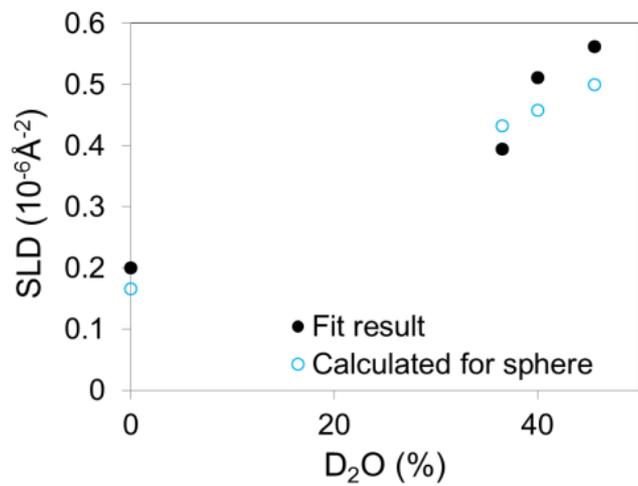
	Sphere model	Core-shell model
sizes	sphere radius	core radius, shell thickness
polydispersity	$p = \sigma/R$	
SLDs	$\rho_{particle}$	$\rho_{core}, \rho_{shell}$

Sphere model



Component	% (w/w)
Protein	4.65
Phospholipid	2.95
Solvent	10.5

- **Average SLD** based on composition
- **contrast dependent SLD** (solvent and hydrophilic protein domains)

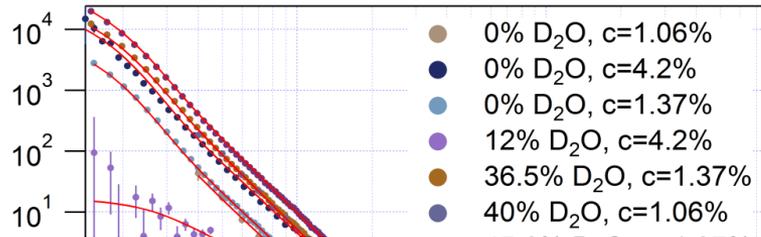


Fit results

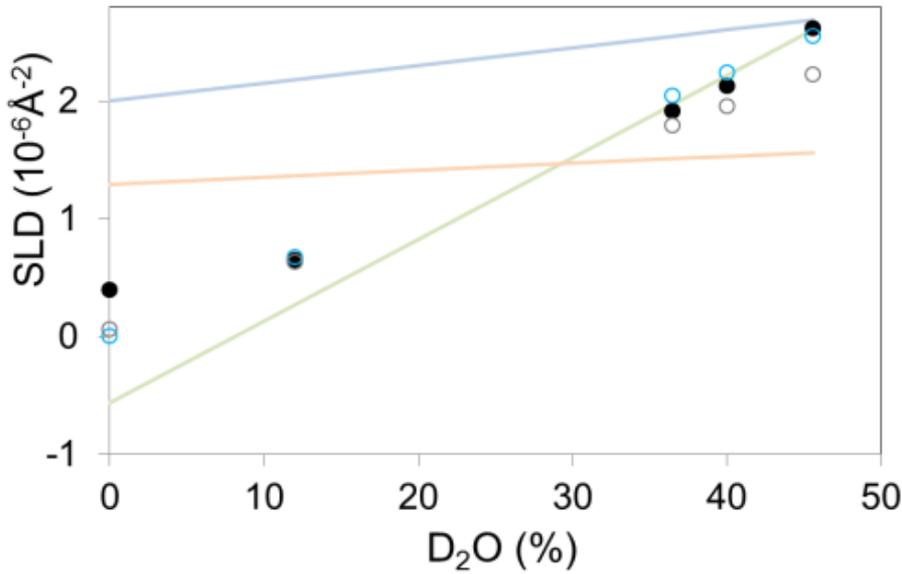
- $R = 1771 \text{ \AA}$
- $p = 0.395$

→ start values for core-shell fit

Core-shell model

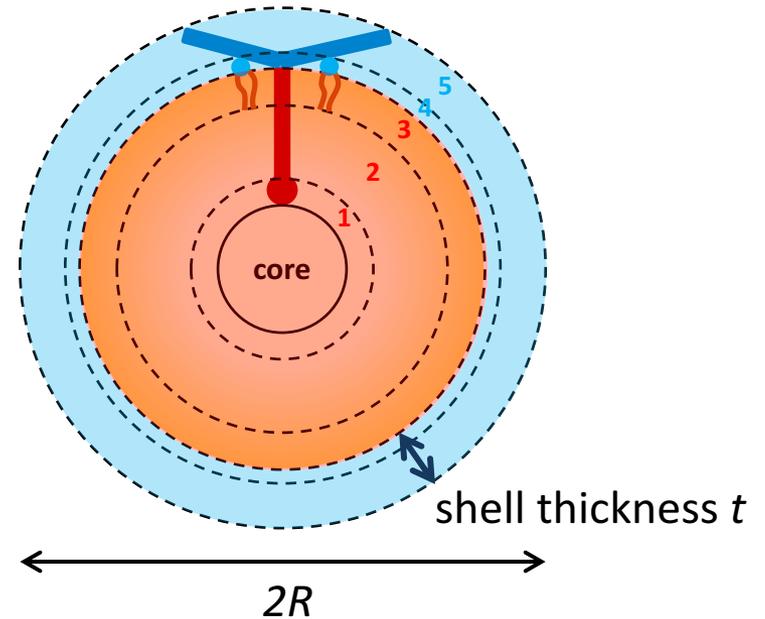


- Fit result
- SLD Solvent
- SLD PL head
- calculated for $d = d4 + d5$
- SLD Protein
- calculated for $d = d3 + d4 + d5$



Fit results

- $R = 1848 \text{ \AA}$
- $p = 0.396$
- $t = 91 \text{ \AA} \rightarrow 6.6 - 8.3 \text{ nm protein shell}$

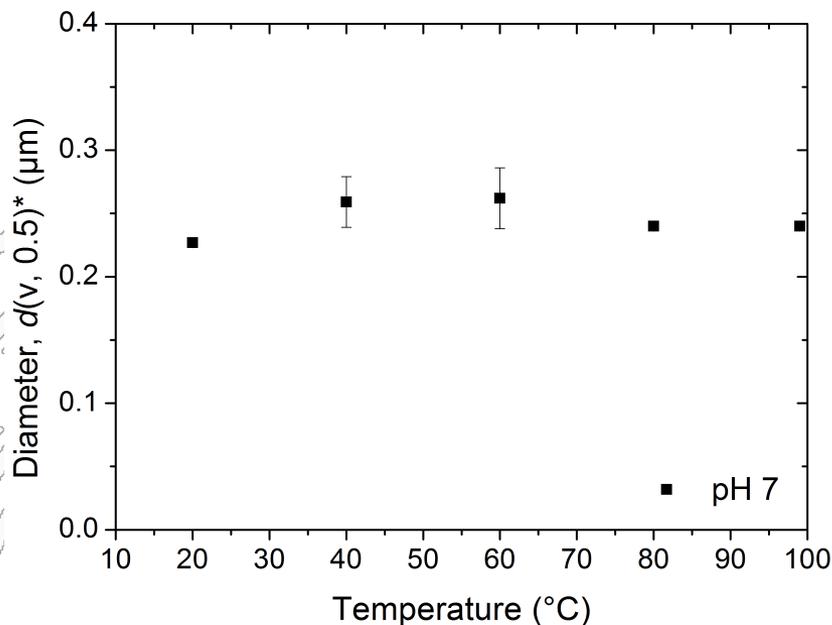


- ❖ theoretical shells
- ❖ calculate **SLDs for individual shells**

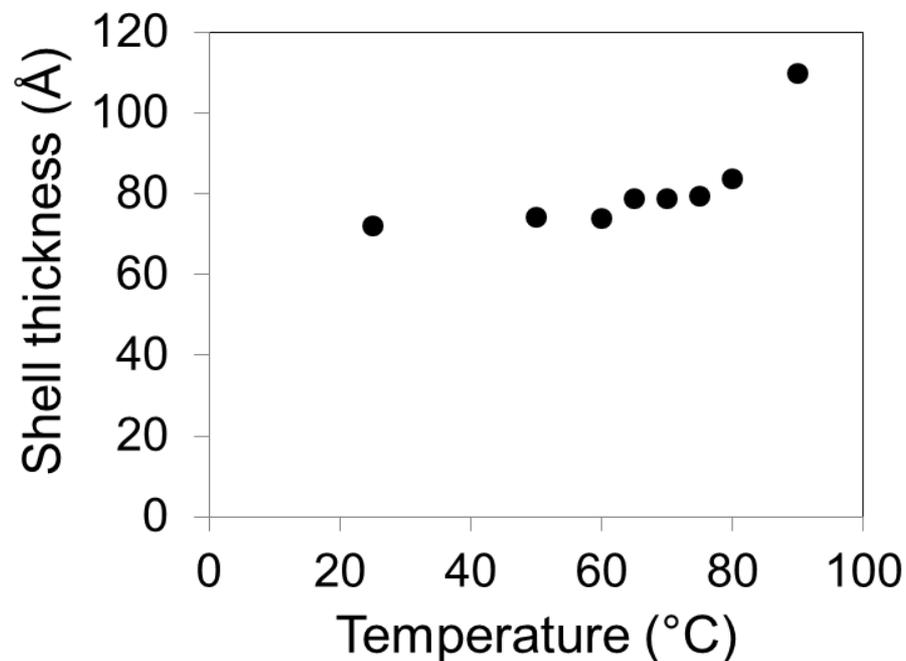
Temperature stability



Laser diffraction



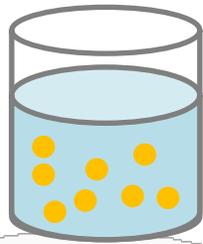
SANS



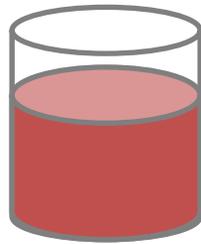
No significant change in shell thickness up to 80 $^{\circ}\text{C}$

→ high temperature stability

Encapsulation

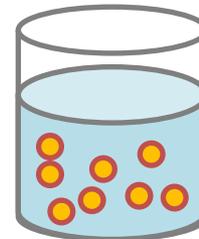


Oleosomes
7% w/w, pH 7

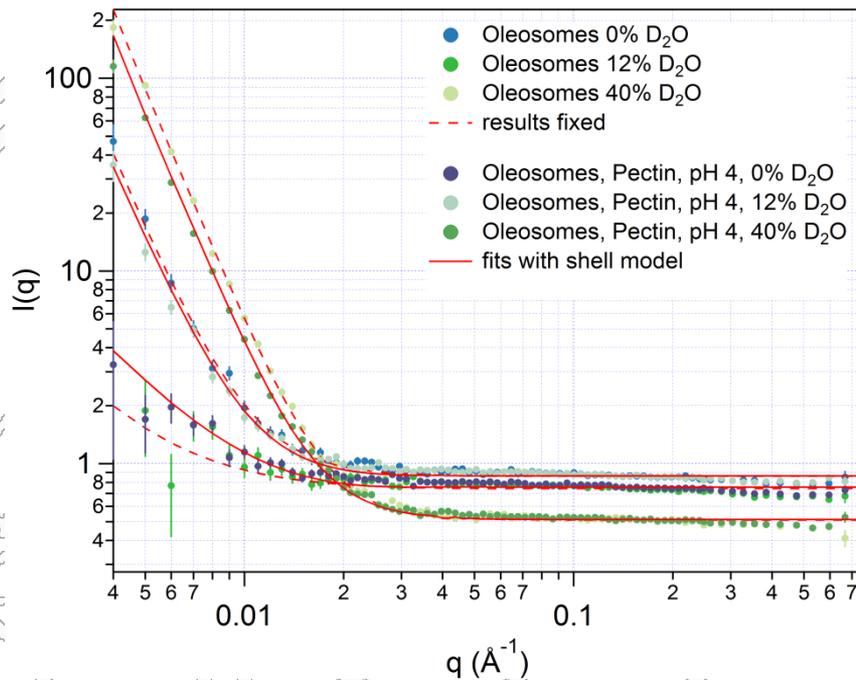


Pectin
0.5% w/w, pH 7

+ HCl
pH 4



Oleosomes
encapsulated
with pectin



Shell thickness for encapsulated
oleosomes 197 Å
→ 106 Å pectin layer

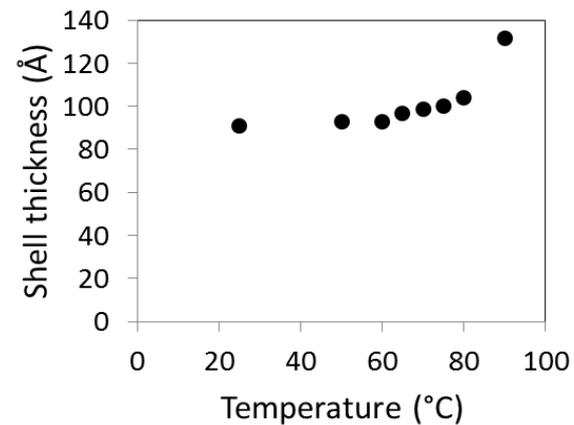
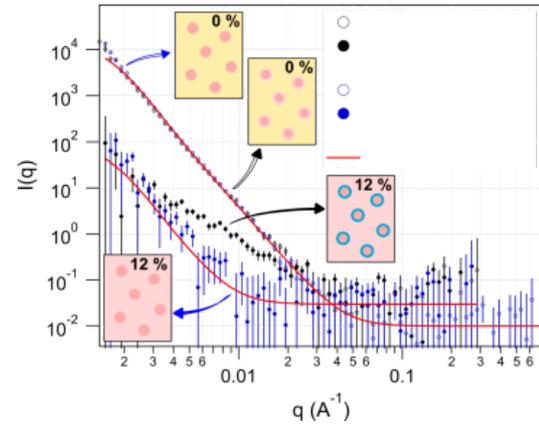
Conclusions

SANS can be used to study

❖ oleosomes' native interface

❖ temperature stability

❖ encapsulation



Thank you!



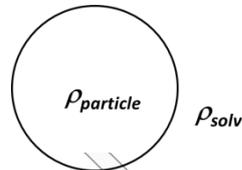


Fitting

IGOR Pro + NIST SANS analysis macro ("*Reduction and Analysis of SANS and USANS Data Using IGOR Pro*") S. R. Kline, J. Appl. Cryst. 39 (2006) 895900

a) Sphere model

- polydisperse spheres



Schulz distribution

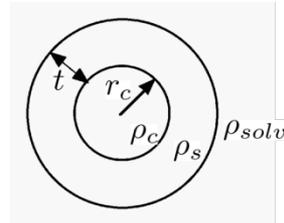
$$f(R) = (z+1)^{z+1} x^z \frac{\exp[-(z+1)x]}{R_{avg} \Gamma(z+1)}$$

Guinier radius:

$$R_g = \sqrt{\frac{3\langle R^8 \rangle}{5\langle R^6 \rangle}} = R_{avg} \sqrt{\frac{3(z+8)(z+7)}{5(z+1)^2}}$$

b) Core-shell model

- polydisperse core
- constant shell thickness



where $x = R/R_{avg}$; $z = 1/p^2 - 1$

with polydispersity $p = \sigma/R_{avg}$

and variance σ^2